BMJ Open *ETS-related gene (ERG)* expression as a predictor of oncological outcomes in patients with high-grade prostate cancer treated with primary androgen deprivation therapy: a cohort study

Mark Rezk,^{1,2} Ashish Chandra,³ Daniel Addis,³ Henrik Møller,⁴ Mina Youssef,^{5,6} Prokar Dasgupta,¹ Hide Yamamoto^{7,8}

To cite: Rezk M, Chandra A, Addis D, et al. *ETS-related gene (ERG)* expression as a predictor of oncological outcomes in patients with high-grade prostate cancer treated with primary androgen deprivation therapy: a cohort study. *BMJ Open* 2019;9:e025161. doi:10.1136/bmjopen-2018-025161

► Prepublication history and additional material for this paper are available online. To view these files, please visit the journal online (<http://dx.doi.org/10.1136/bmjopen-2018-025161>).

Received 2 July 2018
Revised 12 October 2018
Accepted 3 January 2019

© Author(s) (or their employer(s)) 2019. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.

For numbered affiliations see end of article.

Correspondence to

Dr Mark Rezk; mrezk@nhs.net

ABSTRACT

Objectives To determine whether *ETS-related gene (ERG)* expression can be used as a biomarker to predict biochemical recurrence and prostate cancer-specific death in patients with high Gleason grade prostate cancer treated with androgen deprivation therapy (ADT) as monotherapy.

Methods A multicentre retrospective cohort study identifying 149 patients treated with primary ADT for metastatic or non-metastatic prostate cancer with Gleason score 8–10 between 1999 and 2006. Patients planned for adjuvant radiotherapy at diagnosis were excluded. Age at diagnosis, ethnicity, prostate-specific antigen and Charlson-comorbidity score were recorded. Prostatic tissue acquired at biopsy or transurethral resection surgery was assessed for immunohistochemical expression of *ERG*. Failure of ADT defined as prostate specific antigen nadir +2. Vital status and death certification data determined using the UK National Cancer Registry. Primary outcome measures were overall survival (OS) and prostate cancer specific survival (CSS). Secondary outcome was biochemical recurrence-free survival (BRFS).

Results The median OS of our cohort was 60.2 months (CI 52.0 to 68.3). *ERG* expression observed in 51/149 cases (34%). Multivariate Cox proportional hazards analysis showed no significant association between *ERG* expression and OS ($p=0.41$), CSS ($p=0.92$) and BRFS ($p=0.31$). Cox regression analysis showed Gleason score ($p=0.003$) and metastatic status ($p<1\times 10^{-5}$) to be the only significant predictors of prostate CSS.

Conclusions No significant association was found between *ERG* status and any of our outcome measures. Despite a limited sample size, our results suggest that *ERG* does not appear to be a useful biomarker in predicting response to ADT in patients with high risk prostate cancer.

INTRODUCTION

The development of castration resistance is a major clinical hurdle in patients with advanced prostate cancer and is taken as a marker of impending mortality. Early identification of patients who develop castrate resistant prostate cancer can be clinically useful in enabling early

Strengths and limitations of this study

- This observational study consists of a large cohort of solely high-risk cancers treated with initial androgen deprivation therapy (ADT) as monotherapy with subsequent vital status determination through a UK national death certification registry.
- The association between *ETS-related gene (ERG)* expression and oncological survival is explored for the first time in patients on ADT as monotherapy.
- Our study population is of limited sample size. Accuracy of results may be reduced from lack of covariates gained through retrospective data collection.
- Determination of *ERG* status is limited to immunohistochemical detection of the protein without classification of its mutation at a genomic level.

aggressive treatment and therefore in reducing cancer-related deaths.

A recurrent gene fusion event involving the 3' end of *ERG* (*ETS-related gene*) to 5' *TMPRSS2* (*transmembrane protease, serine 2*)¹ is one of the most frequently occurring genetic aberrations in prostate cancer² but its prognostic value is still being explored.³ A meta-analysis evaluating the role of *TMPRSS2:ERG* fusion protein in patients undergoing radical prostatectomy found no association with biochemical recurrence or lethal disease.⁴

Given that *TMPRSS2:ERG* is androgen regulated,⁵ its association with oncological outcomes in patients treated with androgen deprivation therapy (ADT) is possible. *ERG* expression inversely correlates with the levels of androgen receptor protein in the cell and may exert a selective pressure for the development of a castrate-resistant state.⁶ Furthermore, androgen-regulated *ERG* expression

appears to persist following the development of castration resistance.⁷

In vivo validation of *ERG*'s metastatic influence has been controversial. Scheble VJ *et al* had shown a greater proportion of castration resistant metastatic prostate cancer driven by *ERG* negative tumours,⁸ while Perner S *et al* had observed a greater predilection to metastases in fusion positive foci.⁹

The aim of this study is to explore a possible association between *ERG* expression status and oncological outcomes in high grade and advanced prostate cancer patients treated by ADT as monotherapy. The primary end points are overall survival (OS) and prostate cancer specific survival (CSS). The secondary end point is biochemical recurrence-free survival (BRFS).

PATIENTS, MATERIALS AND METHODS

Data collection, study inclusion and exclusion criteria

Patients were identified from the pathology databases at two large neighbouring hospitals, Guy's and St Thomas' hospitals NHS Foundation Trust and King's College Hospital NHS Foundation Trust in London, UK, between January 1999 and August 2006. All patients treated with primary ADT were included in the study. Those were identified among patients with a total Gleason score of 8–10. For each patient, the initial assigned treatment was identified using electronic and paper records. Patients with both metastatic and non-metastatic disease were included in the study. Clinical data collected included the age at diagnosis, the assigned treatment at diagnosis, ethnicity (Caucasian, Afro-Caribbean, or other), Charlson comorbidity score,¹⁰ date of diagnosis, total modified International Society of Urologic Pathology 2005 Gleason Score, radiological evidence of metastasis at diagnosis, history of previous prostate cancer treatment, and serial prostate specific antigen (PSA) values (ng/mL). Patients were excluded from the study for any missing data, if they did not receive ADT or were planned to receive other adjuvant therapies such as radiotherapy. Data on unplanned adjuvant therapy following ADT were not collected due to incomplete follow-up data. The primary end points were OS and CSS. The secondary end point was BRFS.

Vital status and death certification data

Patient vital status data were retrieved from the National Cancer Registry in Public Health England.¹¹ Following institutional approval, unique patient National Health Service numbers were linked to vital status, dates of death and ICD-10 codes on the immediate cause of death (cause 1a), other diseases or conditions leading to 1a (causes 1b and 1c), underlying cause of death and other significant conditions not directly related to death (cause 2).¹² A prostate cancer death was defined as any death stating 'Prostate Cancer' in any of causes 1a, 1b, 1c or an underlying cause. Biochemical recurrence was defined as an increase of more than 2 ng/mL from the PSA nadir

value with censoring on the date when PSA rose more than 2 ng/mL above nadir.¹³

Prostate cancer sample collection, tissue processing and immunohistochemical staining

Prior to retrieval of archived prostate tissue samples, available H&E-stained slides were examined by two consultant histopathologists to select one tissue block for each patient based on the largest cancer volume. Specimen numbers were used to retrieve the corresponding paraffin-embedded blocks from the archives, and 3 µm sections were cut from each block using the Rotary Microtome HM 32S. Immunohistochemistry was performed in batches using the Ventana BenchMark ULTRA IHC/ISH automated stainer (Ventana Medical Systems). Deparaffinisation of the sections was carried by warming up the slides at 72°C in Ventana EZ Prep solution. Endogenous peroxidase activity was blocked using the Ventana inhibition kit and antigen retrieval was carried out by incubating the slides in Cell Conditioning solution-1 and subsequently heating at 100°C for 8 min, then 100 µL of Anti-ERG (EPR3864) Rabbit Monoclonal Primary Antibody was applied on each slide for 32 min. Visualisation was performed using anti-rabbit horseradish peroxidase (HRP)-labelled secondary antibody and 3,3'-diaminobenzidine tetrahydrochloride (DAB) chromogen (Roche/Ventana Ultra View DAB kit). The slides were washed and counterstained with Ventana Haematoxylin and Ventana Bluing Solution.

The IHC nuclear reactivity for *ERG* protein expression in the vascular endothelial cells was used as positive internal controls.¹⁴ Tests were repeated when endothelial cells failed to stain with ERG antibody (see supplementary figure 1).

H-scoring

Semiquantitative IHC analysis of *ERG* expression was conducted by the H-scoring system.¹⁴ Percentages of prostate cancer cells with positive and negative nuclear *ERG* staining were assessed at high magnification for each sample by two consultant histopathologists. The H-score was calculated as: 3× percentage cells with strong *ERG* expression +2× percentage of cells with intermediate *ERG* expression +1× percentage of cells with weak *ERG* expression.¹⁵ The total H-score per sample therefore ranged from 0 to 300. H-scores were classified as negative (0–50), weakly positive (51–100), moderately positive (101–200) or strongly positive (201–300) (see online supplementary figure 2).

Validation of antibody clone against an alternative anti-ERG antibody

Alternative *ERG* staining was carried out on selected cancer tissue samples using an alternative monoclonal ERG antibody (clone 9FY, ab139431). The results are depicted on the photomicrographs shown in supplementary figure 3.

Statistical methods

OS and CSS were determined using the Kaplan-Meier method. Univariate analysis of survival was performed

Table 1 Clinical characteristics of the study population stratified by *ERG* expression status (p values obtained by χ^2 or *t-tests)

	ERG negative (n=98)	%	ERG positive (n=51)	%	P value
Mean age ($\pm$ SD), years	72.3 ($\pm$ 8.3)		75.5 ($\pm$ 8.6)		0.03*
Ethnicity					
Caucasian	51	52	37	73	0.04
Afro-Caribbean	41	42	11	22	
Other	6	6	3	6	
Gleason score					
8	22	52	13	25	0.88
9	71	42	36	71	
10	5	6	2	4	
PSA ($\pm$ SD), ng/mL	1378 ($\pm$ 10849)		283 ($\pm$ 1203)		0.48*
<10.00	4	4	4	8	0.38
10–19	17	18	7	14	
20–49	24	25	18	36	
50–99	13	13	10	20	
$\geq$ 100	39	40	11	22	
Metastasis					
No	60	61	30	59	0.78
Yes	38	39	21	41	
Charlson comorbidity					
0	43	44	25	49	0.05
1	30	31	6	12	
2	16	16	11	22	
$\geq$ 3	9	9	9	18	
Follow-up ($\pm$ SD), months	47.9 ($\pm$ 25.5)		43.7 ($\pm$ 24.9)		0.34
Deaths					
All causes	46	47	28	55	0.39
Prostate cancer specific	29	30	17	33	0.71

ERG, ETS-related gene; PSA, prostate-specific antigen.

using the log-rank method. Multivariable Cox proportional hazards analysis was used to assess OS, CSS and BRFS with adjustments for *ERG* expression, age, ethnicity, Gleason score, PSA, presence of metastasis at presentation and Charlson comorbidity score. Statistical analyses were conducted using SPSS V.22, Graphpad Prism V.5.0 and Microsoft Excel software.

Patient and public involvement

No patients and public persons were involved in the commencement of this research.

RESULTS

Cohort characteristics

A total of 527 patients with high Gleason score prostate cancer were diagnosed on biopsy, of which 169 patients were assigned to primary ADT as monotherapy. Exclusion

of patients was due to tissue samples being unavailable (n=4), lack of vital status data output from the National Cancer Registry (n=4) or one or more missing clinical parameters (n=12). Complete data were available for 149 patients which formed the study population.

Mean follow-up was 46.5 ($\pm$ 25.2) months. Fifty-nine patients (40%) had metastatic disease at presentation. The clinical characteristics of the cohort are shown in (table 1).

ERG expression was observed in 51 cases (34%), of which nearly all demonstrated strong *ERG* expression (92%) (figure 1), (intensity distribution of *ERG* staining shown in supplementary figure 4). No *ERG* expression was found in incidental benign acini within samples. *ERG* positivity was associated with older age, and Caucasian ethnicity (when compared with Afro-Caribbean and Other ethnic groups), but not Gleason score, initial PSA

Figure 1 H-score distribution of ERG positive cases. 47/51 (92%) had a strongly positive H-score. ERG, ETS-related gene .

level, or presence of metastatic disease at presentation (table 1).

National Cancer Registry-linked oncological survival outcomes following primary androgen deprivation therapy in metastatic and non-metastatic high Gleason-grade prostate cancer

The National Cancer Registry was used to determine the vital status and death certification details for each patient. Seventy-five patients (50%) had died during follow-up, of whom 55 had died as a result of prostate cancer. Median OS for the cohort was 60.2 months. OS, CSS and BRFS for the cohort are shown (figure 2).

Presence of metastatic disease at diagnosis significantly affected OS ($p=0.001$), CSS ($p<1\times 10^{-7}$) and BRFS ($p<1\times 10^{-6}$). Gleason score significantly affected OS ($p=0.004$) and CSS ($p=0.004$) but not BRFS ($p=0.72$). PSA at presentation only affected BRFS ($p=1\times 10^{-5}$). Those associations were calculated using log-rank analysis.

Association of ETS-related gene expression and oncological outcomes in high risk cases treated by primary androgen deprivation therapy

Log-rank analysis was first conducted to determine whether *ERG* expression predicted oncological outcomes in the high-risk cohort stratified by *ERG* expression status

(figure 3). No statistically significant association was observed between *ERG* expression and OS, CSS or BRFS.

Cox proportional hazards regression analysis was conducted to determine independent predictors of oncological outcomes. Mutual adjustments were made for *ERG* expression, age, ethnicity, Gleason score, PSA, presence of metastasis at presentation and Charlson comorbidity (table 2). The presence of metastatic disease was significantly associated with OS (HR 2.60, 95% CI 1.54 to 4.40), CSS (HR 4.51, 95% CI 2.36 to 8.60) and BRFS (HR 3.15, 95% CI 1.93 to 5.16). Total Gleason score was significantly associated with OS (Gleason 9; HR 2.33, 95% CI 1.2 to 4.53 and Gleason 10; HR 5.81, 95% CI 2.04 to 16.52, reference group Gleason 8) and CSS (Gleason 9; HR 2.56, 95% CI 1.13 to 5.83 and Gleason 10; HR 6.45, 95% CI 2.04 to 16.52, reference group Gleason 8) but not BRFS. Age was significantly associated with OS only. We found no statistically significant association between *ERG* expression and OS, CSS or BRFS. The results did not change when *ERG* expression status was replaced with the H-score (results not shown).

DISCUSSION

In this cohort, we examined the association of *ERG* expression with survival endpoints in patients treated by primary

Figure 2 Oncological outcomes of high-risk prostate cancer following primary androgen deprivation therapy. Significant associations shown in bold. BSPPositive, bone scan positive.

ADT. Following multivariate analysis, we found no association of *ERG* expression with OS, CSS or BRFS (table 2). Advances in planned adjuvant treatments for high-risk prostate cancer such as radiotherapy or chemotherapy confounds the assessment of biomarkers in patients receiving ADT in more recent cohorts. Linkage of clinical data was made with the National Cancer Registry which

provided an up-to-date vital status on all patients residing in England.

ERG is commonly described as an oncogene although its ubiquitous expression in endothelial and haematopoietic stem cells suggest an essential role in angiogenesis, endothelial cell function and haematopoiesis.^{16 17} Since the discovery of the *ERG* and androgen regulated

Figure 3 Kaplan-Meier survival curves stratified by *ERG* expression status for OS, CSS and BRFS.

Table 2 Multivariate Cox proportional hazards analysis of *ERG* expression with other known oncological outcome parameters

	P value	HR	95% CI		P value	HR	95% CI		P value	HR	95% CI	
			Lower	Upper			Lower	Upper			Lower	Upper
<i>ERG</i> expression	0.41	1.24	0.74	2.05	0.92	1.03	0.57	1.87	0.31	0.78	0.47	1.27
Age	0.02	1.04	1.01	1.07	0.19	1.02	0.99	1.06	0.82	1.00	0.97	1.03
Ethnicity												
Caucasian (ref)	0.75				0.98				0.44			
Afro-Caribbean	0.70	0.90	0.53	1.53	0.86	0.94	0.51	1.75	0.52	0.85	0.52	1.38
Other	0.57	1.35	0.47	3.86	0.93	0.94	0.22	4.10	0.24	0.49	0.15	1.59
Gleason score												
8 (ref)	0.003				0.01				0.79			
9	0.01	2.33	1.20	4.53	0.02	2.56	1.13	5.83	0.50	1.20	0.71	2.01
10	<0.001	5.81	2.04	16.52	<0.01	6.45	1.92	21.71	0.73	1.24	0.35	4.38
PSA	0.92	1.00	1.00	1.00	0.96	1.00	1.00	1.00	0.88	1.00	1.00	1.00
Metastasis	<0.001	2.60	1.54	4.40	<1×10⁻⁵	4.51	2.36	8.60	<1×10⁻⁵	3.15	1.93	5.16
Charlson comorbidity												
0 (ref)	0.15				0.48				0.83			
1	0.19	1.52	0.81	2.87	0.85	1.08	0.50	2.35	0.56	1.18	0.68	2.07
2	0.06	1.81	0.97	3.39	0.67	1.18	0.56	2.47	0.75	0.90	0.49	1.67
≥3	0.09	2.00	0.89	4.47	0.12	2.17	0.81	5.84	0.51	1.29	0.60	2.75

Reference groups are indicated for categorical variables.

BRFS, biochemical recurrence-free survival; CSS, cancerspecific survival; *ERG*, ETS related gene; OS, overall survival; PSA, prostate specific antigen.

TMPRSS2 genetic fusion in prostate cancer,¹ its role as a sensitive and prevalent marker for prostate cancer has been shown to be highly replicable.⁴ Recent whole genome sequencing studies have revealed it to be the most frequent genetic aberration in prostate cancer within the entire genome.² Its high prevalence among all grades of disease^{4 18–21} however, supports its significance to be a marker of cancer per se rather than a marker for prognosis. *ERG* overexpression in animal models produces prostate intraepithelial neoplasia but not invasive cancer, suggesting it to be an early event in the natural history of prostate cancer.²²

Androgen receptor is known to play a role in the development of castrate resistance in prostate cancer⁵ and its levels have been shown to correlate with *ERG* expression.⁶ To the best of our knowledge, only a few studies have looked into the possible association of primary ADT on *ERG* positive cancer with varying conclusions.^{23–25} Similar to the findings of our study, Berg *et al* suggest no association between *ERG* expression and the development of castrate resistance in patients treated with primary ADT.²⁴ Huang *et al* had shown that combined *ERG* and androgen receptor status was significant in its association with a worsened survival in prostate cancer.²³ However, sole expression of *ERG* had not conferred worsened survival outcomes in patients with prostate cancer. Graff *et al* suggest a protective benefit in managing *ERG* positive prostate cancer with ADT.²⁵

Our study is the largest cohort of solely high-risk cancers homogeneously treated with initial ADT as planned monotherapy with subsequent high-quality vital status determination through a national registry.

In organ confined prostate cancer, *ERG* expression and its association with clinical outcomes has been the subject of numerous studies with conflicting outcomes.^{3 26} A meta-analysis describing *ERG* fusion positive cancer and its associated outcomes in postprostatectomy showed *ERG* fusion events to be associated with a higher clinical stage at diagnosis of T3 over T2 with a risk ratio of 1.23, yet no association was found for cancer specific survival or disease recurrence.⁴

Although we used IHC to estimate *TMPRSS2:ERG* gene fusion status, studies have shown very high concordance between more accurate fluorescence in situ hybridization (FISH) techniques.^{27–30} The reproducibility of the technique was assessed using an additional antibody clone, as well as determining technical success for each sample using endothelial cell expression as internal controls (supplementary figures 1 and 3). The prevalence (34%) of *ERG* expression is in line with previous studies.⁴ The association between Caucasian ethnicity and *ERG* expression agrees with a previous study evaluating *TMPRSS2:ERG* fusion events.³¹ Higher age was significantly associated with *ERG* expression in our cohort ($p=0.03$), in contrast to other studies that showed a higher proportional expression in the younger men³² or no correlation at all.^{4 14} It is

possible that this is an association seen only in high grade prostate cancer cases.

Limitations

The retrospective nature of the study had resulted in a reduction in the collection of other covariates such as stage at diagnosis.⁴

Moreover, a subset of patients within the cohort received unplanned adjuvant therapy in addition to androgen deprivation monotherapy which may have influenced the OS. This was not assessed as a covariate due to the heterogeneous nature of the treatment and small patient numbers.

Patients who did not have a complete dataset were excluded from the study although this had represented a small proportion of patients (20/169). In addition, despite being the largest cohort of patients solely treated with primary ADT, the sample size remains small reducing the power of this study population.

Both quantitative³³ and qualitative differences in the *ERG* mutation have been implicated in prognostication of prostate cancer. Patients with cancer cells exhibiting an aberration consisting of both a duplication and deletion of the 5' end of *ERG* (known as 2+ 'Edel') were predisposed to a poorer disease specific survival.^{34 35} In this context, the use of IHC was a limitation as it cannot detect the genomic fusion quantitatively or qualitatively but only isolated expression of the *ERG* protein.^{7 27} It is important to note that the H score does not provide a quantitative measurement of the *ERG* aberration.²⁷ With this knowledge at hand, subclassifications of *ERG* mutations when assessing prognostic indicators is recommended in future clinical studies. IHC may be controversial as a detection method of the *TMPRSS2:ERG* fusion gene. Sung JY *et al* expressed caution to its use for its false positive rate³⁶ while Gsponer and colleagues identified a subgroup of *ERG* genetic alterations that are undetectable at a protein level.³⁷

CONCLUSION

While *ERG* expression is known to be strongly associated with oncogenesis, we show that *ERG* expression did not predict oncological survival in prostate cancer patients treated with ADT. Our findings are in line with other studies showing a lack of association between *ERG* expression and prostate cancer treatment outcomes.

Author affiliations

¹NIHR Biomedical Research Centre, Kings College London, London, UK

²Intensive Care Unit, Torbay and South Devon NHS Foundation Trust, Torquay, UK

³Department of Histopathology and Cytology, Guy's and St Thomas' NHS Foundation Trust, London, UK

⁴Department of Cancer Epidemiology & Populational Health, Kings College London, London, UK

⁵General Surgery Department, Norfolk and Norwich University Hospitals NHS Foundation Trust, Norwich, UK

⁶Surgical Oncology Department, National Cancer Institute, Cairo, Egypt

⁷Department of Urology, Guy's and St Thomas' NHS Foundation Trust, London, UK

⁸Department of Urology, Maidstone Hospital, Maidstone, UK

Contributors HY led the study design, analysed data and was involved in revision, drafting and approval of the final manuscript. MR was involved in sample and data collection, data analysis and drafting of the manuscript. AC and DA were both involved in analysis of the prostate cancer tissue. HM, MY and PD provided critical review of the study design and manuscript. All authors meet the ICMJE criteria for authorship.

Funding This research received no specific grant from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests None declared.

Patient consent for publication Not required.

Ethics approval Institutional approval was granted prior to the study.

Provenance and peer review Not commissioned; externally peer reviewed.

Data sharing statement Due to the confidential nature of our data which may identify individuals, even following anonymisation, we have chosen not to make our data publicly available.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>.

REFERENCES

- Tomlins SA, Rhodes DR, Perner S, *et al*. Recurrent fusion of *TMPRSS2* and *ETS* transcription factor genes in prostate cancer. *Science* 2005;310:644–8.
- Fraser M, Sabelnykova VY, Yamaguchi TN, *et al*. Genomic hallmarks of localized, non-indolent prostate cancer. *Nature* 2017;541:359–64.
- Sreenath TL, Dobi A, Petrovics G, *et al*. Oncogenic activation of *ERG*: A predominant mechanism in prostate cancer. *J Carcinog* 2011;10:37.
- Pettersson A, Graff RE, Bauer SR, *et al*. The *TMPRSS2:ERG* rearrangement, *ERG* expression, and prostate cancer outcomes: a cohort study and meta-analysis. *Cancer Epidemiol Biomarkers Prev* 2012;21:1497–509.
- Clinckemalie L, Spans L, Dubois V, *et al*. Androgen regulation of the *TMPRSS2* gene and the effect of a SNP in an androgen response element. *Mol Endocrinol* 2013;27:2028–40.
- Yu J, Yu J, Mani RS, *et al*. An integrated network of androgen receptor, polycomb, and *TMPRSS2-ERG* gene fusions in prostate cancer progression. *Cancer Cell* 2010;17:443–54.
- Attard G, Swennenhuis JF, Olmos D, *et al*. Characterization of *ERG*, *AR* and *PTEN* gene status in circulating tumor cells from patients with castration-resistant prostate cancer. *Cancer Res* 2009;69:2912–8.
- Scheble VJ, Scharf G, Braun M, *et al*. *ERG* rearrangement in local recurrences compared to distant metastases of castration-resistant prostate cancer. *Virchows Arch* 2012;461:157–62.
- Perner S, Svensson MA, Hossain RR, *et al*. *ERG* rearrangement metastasis patterns in locally advanced prostate cancer. *Urology* 2010;75:762–7.
- Hall WH, Ramachandran R, Narayan S, *et al*. An electronic application for rapidly calculating Charlson comorbidity score. *BMC Cancer* 2004;4:94.
- NCRAS. The National Cancer Registration and Analysis Service. 2017 <https://www.gov.uk/guidance/national-cancer-registration-and-analysis-service-ncras> (Cited 10 November 2015).
- Office of National Statistics. Guidance for doctors completing Medical Certificates of Cause of Death in England and Wales. 2010 http://www.gro.gov.uk/images/medcert_july_2010.pdf (Cited 15 Nov 2015).
- Roach M, Hanks G, Thames H, *et al*. Defining biochemical failure following radiotherapy with or without hormonal therapy in men with clinically localized prostate cancer: Recommendations of the RTOG-ASTRO Phoenix Consensus Conference. *Int J Radiat Oncol Biol Phys* 2006;65:965–74.
- Hoogland AM, Jenster G, van Weerden WM, *et al*. *ERG* immunohistochemistry is not predictive for PSA recurrence, local recurrence or overall survival after radical prostatectomy for prostate cancer. *Mod Pathol* 2012;25:471–9.
- Kraus JA, Dabbs DJ, Beriwal S, *et al*. Semi-quantitative immunohistochemical assay versus oncotype DX(®) qRT-PCR assay

- for estrogen and progesterone receptors: an independent quality assurance study. *Mod Pathol* 2012;25:869–76.
16. Wagner W, Ansorge A, Wirkner U, *et al.* Molecular evidence for stem cell function of the slow-dividing fraction among human hematopoietic progenitor cells by genome-wide analysis. *Blood* 2004;104:675–86.
 17. Birdsey GM, Dryden NH, Amsellem V, *et al.* Transcription factor Erg regulates angiogenesis and endothelial apoptosis through VE-cadherin. *Blood* 2008;111:3498–506.
 18. Mehra R, Tomlins SA, Shen R, *et al.* Comprehensive assessment of TMPRSS2 and ETS family gene aberrations in clinically localized prostate cancer. *Mod Pathol* 2007;20:538–44.
 19. Font-Tello A, Juanpere N, de Muga S, *et al.* Association of ERG and TMPRSS2-ERG with grade, stage, and prognosis of prostate cancer is dependent on their expression levels. *Prostate* 2015;75:1216–26.
 20. Bismar TA, Dolph M, Teng LH, *et al.* ERG protein expression reflects hormonal treatment response and is associated with Gleason score and prostate cancer specific mortality. *Eur J Cancer* 2012;48:538–46.
 21. Leinonen KA, Saramäki OR, Furusato B, *et al.* Loss of PTEN is associated with aggressive behavior in ERG-positive prostate cancer. *Cancer Epidemiol Biomarkers Prev* 2013;22:2333–44.
 22. Tomlins SA, Laxman B, Varambally S, *et al.* Role of the TMPRSS2-ERG gene fusion in prostate cancer. *Neoplasia* 2008;10:177–IN9.
 23. Huang KC, Alshalalfa M, Hegazy SA, *et al.* The prognostic significance of combined ERG and androgen receptor expression in patients with prostate cancer managed by androgen deprivation therapy. *Cancer Biol Ther* 2014;15:1120–8.
 24. Berg KD, Røder MA, Thomsen FB, *et al.* The predictive value of ERG protein expression for development of castration-resistant prostate cancer in hormone-naïve advanced prostate cancer treated with primary androgen deprivation therapy. *Prostate* 2015;75:1499–509.
 25. Graff RE, Pettersson A, Lis RT, *et al.* The TMPRSS2:ERG fusion and response to androgen deprivation therapy for prostate cancer. *Prostate* 2015;75:897–906.
 26. Adamo P, Ladomery MR. The oncogene ERG: a key factor in prostate cancer. *Oncogene* 2016;35:403–.
 27. Chaux A, Albadine R, Toubaji A, *et al.* Immunohistochemistry for ERG expression as a surrogate for TMPRSS2-ERG fusion detection in prostatic adenocarcinomas. *Am J Surg Pathol* 2011;35:1014–20.
 28. Schelling LA, Williamson SR, Zhang S, *et al.* Frequent TMPRSS2-ERG rearrangement in prostatic small cell carcinoma detected by fluorescence in situ hybridization: the superiority of fluorescence in situ hybridization over ERG immunohistochemistry. *Hum Pathol* 2013;44:2227–33.
 29. Falzarano SM, Zhou M, Carver P, *et al.* ERG gene rearrangement status in prostate cancer detected by immunohistochemistry. *Virchows Arch* 2011;459:441–7.
 30. Park K, Tomlins SA, Mudaliar KM, *et al.* Antibody-based detection of ERG rearrangement-positive prostate cancer. *Neoplasia* 2010;12:590–8.
 31. Magi-Galluzzi C, Tsusuki T, Elson P, *et al.* TMPRSS2-ERG gene fusion prevalence and class are significantly different in prostate cancer of Caucasian, African-American and Japanese patients. *Prostate* 2011;71:489–97.
 32. Schaefer G, Mosquera JM, Ramoner R, *et al.* Distinct ERG rearrangement prevalence in prostate cancer: higher frequency in young age and in low PSA prostate cancer. *Prostate Cancer Prostatic Dis* 2013;16:132–8.
 33. FitzGerald LM, Agalliu I, Johnson K, *et al.* Association of TMPRSS2-ERG gene fusion with clinical characteristics and outcomes: results from a population-based study of prostate cancer. *BMC Cancer* 2008;8:1.
 34. Attard G, Clark J, Ambrosine L, *et al.* Duplication of the fusion of TMPRSS2 to ERG sequences identifies fatal human prostate cancer. *Oncogene* 2008;27:253–63.
 35. Mehra R, Tomlins SA, Yu J, *et al.* Characterization of TMPRSS2-ETS gene aberrations in androgen-independent metastatic prostate cancer. *Cancer Res* 2008;68:3584–90.
 36. Sung JY, Jeon HG, Jeong BC, *et al.* Correlation of ERG immunohistochemistry with molecular detection of TMPRSS2-ERG gene fusion. *J Clin Pathol* 2016;69:586–92.
 37. Gsponer JR, Braun M, Scheble VJ, *et al.* ERG rearrangement and protein expression in the progression to castration-resistant prostate cancer. *Prostate Cancer Prostatic Dis* 2014;17:126–31.