Is famine exposure during developmental life in rural Bangladesh associated with a metabolic and epigenetic signature in young adulthood? A historical cohort study

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ABSTRACT

Objectives: Famine exposure in utero can ‘programme’ an individual towards type 2 diabetes and obesity in later life. We sought to identify, (1) whether Bangladeshis exposed to famine during developmental life are programmed towards diabetes and obesity, (2) whether this programming was specific to gestational or postnatal exposure windows and (3) whether epigenetic differences were associated with famine exposure.

Design: A historical cohort study was performed as part of a wider cross-sectional survey. Exposure to famine was defined through birth date and historical records and participants were selected according to: (A) exposure to famine in postnatal life, (B) exposure to famine during gestation and (C) unexposed.

Setting: Matlab, a rural area in the Chittagong division of Bangladesh.

Participants: Young adult men and women (n=190) recruited to a historical cohort study with a randomised subsample included in an epigenetic study (n=143).

Outcome measures: Primary outcome measures of weight, body mass index and oral glucose tolerance tests (0 and 120 min glucose). Secondary outcome measures included DNA methylation using genome-wide and targeted analysis of metastable epialleles sensitive to maternal nutrition.

Results: More young adults exposed to famine in gestation were overweight than those postnatally exposed or unexposed. In contrast, more young adults exposed to famine postnatally were overweight compared to those gestationally exposed or unexposed. Underweight adults exposed to famine in gestation in utero were hyperglycaemic following a glucose tolerance test, and those exposed postnatally had elevated fasting glucose, compared to those unexposed. Significant differences in DNA methylation at seven metastable epialleles (VTRNA2-1, PAX8, PRDM-9, near ZFP57, near BOLA, EXD3) known to vary with gestational famine exposure were identified.

Conclusions: Famine exposure in developmental life programmed Bangladeshi offspring towards diabetes and obesity in adulthood but gestational and postnatal windows of exposure had variable effects on phenotype. DNA methylation differences were replicated at previously identified metastable epialleles sensitive to periconceptual famine exposure.

INTRODUCTION

The global prevalence of type 2 diabetes and obesity is increasing rapidly and their associated vascular complications bring high

Strengths and limitations of this study

▪ This study uses both clinical and molecular tools to investigate the developmental programming hypothesis in a Bangladeshi population at high background risk of type 2 diabetes.

▪ A historical cohort study has allowed detailed analysis of the effects of famine exposure windows during gestation and postnatal life.

▪ We provide important evidence that DNA methylation differences at metastable epialleles sensitive to periconceptual nutritional availability can be replicated in a different setting and ethnic group.

▪ Our genome-wide analysis of epigenetic differences associated with famine exposure in early life was not able to detect DNA methylation differences according to the developmental windows studied, due to the insufficient sample size to overcome the potential multiple hypothesis testing bias.

▪ Future studies should expand on this study design and scaled-up longitudinal resampling of this population for phenotypic and molecular studies will provide an important setting with which to investigate the causative role of early life famine exposure in the programming of future type 2 diabetes.
morbidity and mortality.1 Bangladesh is a South Asian country undergoing demographic transition and with rapidly rising type 2 diabetes prevalence, increasing from 4% in 1990 to 10% in 2010 and projected to have over 10 million people with type 2 diabetes by 2030. Since the observation that low birth weight was associated with increased risk of type 2 diabetes in later life,2 the concept of developmental programming has been considered a possible contributor to this rising prevalence and its ‘missing heritability’, that is, the risk not explained by polygenic or environmental influence alone.3 Exposure to severe nutritional deprivation in early life development has been associated with future risk of type 2 diabetes and obesity in several human and animal models.4-9 South Asian populations are at high risk of type 2 diabetes and obesity,1 and the contribution of fetal programming to this is poorly understood. It is hypothesised that the rapid increase in diabetes risk in South Asians may reflect a mismatch between the development of a fetus in an environment of ‘thrift’ and transition in later life into an environment of nutritional excess, reduced energy expenditure due to changing behaviour patterns, migration (rural to urban, or between countries) and sociodemographic changes.10 11 These concepts may also be important to understand the varied characteristics of diabetes and obesity, including the observation that visceral adiposity despite a relatively low body mass index (BMI) is associated with poorer metabolic health in people of South Asian origin (the so-called ‘thin-fat’ phenotype),12 that severe famine can lead to diabetes in underweight individuals,13 and the existence of a dual burden of overnutrition and undernutrition in transitional populations.14

The aetiology of developmental programming is poorly understood; however, recent epigenetic studies have shown differences in DNA methylation (an epigenetic mark) in association with periconceptual exposure to undernutrition15 and such processes may underlie the gene-environment interactions in the pathogenesis of complex disease16 and developmental programming.17 18 Most early epigenetic studies applied to developmental programming have been limited by lack of replication and/or the use of a candidate gene approach.17 More recent studies designed to identify metastable epialleles (MEs) or that use an epigenome-wide association study approach have been informative and identify methylation differences according to early gestation/periconceptual exposures.15 19-25 The next important step for these data is to replicate across studies in different populations, to identify whether they have a causal role with known programmed phenotypic outcomes such as diabetes and obesity, and whether they contribute to the high burden of these chronic diseases in South Asian populations.

Until now, most studies of developmental programming in South Asians are limited to single micronutrient deficiencies associated with specific dietary behaviours, limiting their wider application to other populations.24 Furthermore, there are no published studies that show programmed phenotypic outcomes in association with molecular aetiology, such as epigenetic differences, in South Asian populations. Developmental programming studies in The Gambia have identified multiple robust regions of epigenetic (DNA methylation) variation in children that are associated with differences in periconceptual nutrient availability, but until now have not found an association with a programmed phenotype. These so-called ‘metastable epialleles’ occur at regions of the epigenome established stochastically in the early embryo, maintained across different cell lineages, and show intrainternal and intertissue stability,19 26 and therefore any inter-individual variation detected at these loci could have significant biological and functional relevance, such as the risk of complex disease in later life. While other studies looking at methylation variants in fetal programming exist, we chose to investigate these MEs due to the robust nature in which they were first identified and characterisation in a contrasting ethnic population, meaning that replication in our cohort would be of potential significance.

With these background data, and their limitations in mind, we performed a historical cohort study of young adults exposed to famine in early life in Matlab, a rural area in the Chittagong division of Bangladesh, using both clinical and epigenetic outcomes. A severe monsoon in 1974 destroyed the majority of the annual rice crop and, in combination with inadequacies in the food distribution system, unemployment and lower purchasing power, a severe famine ensued.20 In Matlab, this famine was defined as occurring between July 1974 and June 1975, and its severity implicated deficiencies of multiple macronutrients and micronutrients in the diet of the local population during that time.21 Increased mortality of the general population in Matlab during and immediately after the famine was noted, as was neonatal and infant mortality.22 We hypothesised that young adults exposed to famine in developmental life would show evidence of programming towards diabetes and obesity, and that there may be varied effects according to timing of exposure, that is, gestational or postnatal exposure. Furthermore, we hypothesised that gestational famine exposure would be accompanied by epigenetic differences at specific metastable epialleles, previously identified to be sensitive to periconceptual nutrition availability in Gambian children.15

METHODS

Participants were recruited across the Matlab area as part of a wider cross-sectional unselected pilot survey of men and women aged 25–64 years.28 Famine exposure windows were determined from birth records and using the famine start and end dates previously described. All individuals included in the survey were grouped according to exposure window resulting in the following: group A (n=81): postnatal famine exposure (born 1–2 years...
before the start of famine); group B (n=40); famine exposure in gestation (including at least 7 months of famine exposure during gestation) and group C (n=70); unexposed (conceived 6 months to 2 years after famine). Group (D) was also defined (n=112), incorporating older offspring exposed after 16 years of age for comparison to a background population. A randomised subsample of individuals from within groups A, B and C was included in the epigenetic study.

Clinical phenotyping was performed during a single study visit, incorporating measurement of BMI, anthropometry, 75 g WHO standard oral glucose tolerance tests (OGTT) and fasting lipids. Standard criteria were used to define diagnoses of metabolic syndrome,\textsuperscript{29} impaired fasting glycaemia, impaired glucose tolerance and type 2 diabetes,\textsuperscript{30} and abnormalities of BMI,\textsuperscript{31} using sex-specific and ethnicity-specific cut-offs where relevant. Standardised questionnaires were used to capture health status and sociodemographic factors. Phenotypic differences relevant to type 2 diabetes and obesity were compared between groups A, B and C were examined using three-way analysis of variance (ANOVA) tests for continuous variables and $\chi^2$ tests for categorical data.

A randomised subset of participants from groups A, B and C (n=148) were selected for the epigenetic study. Whole blood samples from these participants were stored at $-80^\circ$ until genomic DNA extraction was performed, using standard kits (Qiagen, UK). Randomly selected DNA aliquots from groups A, B and C were transported to the UK under a Material Transfer Agreement that involved training Bangladeshi scientists at the receiving UK institution. Genomic DNA samples with a nanodrop 260/280 ratio $<$1.75 and a 260/290 ratio $<$1.8 were discarded from further analysis. DNA was bisulphite-converted using the EZ-methylation kit (Zymo) and efficiency assessed in bioinformatics quality control checks.

The Illumina HumanMethylation 450BeadChip (Illumina, California, USA) was used to assay DNA methylation at CpG dinucleotides on a genome-wide scale, incorporating all designable RefSeq genes and enriched for CpG islands and regulatory regions. The experimental methods used have been previously described\textsuperscript{32} and use bisulphite conversion and Infinium bead chemistry\textsuperscript{33} to quantify methylation at CpG dinucleotides. Normalised, filtered and quality-checked quantitative methylation data were computed; $\beta$ values (0–100% methylation) were derived from the analysis and data were analysed on a genome-wide scale using F-tests between groups A, B and C using Limma\textsuperscript{34} and a Benjamini and Hochberg false discovery rate control.\textsuperscript{35}

We interrogated probe $\beta$ values at probes within 16 MEs (VTRNA2-1, PAX8, PRDM9, HLA-DQB2, PLD6, near ZFDP57, AKAP12, ATP5B, LRRCL1B, SPG20, near BOLA, RBM46, ZFYVE28, EXD3, PARDEG, ZNF678 and ZFYVE28), ‘metastable epialleles’ that showed methylation variation in Gambian children according to nutritional conditions at conception.\textsuperscript{15} 19 20 We also interrogated our data at sites of methylation variation identified in other genome-wide developmental programming studies using a non-MEs approach in the Dutch Winter Hunger and Barbados famine studies.\textsuperscript{21–23}

Targeted bisulphite pyrosequencing was performed as validation in all experimental samples at the PAX8 metastable epialleles.\textsuperscript{15} Samples were assayed in duplicate and mean $\beta$ values used. Primer sequences are available on request. Genomic DNA was converted using a Zymo EZ-DNA Methylation kit (kit number D5001) according to the manufacturer’s protocol. Bisulphite-converted DNA was recovered in 20 $\mu$L of elution buffer. Forward and reverse primers (one of them being biotinylated) flanking the region of interest were designed using the Pyromark Assay Design v2.0.1.15 software and used with Platinum Taq DNA Polymerase to amplify 20 ng of BS-converted DNA. Quantitative short read sequencing of amplified BS-DNA was performed on the pyrosequencer PSQ96MD system using the PyroGold SQA Reagent Kit and analysed with Q-CpG software (V1.0.9, Pyrosequencing AB). $\beta$ values from both targeted bisulphite pyrosequencing and array probe interrogation were compared using three-way ANOVAs and across relevant genomic features (eg, a gene’s promoter) annotated by a gene browser (http://www.ensembl.org).

**RESULTS**

**Clinical data**

Clinical data of the participants, including sex, wealth index, marital status, occupation, educational status, smoking and phenotypic measures, are presented in online Supplementary file 1, table 1. BMI was categorised into underweight (BMI $<$18.5 kg/m$^2$), normal weight (BMI 18.5–22.9 kg/m$^2$) and overweight and obese (BMI $\geq$23 kg/m$^2$) using the WHO reference criteria for Asian populations.\textsuperscript{31} Age differences were present between groups due to the study design. Ordinal regression analysis showed that BMI categorisation was not affected by putative confounding variables (age at follow-up, sex, wealth status, marital status, occupation, educational attainment, smoking or arsenic exposure) (pseudo $R^2$ 0.053, $p=0.34$). Significant differences (Proportional odds model, $p=0.039$) were identified between exposure groups according to weight category: an excess of underweight individuals was identified in those exposed during gestation (group B) (48%) compared to postnatal exposed (group A) (32%) and unexposed (group C) (30%); greater numbers of overweight offspring (26%) were identified in the postnatal exposed group compared to gestational exposed (8%) or unexposed (14%) (see table 1). Comparison of these data with the ‘older’ group of individuals (mean age 48 years, exposed to famine after age 16 years) suggests that both findings are disproportionate to the prevalence of underweight and overweight in the background (and older) population. Data from this older population also highlight the burden of hyperglycaemic
Table 1  Age, body mass, weight circumference and glucose parameters from offspring exposed postnatally, in utero and unexposed to famine, and statistical comparisons of groups A, B and C (proportional odds model for weight as an ordered categorical variable, and ANOVA for continuous variables)

<table>
<thead>
<tr>
<th>Group</th>
<th>Exposure</th>
<th>Group A</th>
<th>Group B</th>
<th>Group C</th>
<th>Statistical tests (groups A,B,C)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Postnatal exposed</td>
<td>In utero exposed</td>
<td>Unexposed</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>n=68</td>
<td>n=70</td>
<td>n=70</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Age (years) Mean 31</td>
<td>30</td>
<td>28</td>
<td>ANOVA p&lt;0.0001</td>
<td>48</td>
</tr>
<tr>
<td></td>
<td>SD 0.4</td>
<td>0.3</td>
<td>0.3</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>BMI (kg/m²) Mean 20.3</td>
<td>19.4</td>
<td>20.0</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>SD 3.1</td>
<td>2.9</td>
<td>2.6</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>BMI category Underweight n (%) 26 (32)</td>
<td>33 (49)</td>
<td>21 (30)</td>
<td>Proportional odds model df=2, deviance ratio=3.25, p=0.039</td>
<td>34 (30)</td>
</tr>
<tr>
<td></td>
<td>Normal weight n (%) 34 (42)</td>
<td>26 (38)</td>
<td>39 (56)</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>45 (40)</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Overweight n (%) 21 (26)</td>
<td>9 (13)</td>
<td>10 (14)</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>33 (30)</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Waist circumference Normal 71 (88)</td>
<td>63 (93)</td>
<td>65 (93)</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>High (M≥ 90 cm; F≥ 80 cm) 10 (12)</td>
<td>5 (7)</td>
<td>5 (7)</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>23 (21)</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>0 min glucose (mmol/l) Mean 4.8</td>
<td>4.8</td>
<td>4.7</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>SD 0.5</td>
<td>0.5</td>
<td>0.6</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>9 (13)</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3.7</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>120 min glucose (mmol/l) Mean 5.2</td>
<td>5.8</td>
<td>5.6</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>SD 1.4</td>
<td>1.6</td>
<td>2.0</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3.7</td>
<td>ns</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>IFG/IGT/T2D n (%) 0=no 74 (81)</td>
<td>59 (87)</td>
<td>60 (86)</td>
<td>ns</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1=yes 7 (9)</td>
<td>10 (14)</td>
<td>ns</td>
<td>ns</td>
<td></td>
</tr>
</tbody>
</table>

Non-significant statistical comparisons (p≥0.05) are marked ‘ns’. Data on older offspring exposed to famine after the age of 16 years are presented (in italics) for reference but are not included in statistical testing.

ANOVA, analysis of variance; BMI, body mass index; IFG, impaired fasting glucose IGT, impaired glucose tolerance; T2D, type 2 diabetes.

Targeted DNA methylation analysis

We used our 450k array data set to assess quantitative DNA methylation at β values [0–1], unmethylated to fully methylated (β=1). All other probes within these genes were interrogated to assess regional patterns of methylation differences, as described. At each probe (n=18) using samples from groups A (n=30), B (n=13) and C (n=36), we performed comparative analysis of mean methylation values from groups A, B and C using F-tests. Neither analysis nor application of a false discovery rate (adjusted p<0.05), and we therefore eliminated any groupwise bias in the randomisation of each exposure group by a latent variable approach to correction yielded methylation differences. A standard t-test was performed, and normalized β values from groups A (n=14), B (n=10) and C (n=16), and a control group to the gestationally exposed group (n=5) that had failed quality control checks, were analyzed with linear models.

We generated high quality single CpG resolution methylation values from 483 382 probes on the Illumina HumanMethylation450 beadchip array (450k array) for 148 samples (74 (A), 35 (B) and 39 (C)), selected from the 6 of the 16 previously reported MEs (MEs) that had a mean 120 min glucose values from groups A, B and C using F-tests. Neither analysis nor application of a false discovery rate (adjusted p<0.05), and we therefore eliminated any groupwise bias in the randomisation of each exposure group by a latent variable approach to correction yielded methylation differences. A standard t-test was performed, and normalized β values from groups A (n=14), B (n=10) and C (n=16), and a control group to the gestationally exposed group (n=5) that had failed quality control checks, were analyzed with linear models.

Genome-wide DNA methylation analysis

We performed comparative analysis of mean methylation values from groups A, B and C in a three-way comparative analysis and two-way analysis (combining groups A+C and B) using F-tests. Neither analysis nor application of a false discovery rate (adjusted p<0.05), and we therefore eliminated any groupwise bias in the randomisation of each exposure group by a latent variable approach to correction yielded methylation differences. A standard t-test was performed, and normalized β values from groups A (n=14), B (n=10) and C (n=16), and a control group to the gestationally exposed group (n=5) that had failed quality control checks, were analyzed with linear models.

There were no significant differences in mean blood glucose, impaired fasting glucose (IFG, [4.8 mmol/L]) or unexposed (4.8 mmol/L) (ANOVA p<0.03) (see table 2 and figure 1). There were no significant differences in mean age, age, fasting lipid measures between groups (data not shown). There were no significant differences in mean 120 min glucose between exposure groups in normal or overweight individuals (table 3).

However, after stratification, those exposed in gestation who were underweight at the time of sampling had a significantly lower mean 120 min glucose (13.9 mmol/L) compared to overweight individuals (16.3 mmol/L) (ANOVA p<0.03) (see table 2 and figure 1). There were no significant differences in mean age, age, fasting lipid measures between groups (data not shown). There were no significant differences in mean 120 min glucose between exposure groups in normal or overweight individuals (table 3).
Table 2 0 min glucose (as part of an oral glucose tolerance test) according to exposure category and stratified by body weight category

<table>
<thead>
<tr>
<th>Weight Category</th>
<th>Group A Postnatal exposed</th>
<th>Group B Gestational exposed</th>
<th>Group C Unexposed</th>
<th>Statistical tests</th>
<th>Group D Older</th>
</tr>
</thead>
<tbody>
<tr>
<td>Underweight</td>
<td>4.9 (n=26)</td>
<td>4.7 (n=33)</td>
<td>4.6 (n=21)</td>
<td>ns</td>
<td>4.8 (n=34)</td>
</tr>
<tr>
<td>Normal weight</td>
<td>4.7 (n=34)</td>
<td>4.9 (n=26)</td>
<td>4.7 (n=39)</td>
<td>ns</td>
<td>5.3 (n=56)</td>
</tr>
<tr>
<td>Overweight</td>
<td>4.7 (n=21)</td>
<td>4.8 (n=9)</td>
<td>4.8 (n=10)</td>
<td>ns</td>
<td>6.1 (n=22)</td>
</tr>
</tbody>
</table>

Sample size per subcategory is noted (n=). Statistical testing performed on groups A, B and C only. Group D is shown for comparison only. ns, non-significant.

Figure 1 Distribution of 120 min glucose values in underweight individuals in groups A (postnatal exposed), B (gestational exposed) and C (unexposed).

Quantitative methylation values derived from the 450k array at PAX8 were validated against a BS-pyrosequencing amplicon targeted to 8 CpG sites overlying and adjacent to the metastable epiallele. BS-pyrosequencing was performed across a wider sample number (due to the higher quality bisulfite conversion) and was highly correlated (R²=0.92).

DISCUSSION
In this cross-sectional study of a rural Bangladeshi population, we have identified diabetes-related phenotypes in young adults exposed to famine in early developmental life. These phenotypes were evident only in those individuals remaining underweight (BMI <18.5 kg/m²) at the time of this follow-up study, when aged 29–31 years. These data add to existing studies suggesting that early life nutritional deficits ‘programme’ offspring towards metabolic disease in later life.4 5 24 Our study adds to existing studies by a clear definition of exposure windows, including gestational exposure, as well as a window of postnatal famine exposure starting at <2 years of age. This study is the first that follows up famine-exposed individuals in a South Asian population and that incorporates the study of potential epigenetic mechanisms underlying the double burden of disease.14

We identified a 30–32% prevalence of underweight across postnatal exposed, unexposed and older experimental groups, suggesting a non-age-dependent background prevalence. In contrast, we observe an excess prevalence of underweight (49%) in young adults exposed to famine in utero, and this appears to be independent of confounding sociodemographic factors. Those adults exposed to famine during both conception and gestation who remain underweight have a higher mean glucose at 120 min postoral glucose challenge, compared to those postnatally exposed or unexposed. The finding of impaired glucose tolerance in the underweight exposed in gestation will benefit from replication in future studies and may suggest a programmed insulin resistance phenotype previously described by others in similar models,2 4 and supports the concept of malnutrition-related diabetes and the ‘thin-fat phenotype’.12 37 We also observed that overweight and obesity were twice as common in young adults who had been exposed to famine during postnatal life (<2 years old), driven by group B (gestationally exposed) compared to either unexposed and/or postnatally exposed. Comparison using non-parametric testing and with/without the inclusion of single nucleotide polymorphism probes did not affect the results. Furthermore, analysis using z-scores of mean methylation across all 16 MEs (to reduce the interepiallele variance in methylation) showed significant and relative hypomethylation in group B (mean z-score −0.24), compared to groups A and C (mean z scores −0.14 and −0.15, respectively) (3-way ANOVA p value 0.0003, Bonferroni multiple comparison test significant between groups A vs B and B vs C). These findings are consistent with the findings of Dominguez-Salasz et al19 and Silver et al20 who showed hypomethylation in Gambian offspring conceived during the rainy season (ie, during nutritional depletion) and hypothesise that this is due to reduced methyl donor availability. Interestingly, PAX8 showed relative hypermethylation in group B compared to groups A and C, but this did not affect the pattern of mean hypomethylation across all epialleles. Methylation data and probe coordinates from all 16 MEs are presented in online supplementary file 1, tables 2 and 3. There was no significant replication of the methylation differences identified in the recent genome-wide Dutch Winter Hunger or Barbados famine studies (see online supplementary file 2).
compared to those exposed in utero or unexposed. The varied body weight phenotype associated with the in utero and postnatal exposure windows suggests a variable influence on developing organ systems and behavioural systems, and supports our inclusion of a distinct postnatal exposure group to investigate the effects of postnatal programming on appetite mechanisms. Other studies include postnatally exposed individuals as ‘unexposed’ controls, but our data suggest the importance of considering this exposure window as a

Table 3  120 min glucose (as part of an oral glucose tolerance test) according to exposure category and stratified by body weight category

<table>
<thead>
<tr>
<th></th>
<th>Group A Postnatal exposed</th>
<th>Group B In utero exposed</th>
<th>Group C Unexposed</th>
<th>Statistical tests</th>
<th>Group D Older</th>
</tr>
</thead>
<tbody>
<tr>
<td>Underweight</td>
<td>4.8 (n=26)</td>
<td>5.8 (n=31)</td>
<td>4.8 (n=21)</td>
<td>ANOVA p&lt;0.02</td>
<td>4.9 (n=34)</td>
</tr>
<tr>
<td>Normal weight</td>
<td>5.3 (n=34)</td>
<td>5.6 (n=26)</td>
<td>5.9 (n=39)</td>
<td>ns</td>
<td>6.4 (n=56)</td>
</tr>
<tr>
<td>Overweight</td>
<td>5.5 (n=21)</td>
<td>5.9 (n=9)</td>
<td>5.8 (n=10)</td>
<td>ns</td>
<td>7.9 (n=22)</td>
</tr>
</tbody>
</table>

Sample size per subcategory is noted (n=). Statistical testing performed on groups A, B and C only. Group D is shown for comparison only. ANOVA, analysis of variance; ns, non-significant.

Figure 2  Box and whisker plots showing β values at 450k probes located at/near metastable epialleles. Boxes (median, 25th and 75th quartiles) show β values from groups A (blue), B (red) and C (green). Probe labels along the X axis in bold denote those probes overlying, or within 100 bp, of the target metastable epialleles (MEs) (Dominguez-Salazar, 2014; Silver, 2015). Three-way group comparisons (ANOVA) showing differences reaching statistical significance (*p<0.05, **p<0.005, ***p<0.001, ****p<0.0001). Mean methylation z-scores across each MEs in the ‘all MEs’ plot are combined to reduce interepiallele variance. ANOVA, analysis of variance; MEs, metastable epialleles.
separate experimental group with that could be susceptible to different epigenetic and phenotypic variation, compared to in utero exposure. Animal and human studies that focus on postnatal life suggest that ‘catch-up growth’ in the first 2 years of life following in utero growth restriction is associated with childhood obesity. It is important to note that famine exposure at any age is likely to be followed by a period of rapid growth acceleration due to sudden nutritional repletion as famine conditions were reversed. It is not possible to tell from this, or other studies, whether it is the growth restriction, acceleration or timing of either that is the determinant of future metabolic disease programming.

Despite strong clinical and epidemiological evidence for fetal programming, there is little understanding of its molecular basis. Current epigenetic studies in this context are limited by methodological issues, lack of replication and validation, use of surrogate tissues and inability to prove functional or causal effects on phenotype. We sought to improve on these limitations with a carefully designed study during different exposure windows and by replicating and validating our data across technological platforms. Our genome-wide, discovery-based approach was limited by a small sample size. We were unable to control for false discovery in the context of multiple hypothesis testing and were therefore unable to produce robust findings with this approach. However, we investigated whether the MEs identified from rigorous epigenetic studies applied to Gambian children exposed to periconceptual nutritional deficiency were also differentially methylated in Bangladeshi adults. We studied the 16 previously identified MEs using data from our 450k arrays and identified methylation differences at 6 of them. These MEs showed relative hypomethylation in association with gestational famine exposure, consistent with the observation of hypomethylation at these regions in periconceptually exposed Gambian children. Interestingly, the MEs at PAX8 and near BOLA showed relative hypermethylation in those exposed during gestation and future studies will need to define more precise exposure windows to elucidate whether there are variable effects according to more detailed windows of exposure or the frequency and characteristics of the famine itself. We note that all historical cohort and epidemiological studies of famine conditions have the potential to be limited by variable exposure and susceptibility to the conditions of famine on an individual level and this could alter the molecular read-out. We validated our 450k methylation data at PAX8 using a bisulphite-pyrosequencing assay and found both platforms to correlate strongly. The design of the 450k array, with probes designed to multiple CpG sites in most genes, allowed us to interrogate regional methylation and we showed spreading patterns of methylation variation around the metastable epialleles. PAX8 is a widely expressed transcription factor that is known to have multiple roles in thyroid cell differentiation and function via transcriptional regulation of a wide gene network. In providing evidence of methylation variation at six MEs in Matlab offspring, we are confident that these epigenetic marks of exposure are unaffected by age at sampling, ethnicity or geographical setting. These findings lend support to the use of whole blood as a means of detecting functionally relevant, robustly identified epigenetic variants, such as metastable epialleles, that might have been set down in early development prior to separation of the three germ layers, or that are stable across tissues. Our lack of replication of the top hits identified in the Dutch Hunger Winter and Barbados famine studies may be due to differences in study design, ethnicity or lack of power, and highlight the challenges of replicating small methylation variants from association-based studies.

There are limitations of our historical cohort study. First, the retrospective analysis of cohort data is unable to elucidate causation and, despite high participation rates in the study, there may be selection bias, for example, favouring a ‘survivor effect’. Second, our study includes only small numbers of individuals and validation of both phenotypic and epigenetic outcomes in larger numbers are now planned to allow multivariate analysis and sex-specificity of outcomes. The sample size also limited our ability to detect epigenomewide differences in DNA methylation that withheld correction for multiple testing. Despite these limitations, our study provides additional important proof-of-concept in support of fetal programming hypotheses, adds to the current literature which relates either to specific micronutrient deficiencies, or, in the case of other famine-based studies, does not show association of both phenotypic and epigenetic outcomes. Finally, we add important insight into the populations at high risk of diabetes and obesity and the notion of the ‘thin-fat’ phenotype in this Bangladeshi group that have been developmentally programmed and are at high risk of diabetes even if underweight.

In conclusion, we have found clinical evidence of developmental programming towards hyperglycaemia in a cohort of underweight young adult Bangladeshi exposed to famine during gestation and postnatal life. We have replicated DNA methylation differences at MEs described in an independent study in those offspring exposed to famine during gestation. Furthermore, we suggest that postnatal exposure to famine may be an important window of exposure to include in this type of study. Our epigenomic study was underpowered to identify new methylation variants using a hypothesis-free approach. Our study has overcome some of the concerns raised by many epigenetic studies, namely lack of reproducibility and external replication. However, larger studies are required to overcome false discovery in epigenomic studies and technical variation between platforms and this data set provides important proof-of-principle and a source of data for future meta-analysis or replication studies. Our findings also highlight the importance of studying the varied clinical characteristics of people at
risk of diabetes, and the contribution of different aetiological factors, such as developmental origins, ethnicity and sociodemographic factors, is crucial to understanding the complex basis of the disease and its increasing global burden. Future studies should also incorporate longitudinal sampling to examine the progression of phenotypes with age, elucidate the stability of the MEs and to add insight into whether they have an associative or causative role. The identification of MEs associated with developmental programming may offer a useful molecular signature of early life influences associated with an increased risk of diabetes. Future studies should investigate whether the findings of developmental programming, via an extreme early life exposure such as famine, can be generalised to a wider population at risk of diabetes, for example, across generations or via less extreme insults. A combined approach of detailed clinical and molecular studies may help to improve diabetes risk prediction in populations undergoing transition, and could help define windows of susceptibility and/or reversibility with which to focus and tailor intervention.

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Contributors
DA, GAH and SF conceived and designed the study, SP and DA recruited to and implemented the clinical study, SF and SI transferred samples from ICDDR,B to London, UK and performed DNA extraction and quality control checks. SF, SI, BWO and MS processed DNA for 450k analysis and epialleles. SF, SI, BWO and MS performed array hybridisation and pyrosequencing. Brian Shine and Stephen Senn gave valuable contributions to statistical analysis. The authors would like to thank the reviewers of this manuscript for their detailed and constructive contributions.

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Data sharing statement
Extra data can be accessed via the Dryad data repository at http://datadryad.org/ with the doi:10.5061/dryad.k67k7.

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