Mansonella ozzardi corneal lesions in the Amazon: a cross-sectional study

Lucas Monferrari Monteiro Vianna,1 Marilaine Martins,2 Marcos Jacob Cohen,3 Jacob Moyses Cohen,3 Rubens Belfort Jr1

ABSTRACT

Objectives: To characterise and confirm the presence of Mansonella ozzardi microfilariae in the cornea by biomicroscopy and corneal confocal microscopy.

Design: Cross-sectional study.

Settings: Clinical practice study in patients from rural communities in Coari city on the Solimões river, Amazonas state, Brazil.

Participants: The eyes of 212 consecutive volunteer patients were examined using a flash light and their blood checked for the presence of microfilariae by an expert microscopist. Patients with suspicious corneal lesions (characterised as nummular keratitis) were submitted to biomicroscopy, fundoscopy and corneal confocal microscopy evaluation (CCME). In two patients, a biopsy of the limbal conjunctiva adjacent to the nummular keratitis was carried out and blood collected from the surgical wound for microfilariae investigation by thick blood film examination.

Primary and secondary outcome measures: Positive correlation between corneal biomicroscopic and confocal lesions and Mansonella ozzardi microfilaremia.

Results: Of the 212 patients, 56 (26.4%) were positive for microfilaremia. 22 patients with nummular keratitis were positively correlated to microfilaremia (p=0.0001). At biomicroscopy, lesions were classified as quiescent or active. At CCME, lesions were categorised as circular or filiform. The associations between corneal lesions, CCME findings and microfilaremia are shown.

Conclusions: We describe Mansonella ozzardi microfilariae in the cornea and the associated eye pathology. Further studies using ocular tissue PCR and other imaging techniques would be helpful.

INTRODUCTION

Hundreds of types of filariae have been described. Mansonella ozzardi is one of the few causing infection in humans and was first reported as being associated with corneal disease in 1998.1 Mansonella ozzardi is a filaria exclusively found in the America continent and one of the causes of mansonelliasis.2 In Brazil it is found in the states of Roraima, Mato Grosso and Amazonas.3–5 The microfilariae of M ozzardi are found in peripheral blood and can be identified by their morphological characteristics or by molecular biology.6–11

The clinical features of mansonelliasis are not well described. Infected individuals are generally asymptomatic or have signs and symptoms common to other infections such as fever, cold legs, joint pain and headache.12 Signs and symptoms may also include skin and ocular lesions1–3 13–15 and be associated with Wolbachiae, which are bacterial endosymbionts of insects and many filarial nematodes and whose products trigger inflammatory responses.14

Coari city on the Solimões river (04°08’S, 65°07’W) has a high prevalence of M ozzardi infection. Studies showed an average infection rate of between 13.3% and 18.9%, with a higher prevalence in rural areas and older patients.2 14 Other studies conducted
in non-endemic areas found no *M. ozzardi* infection.\textsuperscript{15} Onchocerciasis is not found in this region of the Amazon.\textsuperscript{16}

The objective of the present study is to characterise and confirm the presence of *M. ozzardi* microfilariae in the cornea and to demonstrate the positive correlation between corneal biomicroscopic and confocal lesions and *M. ozzardi* microfilaremia.

**METHODS**

This was a cross-sectional, clinical practice study of 212 consecutive volunteer patients from rural communities of Coari city on the Solimões river, Amazonas state, Brazil, examined in August 2010. All patients had their eyes examined using a flash light and their blood checked for the presence of microfilariae by an expert microscopist. Peripheral blood infection by *M. ozzardi* was determined by light microscopy examination (200× and 400×) of Giemsa-stained thick smears of peripheral blood obtained by digital puncture with a sterile disposable lancet,\textsuperscript{6} Knott and polycarbonate membrane filtration and by a new identification protocol using PCR (M Martins et al, unpublished observations).

Patients with nummular keratitis, similar to those previously described in the literature,\textsuperscript{1} 13 14 at unilateral or bilateral eye examination were subjected to bilateral biomicroscopy, fundoscopy and corneal confocal microscopy evaluation (CCME) using the Rostock Cornea Module of the Heidelberg Retina Tomograph. Slit lamp photographs were taken. The CCME was performed on the central cornea of both eyes and on the corneal regions affected by nummular keratitis.

Two patients underwent biopsy of the limbal conjunctiva adjacent to the area of nummular keratitis and blood was collected from the surgical wound for microfilariae investigation by thick blood film examination.

Inclusion criteria were volunteering for a blood check for *M. ozzardi* microfilaremia and signing the informed consent form or have it signed by a legal representative. Exclusion criteria were not volunteering or unable to sign the informed consent form.

This was a prospective study and was approved by the UNIFESP Ethics Committee (number 0767/10). All participants signed the protocol’s informed consent form.

Data were presented in contingency tables and Fisher’s exact test was used to compare proportions. \( p \) Values less than 0.05 were considered statistically significant. Analyses were carried out in Stata V.11 (College Station, Texas, USA).

**RESULTS**

Of the 212 patients, 56 (26.4\%) were positive for microfilaremia. Twenty two patients with nummular keratitis

<p>| Table 1 Association between corneal lesions and microfilaremia detected by Giemsa staining, Knott or polycarbonate membrane filtration and/or PCR* |
|--------------------------------------|-----------------|-----------------|---------|</p>
<table>
<thead>
<tr>
<th>Corneal lesions</th>
<th>Positive (+)</th>
<th>Negative (−)</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Microfilaremia</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Positive (+)</td>
<td>14 (63.6%)</td>
<td>42 (22%)</td>
<td>56</td>
</tr>
<tr>
<td>Negative (−)</td>
<td>8 (33.4%)</td>
<td>148 (88%)</td>
<td>156</td>
</tr>
<tr>
<td>Total</td>
<td>22</td>
<td>190</td>
<td>212 (100%)</td>
</tr>
</tbody>
</table>

\(*p=0.0001.\)

**Figure 1** Clinical features. (Left and bottom) Central irregular ‘mottled’ appearance surrounded by an opaque halo; (right) four typical keratitis lesions with a normal translucent area between the lesions and the limbus, but no corneal neovascularisation.
identified by flash light examination underwent biomicroscopy and CCME. Four of the patients had bilateral lesions. The statistically significant association between microfilaremia and corneal lesions is shown in table 1 (p=0.0001).

At biomicroscopy, some lesions were characterised as nummular, mid periphery and anterior stromal corneal opacities, 0.5–1.0 mm in diameter with a central mottled appearance surrounded by an opaque halo, and with a marked boundary between the white opacity and the clear cornea. Other lesions were characterised by nummular keratitis, also located at the mid periphery and anterior stroma. In all cases there was a normal translucent area between the corneal lesions and the limbus and an absence of corneal neovascularisation or other changes (figure 1). The clinical characteristics of the lesions as well as their high prevalence exclude the possibility of trauma or other known causes of the differential diagnosis, specifically onchocerciasis. The association between nummular keratitis and microfilaremia is shown in table 2 (p=0.1347).

Among the 22 patients with corneal lesions at biomicroscopy, 12 also had suspected related lesions at CCME. These confocal suspected lesions displayed two patterns: circular (increased reflectivity, measuring about 80 µm in diameter, located at the sub-epithelial level, as shown in figure 2) and linear (filiform lesions 250–300 µm in length and 10–15 µm in width, with one ‘C’-shaped end and the other tapered, as shown in figure 3). Five patients had only circular lesions, one patient had only linear lesions and six patients had both. The association between confocal lesions and microfilaremia is shown in table 3 (p=0.4266).

**Table 2** Association between corneal lesions and microfilaremia detected by Giemsa staining, Knott or polycarbonate membrane filtration and/or PCR*.

<table>
<thead>
<tr>
<th>Microfilaremia</th>
<th>Keratitis</th>
<th>Opacities</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive (+)</td>
<td>10 (76.9%)</td>
<td>4 (44%)</td>
<td>14</td>
</tr>
<tr>
<td>Negative (−)</td>
<td>3 (23.1%)</td>
<td>5 (56%)</td>
<td>8</td>
</tr>
<tr>
<td>Total</td>
<td>13</td>
<td>9</td>
<td>22 (100%)</td>
</tr>
</tbody>
</table>

*p=0.1347.

Examination of thick blood smears from the limbal conjunctiva in two patients with linear lesions confirmed microfilaremia. Figure 4 compares the *M ozzardi* microfilaremia seen in the thick blood smear with the confocal microscopy image.

**DISCUSSION**

There is a high prevalence of *M ozzardi* in some riverside communities in the Amazon with the increasing occupation of the rainforest. Previous studies in Coari city showed average infection rates of between 13.3% and 18.9%, with a higher prevalence in rural areas and older patients. Other studies conducted in non-endemic areas show no *M ozzardi* infection. Prevalence was 26.4% in the present study.

Presumed keratitis caused by *M ozzardi* has also been described in Brazil by previous authors, with no other associated aetiology being identified. Comparison with other filarial diseases shows some similarities with onchocerciasis, but onchocerciasis has a different clinical picture and is rare in Brazil: it has different microfilarial tail characteristics and is only found in the Yanomami Indian Reservation close to the Venezuelan border. None of the patients had lived in or visited *Onchoerca* areas.

A study conducted in Pauini on the Purus river identified 20 non-Indian inhabitants with peripheral corneal opacities among 524 patients examined. No microfilariae were seen in the skin snips and *Mansonella* sp. was only found in the blood of the two patients from whom it was possible to collect blood. Another study conducted in São Gabriel da Cachoeira (on the Negro river) showed a positive association between *M ozzardi* and corneal lesions as patients with ocular changes also had *M ozzardi* microfilariae in blood samples. However, all skin biopsies were negative for microfilaria including *Onchoerca*. Cohen et al also reported a positive relationship between keratitis and microfilaremia in the Amazon. This association was also found to be statistically significant in our study, as shown in table 1 (p=0.0001).

Corneal lesions were classified as either corneal opacities or keratitis, as shown in figure 1. Keratitis was more common among patients positive for microfilaremia, while corneal opacity was more common among patients...
negative for microfilaria, but neither finding was statistically significant (table 2, p=0.1347). One explanation for negative microfilaraemia in patients with keratitis could be false negative blood tests, while chronic infection may explain the finding of microfilaraemia in patients with corneal opacity.

As in previous studies, we were unable to detect filariae by biomicroscopy in any ocular tissue, but corneal lesions similar to those previously described were found. Using CCME, we detected lesions which had not been previously reported. Patients positive for microfilaraemia had a higher proportion of confocal lesions (table 3, p=0.4266). Confocal lesions were categorised as linear or circular. More patients with lesions identified by CCME were positive for microfilaraemia and more patients negative for microfilaraemia had circular lesions, although this was not statistically significant (table 3, p=0.4266). This suggests that linear lesions are associated with active disease and circular lesions with inflammatory scars, associated or not with active disease. Linear lesions were larger (250–300 µm) on CCME than in microbiological studies (149–240 µm). The microfilariae seen in ocular tissue are probably dead and could have been enlarged due to an inflammatory reaction. In addition, circular lesions could be cross-sections of the remnants of adult worms (26–49 mm in length by 0.7–0.15 mm in diameter).

Unaffected patients were not tested because of fieldwork difficulties. We carried out confocal microscopy in both eyes of all 22 patients with keratitis. In the 18 with unilateral keratitis, the non-affected eye showed no alterations on confocal microscopy.

New molecular biology techniques for *M. ozzardi* identification could be helpful in future PCR studies of ocular tissue to confirm the association between microfilaraemia and ocular lesions.

### CONCLUSION

We describe *M. ozzardi* microfilariae in the cornea and the associated eye pathology. Further studies using ocular tissue PCR and other imaging techniques would be helpful.

**Acknowledgements** We wish to thank the following members of the Mansonella-Brazil Group: M Borborema, P Cunha, G Fontes, JF Medeiros, FAC Pessoa and T Zanelato. We also thank Nakano EM for helping with confocal microscopy analysis.

**Contributors** LMMV and RBJ participated in the conception and design of the study, and drafted and wrote the manuscript. LMMV, RBJ, JMC, MJC and MM participated in the collection, analysis and interpretation of data, and in the revision and final approval of the manuscript.

**Funding** FAPESP (Fundação de Amparo à Pesquisa do Estado de São Paulo or Foundation for Research Support of São Paulo State) funded this study.

**Competing interests** None.

**Ethics approval** UNIFESP Ethics Committee (number 0767/10) approved this study.

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### Table 3

<table>
<thead>
<tr>
<th>Confocal lesions</th>
<th>Microfilaraemia</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive (+), circular/linear/both</td>
</tr>
<tr>
<td>Microfilaraemia</td>
<td>2 (25%)/1 (12.5%)/5 (62.5%)</td>
</tr>
<tr>
<td>Positive (+)</td>
<td>3 (75%)/0/1 (25%)</td>
</tr>
<tr>
<td>Negative (−)</td>
<td>12</td>
</tr>
</tbody>
</table>

*p=0.4266.
Data sharing statement Further information such as population data and additional images of ophthalmological examination (biomicroscopy and confocal microscopy) can be obtained from the corresponding author.

Provenance and peer review Not commissioned; externally peer reviewed.

REFERENCES