ABSTRACT

Objectives: Identification of patient subpopulations susceptible to develop myocardial infarction (MI) or, conversely, those displaying either intrinsic cardioprotective phenotypes or highly responsive to protective interventions remain high-priority knowledge gaps. We sought to identify novel common genetic variants associated with perioperative MI in patients undergoing coronary artery bypass grafting using genome-wide association methodology.

Setting: 107 secondary and tertiary cardiac surgery centres across the USA.

Participants: We conducted a stage I genome-wide association study (GWAS) in 1433 ethnically diverse patients of both genders (112 cases/1321 controls) from the Genetics of Myocardial Adverse Outcomes and Graft Failure (GeneMAGIC) study, and a stage II analysis in an expanded population of 2055 patients (225 cases/1830 controls) combined from the GeneMAGIC and Duke Perioperative Genetics and Safety Outcomes (PEGASUS) studies. Patients undergoing primary non-emergent coronary artery bypass grafting were included.

Primary and secondary outcome measures: The primary outcome variable was perioperative MI, defined as creatine kinase MB isoenzyme (CK-MB) values ≥ 10× upper limit of normal during the first postoperative day, and not attributable to preoperative MI. Secondary outcomes included postoperative CK-MB as a quantitative trait, or a dichotomised phenotype based on extreme quartiles of the CK-MB distribution.

Results: Following quality control and adjustment for clinical covariates, we identified 521 single nucleotide polymorphisms in the stage I GWAS analysis. Among these, 8 common variants in 3 genes or intergenic regions met p<10−5 in stage II. A secondary analysis using CK-MB as a quantitative trait (minimum p=1.26×10−3 for rs609418), or a dichotomised phenotype based on extreme CK-MB values (minimum p=7.72×10−6 for rs4834703) supported these findings.

Pathway analysis revealed that genes harbouring top-scoring variants cluster in pathways of biological relevance to extracellular matrix remodelling, endoplasmic reticulum-to-Golgi transport and inflammation.

Conclusions: Using a two-stage GWAS and pathway analysis, we identified and prioritised several potential susceptibility loci for perioperative MI.

INTRODUCTION

Despite advances in surgical techniques and pharmacological therapy, the incidence of myocardial infarction (MI) after coronary artery bypass grafting (CABG) remains as high as 19%, and is associated with increased mortality and long-term morbidity. Strategies to identify subpopulations of patients at risk for developing large myocardial infarcts on the one hand, or those displaying an intrinsic cardioprotective state on the other hand, remain high-priority knowledge gaps and could inform selection of more specific protective agents. The evidence for heritability of MI is striking, supported both by family studies and, recently, by a number of well powered and replicated genome-wide association studies (GWAS), which primarily implicate common genetic variants at the 9p21 locus in multiple racial groups. However, although family-
based methods are not practical for studying perioperative MI (PMI), its genetic basis is strongly suggested by several observations, including wide variability in incidence and severity that is poorly explained by clinical and procedural risk factors, different racial susceptibility profiles and results from preclinical animal models. Indeed, extensive genetic variability has been found in biological pathways implicated in the pathophysiology of postoperative MI, such as the complex acute inflammatory response to cardiac surgery. Mounting evidence for heritability of a proinflammatory response to cardiac surgery. Yet only a few studies have identified allelic associations with altered susceptibility to myocardial ischaemia-reperfusion injury in cardiac surgical populations, all based on a candidate gene association approach.8–12 Thus, the overall influence of common genetic variation on the incidence of PMI remains poorly understood.

Recently, integrated testing of genes involved in the same biological pathway has emerged as an alternative strategy for evaluating the combined effects of multiple genetic variants with small effect size on a disease phenotype.13–15 Given the polygenic nature of disease susceptibility, this approach is increasingly being used to identify groups of gene variants with shared cellular function that are enriched for disease, while also improving the statistical power of GWAS. In this study, we adopted this strategy by first employing genome-wide association methodology to identify common genetic variants associated with PMI after CABG, followed by pathway-based analyses to uncover biological mechanisms of relevance to PMI.

METHODS

The study design and reporting of the results follow the recommendations by ‘Strengthening the Reporting of Genetic Association Studies’ (STREGA).16 We performed a joint two-stage17 GWAS combined with a pathway analysis approach.

Patient populations

The stage I cohort (discovery cohort) comprised 1493 ethnically diverse subjects who underwent an isolated CABG with cardiopulmonary bypass for the first time and were enrolled between 2002 and 2003 in the Institutional Review Board (IRB) approved Genetics of Myocardial Outcomes and Graft Failure (GeneMAGIC) ancillary study of the multicentre Project of Ex-vivo Vein Graft Engineering via Transfection (PREVENT-IV).18 Of those, 1433 patients met eligibility requirements after applying quality control criteria and excluding patients with missing genotypes or phenotypic information.

In stage II, we tested the top genetic variants in an expanded data set in which an additional 622 patients of self-reported European ancestry were added to the discovery data set, leading to a total of 2055 patients. The additional patients underwent CABG with cardiopulmonary bypass between 1997 and 2006 as part of the Perioperative Genetics and Safety Outcomes Study, an IRB-approved longitudinal study at the Duke University Medical Center.11 12

Definition of PMI

PMI was defined according to the universal definition of MI19 as an elevation in the plasma level of creatine kinase MB isoenzyme (CK-MB) that was >10 times the upper limit of normal, as measured by a core laboratory within 24 h after surgery, and that was not attributable to an intervening clinical event or preoperative MI (adjudicated by the PREVENT-IV Clinical Events Committee).18

Genotyping and quality controls

Genomic DNA was isolated from whole blood or saliva using standard procedures. Genotyping in both cohorts was performed on the Illumina Human610-Quad BeadChip at the Duke Genomic Analysis Facility. Sample and genotype quality control of data flow included assessment of call rates, gender check, cryptic relatedness, SNP missingness and the Hardy-Weinberg equilibrium, as previously described.20 We used principal components derived from the EIGENSTRAT21 method to control for population stratification (see online supplementary methods).

Statistical analysis

Univariate regression analysis was performed to test differences in demographic, clinical and procedural characteristics between patients with and without postoperative MI; statistically significant covariates were subsequently used to adjust genetic association tests. Genome-wide association analyses performed in the stage I cohort used multivariable logistic regression models implemented in PLINK 1.07, assuming an additive genetic model, and including significant clinical covariates and the top 10 principal components to adjust for population stratification (see online supplementary table S1). Statistical significance for stage I analyses was a priori arbitrarily defined as a two-tailed p<0.001, to balance between the overly conservative Bonferroni correction and type I error, given that we had an a priori defined replication data set to obviate type I error. In stage II analyses, the same clinical covariate and principal component adjustments (re-estimated for the expanded data set) were applied, with statistical significance defined as a Bonferroni adjusted p value of 0.05/number of SNPs identified in stage I, to control the overall type I error rate. Haplotype association analysis was performed in the stage II cohort for genes tagged by the significant SNPs, adjusting for the same clinical covariates and principal components. Secondary analyses were performed for the top SNPs identified in the stage II cohort, using the continuous phenotype of CK-MB values and a dichotomised extreme CK-MB
phenotype, which included participants within the first and fourth quartiles of the CK-MB distribution. For CK-MB, a linear regression model was applied with adjustment for the same set of covariates as in the primary analysis. For the extreme CK-MB subset, we reasigned trait status as ‘0’ for participants within the first CK-MB quartile, and ‘1’ for participants within the fourth CK-MB quartile from the stage II cohort. Logistic regression with the same set of covariates was performed. Finally, we employed pathway analysis to prioritise association results and provide biological interpretation, using functional ontology enrichment analysis tools implemented in MetaCore (GeneGO, St. Joseph, MI; see online supplementary methods).

RESULTS
PMI was observed in 112 of 1433 patients (7.8%) and in 225 of 2055 patients (10.9%) in stage I and II cohorts, respectively (see online supplementary table S2). Aortic cross-clamp time, number of coronary artery grafts, and procedures in addition to CABG were significantly associated with PMI in stage I analyses. Stage II analyses showed that extracardiac arteriopathy and year of surgery also played significant roles. All the clinical variables identified above were included in multivariable logistic regression models to adjust the genetic association tests for potential confounders (see online supplementary table S2).

After applying quality control criteria (see online supplementary methods: A.1. Quality control of data flow), 534 390 markers were analysed for association with PMI. The top 2 SNPs (rs10454444 and rs10913237), located in the pregnancy-associated plasma protein A2 gene (PAPPA2), were in high linkage disequilibrium, with p values for stage II of 2.43×10⁻⁶ (OR 0.46; 95% CI 0.33 to 0.63) and 2.47×10⁻⁶ (OR 0.46; 95% CI 0.33 to 0.63), respectively. In addition, one intronic SNP in histone deacetylase-4 (HDAC4), two in the SEC24 family, member D (SEC24D, a member of the cytoplasmic coat protein complex II, COPII), and two located in intergenic regions were associated with PMI (table 1).

When CK-MB was tested as a quantitative trait, all eight SNPs remained significantly associated with plasma levels of CK-MB. However, rs4834703 in SEC24D (p=7.72×10⁻⁶) and rs10200850 in HDAC4 (p=8.26×10⁻⁵) showed the strongest association, followed by the 2 SNPs in PAPPA2 (p=0.0001; table 2). When the dichotomised extreme CK-MB phenotype was studied in the stage II cohort, only the 2 SNPs located in SEC24D (rs4834703 and rs6822035, p=0.002) and rs609418 in the intergenic region between RFXAP and SMAD9 (p=0.001) remained nominally significant (table 2). Overall, the rs4834703 SNP in SEC24D consistently showed strong association signals across postoperative MI, quantitative CK-MB and the extreme CK-MB traits.

The haplotype structures of genic regions surrounding the significant SNPs in PAPPA2, HDAC4 and SEC24D are shown in online supplementary figure S3, and the results of haplotype analysis are shown in table 3 and online supplemental table S3. Haplotype analysis performed for the linkage disequilibrium blocks containing the significant markers showed that the A-A haplotype (rs6822035, rs10518325) in SEC24D had the most.

Figure 1  Manhattan plot of genome-wide association with perioperative myocardial infarction in stage I analysis. The x axis represents the genome in physical order (coloured by chromosome); the y axis showing −log₁₀(p) for all single nucleotide polymorphisms (SNPs). None of the SNPs reached genome-wide significance (p<9.09×10⁻⁸), but 521 SNPs met the prespecified discovery threshold p<0.001 for inclusion in stage II analyses (minimum p=2.76×10⁻⁶, rs2044061 on chromosome 8).
**Table 1. Top eight SNP associated with postoperative myocardial infarction**

<table>
<thead>
<tr>
<th>Chr</th>
<th>SNP</th>
<th>Base pair</th>
<th>Gene symbol</th>
<th>MAF Controls N=1830</th>
<th>MAF Cases N=112</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>rs10489478</td>
<td>176,566,330</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>0.11</td>
<td>0.47 (0.34 to 0.68)</td>
<td>4.70×10^−6</td>
</tr>
<tr>
<td>1</td>
<td>rs10913237</td>
<td>176,630,161</td>
<td>PAPPA2</td>
<td>0.19</td>
<td>0.19</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>4</td>
<td>rs10454444</td>
<td>176,649,181</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>0.11</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>1</td>
<td>rs10518325</td>
<td>176,649,181</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>0.11</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>4</td>
<td>rs102000850</td>
<td>241,240,601</td>
<td>HDAC4</td>
<td>0.18</td>
<td>0.18</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>6</td>
<td>rs10518325</td>
<td>176,649,181</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>0.11</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>6</td>
<td>rs8834703</td>
<td>177,691,621</td>
<td>SEC24D</td>
<td>0.09</td>
<td>0.06</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>12</td>
<td>rs2303970</td>
<td>74,471,340</td>
<td>TRHDE</td>
<td>0.48</td>
<td>0.36</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
<tr>
<td>13</td>
<td>rs609418</td>
<td>37,417,427</td>
<td>SMAD9</td>
<td>0.25</td>
<td>0.36</td>
<td>0.46 (0.33 to 0.63)</td>
<td>2.47×10^−6</td>
</tr>
</tbody>
</table>

*Adjusted for clinical characteristics including extracardiac arteriopathy, recent myocardial infarction, procedure other than CABG only, year of surgery, number of diseased vessels and aortic HDAC4, histone deacetylase-4; PAPPA2, pregnancy-associated plasma protein A2; SEC24D, SEC24-related protein D; SMAD9, mothers against decapentaplegic homolog 9; TRHDE, thyrotropin-releasing hormone-degrading ectoenzyme; CAGB, coronary artery bypass grafting; Chr, chromosome; MAF, minor allele frequency (based on the discovery data set); SNP, single nucleotide polymorphisms.

**DISCUSSION**

We present the first report of a two-stage GWAS involving 225 PMI cases and 1830 controls. After accounting for clinical and procedural covariates, we identified eight significant SNPs mapped to three genes (PAPPA2, HDAC4, SEC24D) and two intergenic regions. The most significant association with PMI was exhibited by rs10116277 SNP previously associated with MI and mortality in non-surgical20 and cardiac surgical8 9 cohorts (see online supplementary figure S4).

The top 10 enriched pathway maps for the stage I analysis SNPs are shown in online supplementary figure S5. The most significant were ‘Cell adhesion: extracellular matrix remodelling’ and ‘Cytoskeleton remodelling: TGF, WNT, and cytoskeletal remodelling’ (enrichment p value=1.9×10^−8 and 7.3×10^−6, respectively). The most significant canonical pathway maps in stage I and II comparative analyses were ‘Immune response: NFAT signalling and leucocyte interaction’ and ‘Cell adhesion: extracellular matrix remodelling’ (p=7.2×10^−4 and 1.1×10^−3, respectively; see online supplementary tables S4 and 5).

In multivariate risk factor-adjusted stage I and II analyses, we found no evidence for association between genetic variation at the 9p21 locus and incident postoperative MI, including the rs10116277 SNP previously associated with MI and mortality in non-surgical20 and cardiac surgical8 9 cohorts (see online supplementary figure S4).

Animal
risk biomarker for the diagnosis and prognosis of acute coronary syndrome; however, the role of PAPPA2 in cardiovascular biology has not been previously reported. Consistent with our single-marker analysis findings, the GWAS pathway analysis identified significant contributions of variants in other IGF system component genes, namely IGF-2 and the IGF-1 receptor (see online supplementary figure S6 and table S5).

**HDAC4**, a member of the HDAC family, mediates changes in the chromatin structure by removing acetyl groups from the core histones, resulting in transcriptional repression. HDAC4 is highly expressed in the myocardium, where it plays an important role in the regulation of gene expression and is involved in myocardial cell cycle progression, differentiation and apoptosis. Experimental HDAC inhibition is associated with a profound reduction in ischaemia-induced myocardial cell death, by triggering preconditioning effects and models of cell death, 

### Table 2

Association of the top eight SNP with CK-MB as a quantitative trait and as an extreme phenotype in the joint analysis data set

<table>
<thead>
<tr>
<th>Chr</th>
<th>SNP</th>
<th>Base pair</th>
<th>Gene symbol</th>
<th>MAF (Overall)</th>
<th>β-Coefficient</th>
<th>p Value</th>
<th>CK-MB as a quantitative trait*</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>rs10489478</td>
<td>176 566 350</td>
<td>PAPPA2</td>
<td>0.17</td>
<td>−5.40</td>
<td>2.27×10^−4</td>
<td>0.77 (0.574 to 1.047)</td>
<td>0.1</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>rs10913237</td>
<td>176 630 416</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>−5.58</td>
<td>1.30×10^−4</td>
<td>0.78 (0.578 to 1.052)</td>
<td>0.1</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>rs10454444</td>
<td>176 649 181</td>
<td>PAPPA2</td>
<td>0.18</td>
<td>−5.59</td>
<td>1.21×10^−4</td>
<td>0.77 (0.575 to 1.043)</td>
<td>0.09</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>rs12000850</td>
<td>240 234 901</td>
<td>HDAC4</td>
<td>0.07</td>
<td>9.26</td>
<td>8.26×10^−5</td>
<td>1.49 (0.950 to 2.326)</td>
<td>0.08</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>rs4834703</td>
<td>119 691 624</td>
<td>SEC24D</td>
<td>0.10</td>
<td>8.54</td>
<td>7.72×10^−6</td>
<td>1.82 (1.242 to 2.654)</td>
<td>0.002</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>rs6822035</td>
<td>119 710 421</td>
<td>SEC24D</td>
<td>0.30</td>
<td>4.42</td>
<td>4.81×10^−4</td>
<td>1.48 (1.152 to 1.903)</td>
<td>0.002</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>rs2303970</td>
<td>74 471 340</td>
<td>TRHDEI</td>
<td>0.47</td>
<td>−2.93</td>
<td>0.01</td>
<td>0.84 (0.669 to 1.047)</td>
<td>0.12</td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>rs609418</td>
<td>37 417 427</td>
<td>RFXAP/SMAD9</td>
<td>0.26</td>
<td>4.80</td>
<td>2.26×10^−4</td>
<td>1.56 (1.189 to 2.034)</td>
<td>0.001</td>
<td></td>
</tr>
</tbody>
</table>

*Adjusted for clinical characteristics including extracardiac arteriopathy, recent myocardial infarction, procedure other than CABG only, year of surgery, number of diseased vessels, aortic cross-clamp time.

CABG, coronary artery bypass grafting; Chr, chromosome; CK-MB, creatine kinase MB isoenzyme; MAF, minor allele frequency (based on the expanded data set); SNP, single nucleotide polymorphisms.

### Table 3

Estimated haplotype frequencies in the **SEC24D** gene and results of association tests with incidence of PMI in the stage II analysis cohort (n=2055)

<table>
<thead>
<tr>
<th>SNP1</th>
<th>SNP2</th>
<th>Haplotype</th>
<th>Patients with PMI (n=225)</th>
<th>Patients without PMI (n=1830)</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs6828577</td>
<td>rs6822035</td>
<td>A-A</td>
<td>0.3956</td>
<td>0.2874</td>
<td>1.65 (1.33 to 2.06)</td>
<td>8.84×10^−6</td>
</tr>
<tr>
<td>rs6828577</td>
<td>rs6822035</td>
<td>G-C</td>
<td>0.6044</td>
<td>0.7126</td>
<td>Reference</td>
<td></td>
</tr>
<tr>
<td>rs6822035</td>
<td>rs10518325</td>
<td>A-G</td>
<td>0.03516</td>
<td>0.0528</td>
<td>0.86 (0.43 to 1.70)</td>
<td>0.657</td>
</tr>
<tr>
<td>rs6822035</td>
<td>rs10518325</td>
<td>C-G</td>
<td>0.00903</td>
<td>0.02636</td>
<td>1.39 (0.67 to 2.90)</td>
<td>0.378</td>
</tr>
<tr>
<td>rs6822035</td>
<td>rs10518325</td>
<td>A-A</td>
<td>0.3631</td>
<td>0.2358</td>
<td>1.87 (1.46 to 2.39)</td>
<td>5.54×10^−7</td>
</tr>
<tr>
<td>rs6822035</td>
<td>rs10518325</td>
<td>C-A</td>
<td>0.5927</td>
<td>0.685</td>
<td>Reference</td>
<td></td>
</tr>
<tr>
<td>rs10518325</td>
<td>rs11098451</td>
<td>A-A</td>
<td>0.03097</td>
<td>0.06231</td>
<td>0.76 (0.48 to 1.19)</td>
<td>0.225</td>
</tr>
<tr>
<td>rs10518325</td>
<td>rs11098451</td>
<td>G-G</td>
<td>0.04425</td>
<td>0.07845</td>
<td>0.87 (0.58 to 1.31)</td>
<td>0.509</td>
</tr>
<tr>
<td>rs10518325</td>
<td>rs11098451</td>
<td>A-G</td>
<td>0.9248</td>
<td>0.8592</td>
<td>Reference</td>
<td></td>
</tr>
</tbody>
</table>

Adjusted for clinical characteristics including extracardiac arteriopathy, recent myocardial infarction, procedure other than CABG only, year of surgery, number of diseased vessels, aortic cross-clamp time.

CABG, coronary artery bypass grafting; PMI, perioperative myocardial infarction; SNP, single nucleotide polymorphisms.
promoting myocardial repair. Genetic variants in HDACs are important determinants of susceptibility to cardiovascular diseases, suggesting functional roles for HDAC4 in modulating perioperative myonecrosis, and their potential use in predicting individual patient responsiveness to HDAC inhibition.

One of the intergenic SNPs associated with PMI (rs609418) is located near the mother against the deca-pentaplegic homologue 9 (SMAD9) gene, part of the transforming growth factor β (TGF-β) signalling pathway, which is markedly activated in the infarcted myocardium. Members of the TGF-β superfamily transduce their signal from the membrane to the nucleus via a distinct combination of transmembrane receptors and downstream effectors—the SMAD proteins. TGF-β plays important and complex roles in regulating postinfarction inflammatory responses. In animal models, signalling through the SMAD transcription factors is associated with resolution of inflammation, repression of cytokine and chemokine gene synthesis, and protection against myocardial ischaemia-reflow injury.

Consistent with this single-locus gene association result, the bone morphogenetic protein pathway, which also transduces its signals via a SMAD9-dependent cascade, was identified as one of the top-scoring pathways in our GWAS pathway analysis (p=5.2×10⁻⁶, see online supplementary table S7). Although no direct functional roles are currently attributed to this intergenic region, a query of the Regulome and Haploreg databases reports that rs609418 is located within active regulatory elements. The GWAS pathway analysis (p=5.2×10⁻⁶, see online supplementary table S7). Although no direct functional roles are currently attributed to this intergenic region, a query of the Regulome and Haploreg databases reports that rs609418 is located within active regulatory elements (GATA2 transcription factor binding site by ENCODE). The regulatory motif in G6f1 and Me2 by the Position-Weight Matrix, see online supplementary figure S7).

Surprisingly, we have been unable to replicate previously reported associations between common genetic variants at the 9p21 locus and risk for PMI or mortality after CABG. Many different reasons may account for this, including inadequate sample size, variation in study design, differences in allele frequencies or variability in the definition of PMI phenotype. Indeed, our observation is consistent with other studies showing that, although genetic variants at the 9p21 locus are associated with incident coronary artery disease (CAD), they may not be associated with the actual risk of MI. This discrepancy between convincing associations of 9p21 with a greater burden of CAD but not with MI in the presence of underlying CAD has been further confirmed by nested case-control studies as well as meta-analyses. Taken together, in subjects with established CAD, such as those included in our study, any lack of association of the 9p21 locus with subsequent MI could have resulted from the presence of CAD in carriers and non-carriers of the risk variants at the 9p21 locus, which seems to primarily mediate an atherosclerotic phenotype.

The strengths of our study are (1) a relatively large population of cardiac surgery patients, (2) a prospective cohort design and (3) a combination of complementary single-marker and pathway-based genome-wide association analyses. This approach allowed us to identify genetic variants that carry only a small disease risk individually, but that jointly can contribute relatively large effects on PMI susceptibility. Furthermore, an application of results from pathway-based analysis may add structure to interpreting genomic data and allow exploration of cellular processes that functionally underpin the observed associations. Finally, by using this approach, replication of association findings at the gene and pathway level is much easier compared to replication at the individual SNP level. Of note, most of the genes identified through pathway enrichment analysis in this study encode targets for therapeutic drugs that have already been developed. Thus, by improving risk assessment and identifying allele-specific therapeutic responses, further investigation of loci and pathways prioritised in this study could yield actionable results for enhancing perioperative cardioprotection.

Several limitations are worth mentioning. Power calculations (see online supplementary results B.6.) show that, based on the current sample size and incidence of PMI, our study can detect a genotypic relative risk approximately 2 with 80% power (assuming a variant with a 10% minor allele frequency and a realistic linkage disequilibrium between the tested marker and the causal locus D²=0.8). Thus, although our study is the largest genetic association study of PMI conducted to date, it is powered to detect only common variants with relatively large effect sizes. Most published genetic association studies of PMI have reported larger effect sizes compared with ambulatory populations (OR range 1.79–3.97). The possibility of rare genetic variants that drive a pronounced clinical phenotype was not explored in this study, because only variants with minor allele frequencies >0.05 were assessed. Also, functional studies to further elucidate the potential biological effects of the SNPs identified were not feasible due to lack of plasma or tissue availability in the GeneMAGIC study. Finally, patients enrolled in our cohorts were predominantly Caucasian, and therefore our findings cannot be generalised to other ethnic groups.

In conclusion, we report the first GWAS in a cohort of patients at risk for MI after CABG surgery. On the basis of our integrated approach utilising primary (PMI) and secondary phenotypes (quantitative CK-MB, extreme CK-MB), single-marker analysis and pathway analysis, we identified several polymorphisms in the insulin growth factor system implicated in the regulation of extracellular matrix remodelling, as well as the ER-to-Golgi secretory pathway potentially involved in adaptive responses to ER stress. As other GWAS of PMI cohorts publish their results, we intend to collaborate on conducting a meta-analysis for this particular phenotype. While our GWAS results are intriguing, follow-up studies are needed to translate these initial findings into biological insights that could lead to predictive and therapeutic advances in perioperative care. For instance, in the...
regions of confirmed associations, causal variants will only occasionally be among those directly genotyped. Moreover, GWAS detect almost exclusively the effects of common SNPs, offering limited power to capture any rare and structural variants, such as insertions, deletions, inversions and translocations. Detailed sequencing may be necessary to further characterise the genetic variations in the PMI-associated regions to enhance the identification of causal variants. Furthermore, as in most cases, the functions of identified genes and their variants, as well as the mechanisms by which they may contribute to PMI pathophysiology, are largely unknown. Currently, very few existing groups can bridge from genetics to molecular biology or cell physiology and to disease or novel therapy; we propose to examine the mechanisms by which variation in the observed genes is involved in myocardial ischaemia-reperfusion injury by examining their functional effects in the human myocardium from patients undergoing cardiac surgery, as well in our previously described preclinical rodent and swine models of cardioplegic arrest and cardiopulmonary bypass. The use of such animal models would allow pharmacological studies targeting the identified pathways to explore the mechanism by which novel cardioprotective drugs would attenuate myocardial ischaemia-reperfusion injury, with the ultimate goal of developing personalised cardioprotective strategies.

Author affiliations
1 Division of Cardiothoracic Anesthesiology, Duke University, Durham, North Carolina, USA
2 Department of Biostatistics and Bioinformatics, Duke University, Durham, North Carolina, USA
3 Duke Molecular Physiology Institute; Duke University, Durham, North Carolina, USA
4 Division of Cardiology, Duke University, Durham, North Carolina, USA
5 Duke Clinical Research Institute; Duke University, Durham, North Carolina, USA
6 Cardiac Surgery; Duke University, Durham, North Carolina, USA

Contributors MDK contributed to the analysis and interpretation of data, as well as the drafting of the manuscript. Y-JL contributed to the analysis and interpretation of data and a critical review of the manuscript. JPM contributed to the acquisition of data and a critical review of the manuscript. Y-JL contributed to the analysis and interpretation of data, as well as the drafting of the manuscript. Y-JL contributed to the analysis and interpretation of data, as well as the drafting of the manuscript. The original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

REFERENCES


Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the user is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/


