

TRIM59, a novel multiple cancer biomarker for immunohistochemical detection of tumorigenesis

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ABSTRACT

Objectives and design: We identified a novel TRIM59 gene, as an early signal transducer in two (SV40Tag and Ras) oncogene pathways in murine prostate cancer (CaP) models. We explore its clinical applications as a multitumour marker detecting early tumorigenesis by immunohistochemistry (IHC).
Setting and participants: 88 CaP patients were from a tissue microarray (TMA) of radical prostatectomy specimen, 42 patients from a 35 multiple tumour TMA, 75 patients with renal cell carcinoma (RCC) and 92 patients from eight different tumour groups (breast, lung, parotid, gastrointestinal, female genital tract, bladder, kidney and prostate cancer).
Results: TRIM59 upregulation specifically in tumour area was determined by IHC in 291 cases of 37 tumour types. To demonstrate that TRIM59 upregulation is 'tumour-specific', we characterised a significant correlation of TRIM59 IHC signals with tumorigenesis and progression, while in control and normal area, TRIM59 IHC signal was all negative or significantly low. TRIM59 protein upregulation in prostate and kidney cancers was detectable in both intensity and extent in early tumorigenesis of prostate intraepithelial neoplasia ($p<0.05$) and grade 1 of RCC ($p<0.05$), and stopped until high grades cancer. The results of the correlation in these two large cohorts of tumour types confirmed and repeated murine CaP model studies. Enhanced TRIM59 expression was identified in most of the 37 different tumours, while the highest intensities were in lung, breast, liver, skin, tongue and mouth (squamous cell cancer) and endometrial cancers. Multiple tumour upregulation was further confirmed by comparing relative scores of TRIM59 IHC signals in eight tumours with a larger patient population; and by a mouse whole-mount embryo (14.5 days post conception) test on the origin of TRIM59 upregulation in epithelial cells.
Conclusions: TRIM59 may be used a novel multiple tumour marker for immunohistochemical detecting early tumorigenesis and could direct a novel strategy for molecular-targeted diagnosis and therapy of cancer.

INTRODUCTION

The TRIM (*Tripartite Motif*) family is an evolutionarily conserved gene family implicated in a number of critical processes including

ARTICLE SUMMARY

Article focus

- We identified a novel TRIM59 gene, as an early signal transducer in two (SV40Tag and Ras) oncogene pathways in murine prostate cancer (CaP) models.
- This is the first report on the clinical application of TRIM59 as a biomarker for multiple human cancers detecting early tumorigenesis by immunohistochemistry.

Key messages

- We first emphasised that the antibody utilised is TRIM59-specific in this work (also in the sister paper).
- Several TRIM59 antibodies obtained even commercially by immunogens of oligo-peptides may not be detecting the true TRIM59 protein.
- To demonstrate that TRIM59 upregulation is tumor tissue specific in multiple tumor tissues, we have presented sufficient controls, including negative staining in most of normal tissues.
- TRIM59 upregulation specifically in tumor area was determined by immunohistochemistry in 291 cases of 37 tumor types.
- In all, 88 CaP patients were from a tissue microarray (TMA) of radical prostatectomy specimen, 42 patients from a 35 multiple tumor TMA, 75 patients with renal cell carcinoma (RCC) and 92 patients from 8 different tumor groups (breast, lung, parotid, gastrointestinal, female genital tract, bladder, kidney and prostate cancer).
- TRIM59 protein upregulation were detectable in both intensity and extent in early tumorigenesis of PIN (prostate intraepithelial neoplasia, $p<0.05$) and grade 1 of RCC ($p<0.05$), and stopped until high grades cancer.
- Enhanced TRIM59 expression was identified in most of the 37 different tumors, while the highest intensities were in lung, breast, liver, skin, tongue and mouth (squamous cell cancer) and endometrial cancers.
- Multiple tumor upregulation was further confirmed by comparing relative scores of TRIM59 IHC signals in eight tumors with a larger patient population; and by a mouse whole-mount embryo (14.5 days post conception) test on the origin of TRIM59 up regulation in epithelial cells.

STRENGTHS AND LIMITATIONS OF THIS STUDY

- TRIM59 may be used as a novel multiple tumor marker for immunohistochemical detecting early tumorigenesis and could direct a novel strategy for molecular-targeted diagnosis and therapy of cancer.
- The work was mostly on human tumor samples by immunohistochemistry.

immunity,^{1–3} antiviral,^{4–8} proliferation,^{6–9} transcriptional regulation,^{6–10} neuro-development,^{11–12} cell differentiation¹² and cancer¹³ (reviewed in refs^{1–5, 14–15}). However, the function of most TRIM family members is poorly understood and was surmised only based on computational analysis from their RBCC (RING finger, B-box, Coiled-Coil) domains. The domain of RING (Really Interesting New Gene) is frequently involved in proteolysis acting as E3 ubiquitin ligases and the ubiquitin-proteasome system in the regulation of numerous cellular processes including cell cycle regulatory proteins, transcription factors and signal transducers.^{6–15} Recent studies demonstrate that various TRIM-NHL proteins function as cofactor for the microRNA-induced silencing complex.^{12–14, 16} An ataxia-telangiectasia group D complementing gene (ATDC) was recently designated as TRIM29, which is elevated in most invasive pancreatic cancers in the Wnt/ β -catenin signalling pathway.¹³

In a previous report,¹⁷ we characterised the function of TRIM59, a novel TRIM family member, in SV40 Tag oncogene-directed genetically engineered mouse (GEM) prostate cancer (CaP) models.^{18–21} The TRIM59 gene was identified to be correlated with the SV40 Tag-initiated tumorigenesis. TRIM59 protein upregulation and hyperphosphorylation started in the prostate cytoplasm in early tumorigenesis from PIN (prostate intraepithelial neoplasia).¹⁷ As a signal pathway effector, the p-Ser/Thr phosphorylated TRIM59 proteins correlate with tumorigenesis, while p-Tyr-TRIM59 proteins correlate with advanced CaP. The function of TRIM59 was identified by shRNA knockdown in human CaP cells resulted in S-phase arrest and cell growth retardation. Although TRIM59 is an effector gene with the SV40Tag oncogene, the initial functional targets of TRIM59 function were actually on the Ras signal pathway as an early and rapid signal transmitter (for a review see ref.²²). In a transgenic mouse test of TRIM59 upregulation specifically in the prostate, TRIM59 demonstrated full oncogenic activity in directing tumorigenesis and progression to highest-grade CaP.^{17–23} The signal pathway of TRIM59 may be possibly linked with two large oncogene routes: the Ras/Raf/MEK/ERK/PI3K/AKT and the SV40 Tag/p53/pRB routes.¹⁷ In this study, we explored the clinical utility of the novel TRIM59 gene to serve as a biomarker for a panel of human cancers. To address the molecular biology basis, we combined results from both basic (animal model^{17–21}) and human clinical studies.

MATERIALS AND METHODS

Patient selection

All patient samples were acquired as part of REB (Research Ethics Board) approved protocols at the Western University (UWO) and Vancouver Prostate Center, University of British Columbia (UBC). Table 1 shows a complete list of 291 patients with 37 different tumour types examined in this study.

Prostate cancer tissue microarrays

Eighty-eight CaP patients between 2006 and 2008 who had no treatment prior to radical prostatectomy were selected from the Vancouver General Hospital, UBC. Each patient block marked as containing benign tissue or cancer was sampled two times with a core diameter of 1 mm arrayed (176 cores) in a rectangular pattern with 0.7 mm between the centres of each core, creating a duplicate tissue microarray (TMA) layout using a manual tissue micro arrayer (Beecher Instruments, Silver Spring, Maryland, USA). The TMA paraffin blocks, were sectioned into 0.5 μ m sections and mounted on the positively charged slides.

Automated image, acquisition and analysis on immunohistochemical staining of CaP-TMA

Immunohistochemical staining was conducted by Ventana autostainer model Discover XT (Ventana Medical System, Tucson, Arizona, USA) with enzyme-labelled biotin streptavidin system and solvent-resistant DAB Map kit (Ventana Medical System). TMA was scanned by Bliss Digital imaging system using $\times 20$ objective (Bacus Laboratories Inc., Centre Valley, Pennsylvania, USA) and stored in the Prostate Centre Saver (<http://bliss.prostatecentre.com>). A value on a four-point scale assigned to each core.

Multiple tumour TMA construction

Tissue samples from 42 patients that encompassed 35 distinct tumour subtypes were selected from London Laboratory Service Group, and the tumour bank in the Department of Pathology (UWO). TMA slides were constructed with triplicate cores for each sample following standard procedure as described.²⁴ Sections of 0.6 mm were prepared from TMA block and re-stained by H&E for each case to confirm the diagnosis.

Histopathological analysis

All cases from 37 tumour types were graded according to standardized histopathology grading systems²⁵ by MM (pathologist) and VK (MD fellow).

Immunohistochemistry and results evaluation

Standard ABC (Avidin Biotin Complex) protocol was performed as previously reported.^{21–26–29} Two kinds of blocking reagents were used: Power Block (Universal Blocking Reagent; BioGenex, San Ramon, California, USA) and Avidin-biotin blocking reagent kit (Vector Labs, Burlingame, California, USA). All B-Raf antibodies were from GenScript (Piscataway, New Jersey, USA): B-Raf

Table 1 Patient list selected in this study

Organ	Patient number	Tumour type	Tumour grade
35 Tumour-TMA	42	Description in table 2	
Prostate TMA	105	Adenocarcinoma	BPH 16 PIN 4 Gleason score 4 5 Gleason score 6 31 Gleason 7 15 Gleason score 8 16 Gleason score 9–10 9 Stroma 3 Absent cores 6
Kidney	75	Clear cell carcinoma 43 Papillary RCC 11 Chromophobe RCC 13 Cystic RCC 6 Sarcomatoid RCC 2	Grade 1 4 Grade 2 38 Grade 3 28 Grade 4 5
Bladder	44	Urothelial carcinoma	Low grade 38 High grade 6
Lung	4	Bronchoalveolar carcinoma 1 Adenocarcinoma 1 Large cell carcinoma 1 Squamous cell carcinoma 1	Grade 1 1 Grade 2 1 Grade 3 1 Grade 4 1
Breast	3	Invasive lobular carcinoma 1 Invasive mammary carcinoma 2	Grade 1 1 Grade 3 2
Female genital tract	5	Endometrial carcinoma 4 Ovary, endometrioid carcinoma 1	Grade 1 4 Grade 2 1
Gastrointestinal tract	2	Colon carcinoma 1 Pancreas neuroendocrine carcinoma 1	Low grade 1 High grade 1
Parotid	3	Mucoepidermoid carcinoma 1 Metastatic SCC 1 Metastatic neuroendocrine carcinoma 1	Low grade 1 High grade 2
Mouth, tongue and larynx	4	Squamous cell carcinoma 4	Moderately differentiated 3 Moderately to poorly differentiated 1
Total	291		

RCC, renal cell carcinoma; SCC, squamous cell carcinoma; TMA, tissue microarray.

antibody (monoclonal antibody, used 1:500 dilution), Raf 1 (polyclonal, 1:50) and Raf -1 (polyclonal, Phospho-Ser²⁵⁹ 1:50).

TRIM59 IHC staining signals were assessed by intensity for cytoplasmic staining and extent to assess the percentage of nuclear staining as previously reported.^{28 29} Since in some tumours TRIM59 showed only cytoplasmic staining, for the purpose of comparing in different tumours, we used a combined relative score system based on both intensity and extent as follows: score 0: 0/0 (intensity/extent); score 1: weak cytoplasmic staining and/or ≤25% nuclear staining; score 2: moderate cytoplasmic staining and/or ≤50% nuclear staining; score 3: strong cytoplasmic staining and/or ≥50% nuclear staining. All relative scores were accessed by two researchers independently MM (pathologist) and VK (MD fellow).

Construction and characterisation of TRIM59 antibody

Online supplementary figure S1 provided details, which combined online supplementary materials of,¹⁷ and more informations. Online supplementary figure S1A

showed that the antibody TRIM59#72 used for mouse and human studies was prepared from C-terminal 136-amino acid peptide (as recombinant GST-fusion protein, see online supplementary figure S1B) of whole-protein 403aa which is TRIM59 specific, while antibody TRIM59#71 was from mouse TRIM59 N-terminal sequence (163aa) containing mostly the common RBCC function domains of more than 70 members the TRIM family.¹⁷ As shown in online supplementary figure S1C, western blots on both mouse and human cell lysates, #72 recognised only one band, while #71 showed multiple bands. We also demonstrated that N-terminal sequence-specific antibody TRIM59#71 can recognise purified proteins from C-terminal sequence-specific, TRIM59#72 Ab affinity column (the same 53 kDa protein, see online supplementary figure S1D). Since these two immunogenic sequences are not overlapped, we thus verified that they are detecting the same protein. We demonstrated that TRIM59#72 can be used for human tissues for immunohistochemistry (IHC) and western blots (see online supplementary figure S1E).

Table 2 Immunohistochemistry (IHC) analysis of TRIM59 as multiple marker in 35 tumour tissue microarray

Tumour type	Patient number	Core number	Pathological grade	Cell type	IHC staining	
					Cytoplasm staining (intensity)	Nuclear staining (extent)
Renal clear cell carcinoma,	2	6	2 3	Epithelial	moderate moderate	– –
Adrenal gland cortical carcinoma	1	3	N/A	Epithelial	Moderate–strong	–
Squamous cell carcinoma (SCC), skin	2	6	WD	Epithelial	Strong	–
Basal cell carcinoma, skin	2	6	MD N/A	Epithelial	Moderate Moderate–strong	– –
Melanoma	1	3	N/A	Epithelial	Weak	50%+
Endometrioid adenocarcinoma	2	6	2 1	Epithelial	Moderate–strong Moderate–strong	– –
Leiomyosarcoma	1	3	N/A	Mesenchymal	Weak	–
Omentum serous adenocarcinoma,	1	3	WD	Epithelial	Weak–moderate	–
Ovary serous adenocarcinoma	1	3	N/A	Epithelial	Weak–moderate	–
Ovary clear cell carcinoma	1	3	PD	Epithelial	Moderate–strong	30%+
Cervix adenocarcinoma	1	3	WD-MD	Epithelial	Moderate–strong	–
Colon adenocarcinoma	1	3	Low grade	Epithelial	Weak	–
Breast ductal adenocarcinoma	1	3	2/3	Epithelial	Moderate–strong	50%+
Bladder urothelial carcinoma	2	6	Low grade 2 (low grade)	Epithelial	Weak Moderate	– –
Stomach GIST	1	3		Epithelial	Weak–moderate	–
Esophagus adenocarcinoma	1	3	MD	Epithelial	0–Weak	–
Thyroid, papillary carcinoma	1	3	N/A	Epithelial	Weak	–
Thyroid, medullary carcinoma	1	3	N/A	Epithelial	Moderate	–
Pancreas adenocarcinoma	2	6	2	Epithelial	Weak–moderate Moderate–strong	– –
Pancreas endocrine tumour	1	3	N/A	Epithelial	Strong	–
Lung SCC	1	3	PD	Epithelial	Strong	–
Lung mesothelioma	1	3	MD-PD		Strong	20%+
Lung adenocarcinoma	1	3	MD		Moderate	50%+
Lung bronchoalveolar carcinoma	1	3	WD		Moderate–strong	50%+
Lung mesothelioma, biphasic	1	3	MD-PD		Strong	20%+
Liver hepatocellular carcinoma (HCCa)	1	3	2/4	Epithelial	Strong	–
Liver metastatic carcinoid	1	3	N/A	Epithelial	Strong	–
Small bowel marginal zone lymphoma	1	3	N/A	Lymphocyte	0–Weak	–
Lymph node, follicular lymphoma	1	3	1/3	Lymphocyte	0–Weak	–
Lymph node, metastatic carcinoid	1	3	Low grade	Epithelial	Strong	–
Spleen, Hodgkin’s lymphoma	1	3	N/A	Lymphocyte	Weak	–
Stomach, malt lymphoma	1	3	Low grade	Lymphocyte	0–Weak	–
Thymus invasive thymoma	1	3	N/A	Epithelial	0–Wweak	–
Appendix, goblet cell carcinoid	1	3	N/A	Epithelial	0	–

GIST, gastrointestinal stromal tumour.

Confocal microscope imaging of immunostaining of mouse embryos 14.5days post conception for TRIM59

The mouse embryos were prepared according to our previous reports.^{19 20} The anti-rabbit secondary antibody was conjugated to Cy3 (1:200, Jackson Immuno Research, West Grove, Pennsylvania, USA). Sections were double-stained with pan-cytokeratin antibody with FICT (Sigma, St. Louis, Missouri, USA), and the stained mouse embryo

sections were visualised using a Carl Zeiss (Oberkochen, Germany) confocal microscope by the LSM Image program.

Statistical analysis

Student’s t tests and one-way analysis of variance were used by programs of Microsoft Excel 2007 or SPSS 10 to

analyse the data with $p < 0.05$ considered to be statistically significant.

RESULTS AND DISCUSSION

TRIM59 upregulation in human prostate cancer TMA: correlation with tumorigenesis and tumor progression by TRIM59 intensity until high grade CaP

We designated TRIM59 as one of the ‘tumorigenesis-associated’ genes correlated with SV40 Tag oncogenesis in mouse prostate cancer (GEM-CaP) models.¹⁷ SV40 Tag is essentially only required for the initiation of tumorigenesis, that is, the ‘hit-and-run’ effect, in GEM-CaP, but not for the tumour progression and metastasis directly. The ‘tumorigenesis-associated’ effect is due to the initial binding of the Tag oncogene with retinoblastoma (pRB), p53 proteins and several transcriptional coactivators. Once this process is initiated, the signal transduction will continue on, even without the initiation effectors.

In GEM-CaP models, the TRIM59 protein upregulation correlated with tumorigenesis and progression, and downregulated in the high-grade CaP by IHC.¹⁷ We assume this ‘tumorigenesis-associated’ effect of TRIM59 may apply to human cancer studies.

We first characterised that TRIM59 antibody (#72) can cross-react with and recognise specifically human TRIM59 counterpart (for details see online supplementary figure S1).

We tested by IHC in a TMA of CaP patients ($n=88$, 176 cores). TRIM59 IHC signals detected in an automated digital image system were identified mostly in cytoplasm of luminal cells (figure 1A), which is different from rapid tumour progression mouse CaP models. The intensity (score=2) in PIN ($n=4$, figure 1A) was higher than in non-tumour area (normal and benign prostatic hyperplasia). Moderate-to-strong expression was observed in Gleason score 6 (3+3, $n=25$), 7 (4+3, $n=15$) and 8 (4+4, $n=14$). In high-grade CaP (score 4+5, $n=8$, figure 1A), TRIM59-IHC signals were lower. As shown in a graph of figure 1B, TRIM59 protein IHC signals correlated significantly ($p=0.014$) with tumorigenesis and progression from PIN to WDCaP (Gleason grade 1–2, scores 2–4) and MDCaP (Gleason grade 2–4, scores 4–8) (graph figure 1B), and decreased in high-grade CaP (Gleason score 9–10) with $p=0.018$, which is similar to GEMs.¹⁷

TRIM59 upregulation in human renal cell carcinoma patients: correlation of tumorigenesis and tumour progression with TRIM59 intensity until high-grade RCC

Next, we demonstrated that TRIM59 is not androgen-responsive and likely not prostate tissue-specific, indicating that the tumorigenesis role of TRIM59 may be applied to all or most of human cancers. Online supplementary figure S2 showed this result in detail of IHC

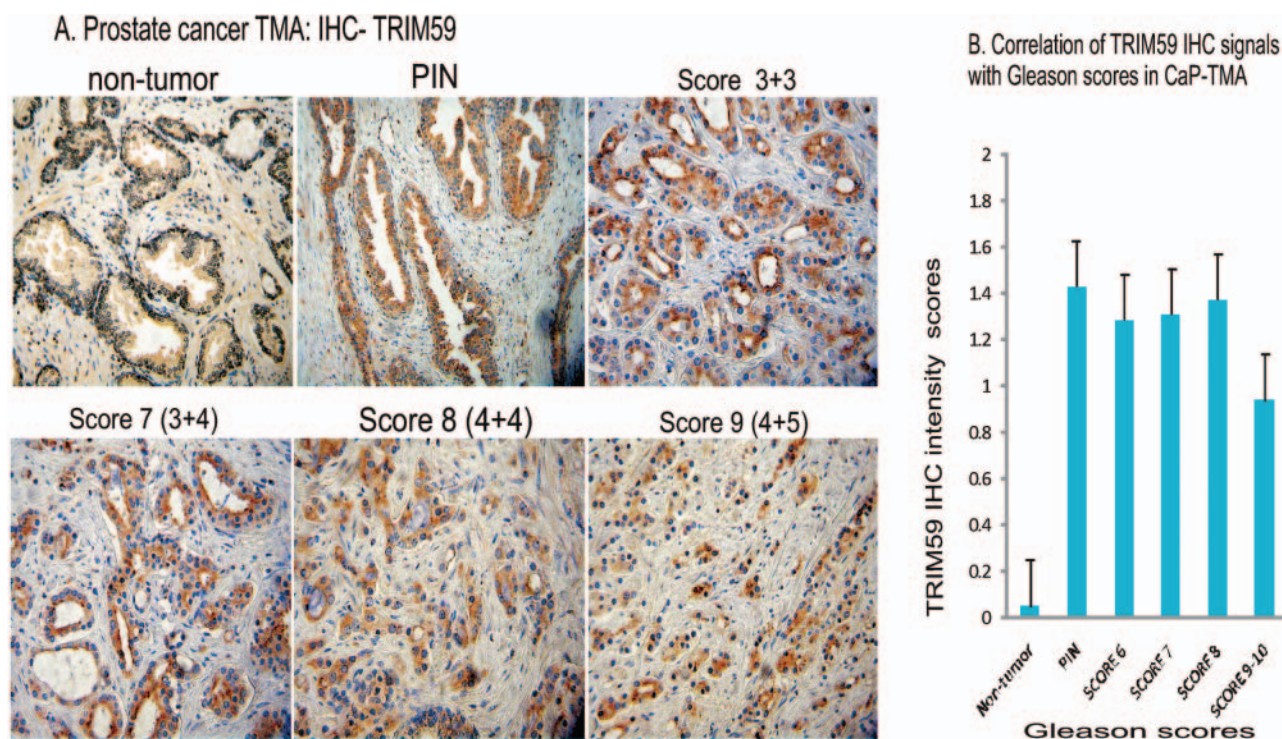


Figure 1 Correlation of TRIM59 immunohistochemistry (IHC) staining in prostate cancer cases in tissue microarray (TMA) assessed by the Gleason grading system. All panels in (A) were shown haematoxylin staining, $\times 20$. Non-tumour: weak or negative, prostate intraepithelial neoplasia (PIN): TRIM59 is located in the cytoplasm of the luminal cells (intensity = 2). Gleason score 3 + 3, TRIM59 is located in the cytoplasm of tumour cells (intensity = 2). Gleason score 3 + 4, TRIM59 is located in the cytoplasm of the tumour cells (intensity = 2). Gleason score 4 + 4, TRIM59 is located in the cytoplasm of the tumour cells (intensity = 2). Gleason score 4 + 5, TRIM59 is located in the cytoplasm of the tumour cells (intensity < 1). Graph B: correlation of TRIM59 protein levels by intensity with Gleason score in prostate cancer TMA analysis. Error bars show mean (\pm SE).

test by TRIM59 antibody#72 on a TMA of the LNCaP human CaP xenografts in nude/severe combined immuno deficiency castrated mice.

We extended results from prostate cancer clinical samples to kidney cancer. We started 75 renal cell carcinoma (RCC) patients including all 5 different types of RCC tumours: 43 clear cell carcinoma (representative IHC figures shown in figure 2A–D), 11 papillary RCC (figure 2E,F), 13 chromophobe RCC (figure 2G–I), 2 sarcomatoid RCC (figure 2J), and 6 cystic RCC (figure 2K,L). RCC cases analysed with Fuhrman grade 1–4 were 4, 38, 28 and 5, respectively. TRIM59-IHC staining in normal area including proximal tubules was negative (figure 2, last row). Background staining was eliminated by testing antibody dilutions (1 : 100,

200, 500, 1200 till 5000) while tumour-specific signals of TRIM59 proteins were noticeable. Endogenous biotin signals were blocked and excluded by additional block reagents (avidin-biotin blocking reagent kit).

TRIM59 IHC staining in tumour areas in RCC (figure 2) was different from cases of CaP-TMA (mainly cytoplasmic). TRIM59 IHC staining was found in both cytoplasm and nucleus in all RCC grades and types (figure 2). We assessed TRIM59-IHC by visual scoring of both intensity (cytoplasmic staining) and extent (% nucleus staining) microscopically. Correlation of TRIM59 IHC signals by scoring the intensity in cytoplasm with grades of all five types of RCC is shown in figure 2M. TRIM59 IHC signals were increased with tumour progression from grade 1–3

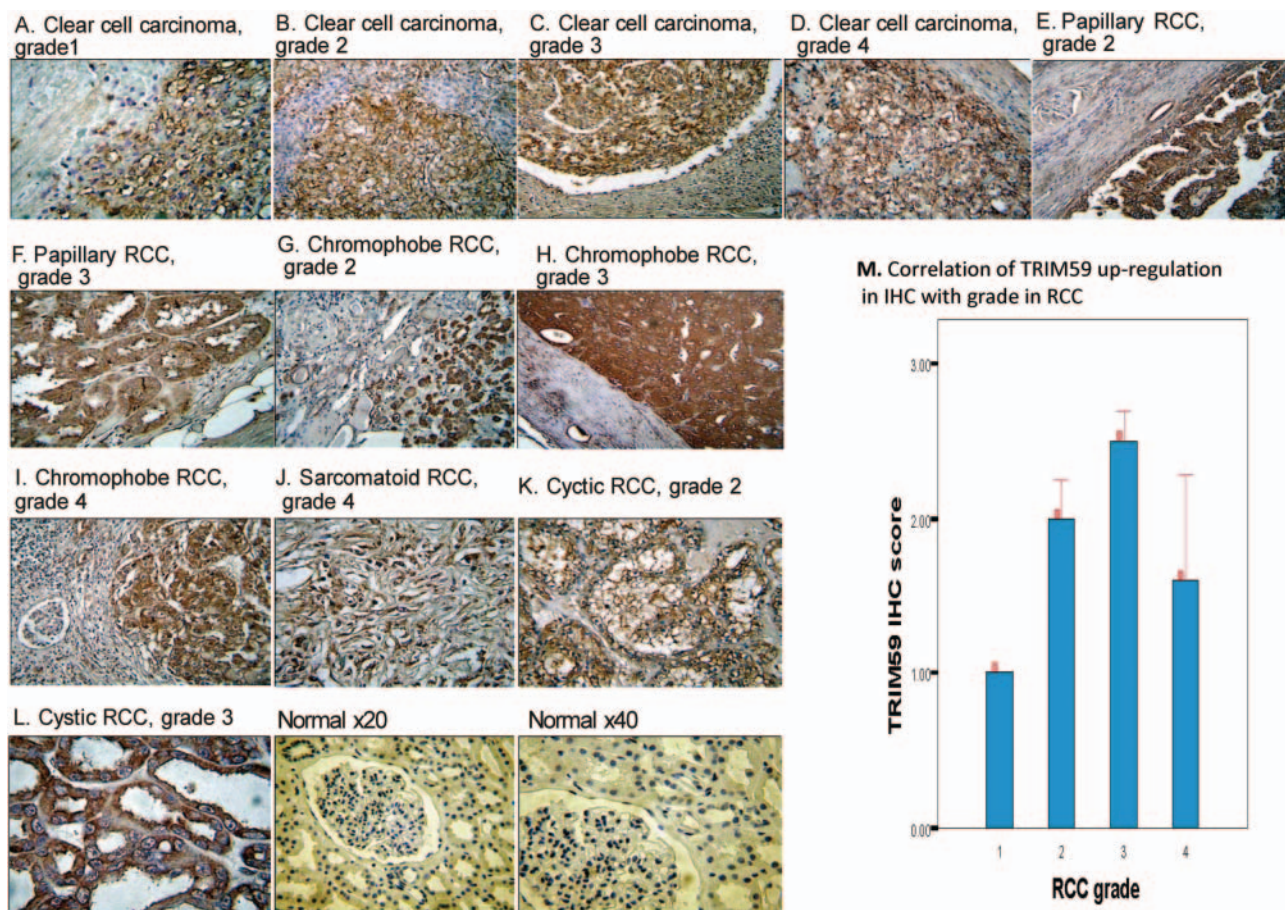


Figure 2 Immunohistochemistry analysis of TRIM59 expression in kidney cancer (RCC, renal cell carcinoma) cases: correlation with tumour grade by intensity detecting early tumorigenesis. Five types of RCC with different grades were shown clear cell carcinoma (A–D), papillary RCC (E and F), chromophobe RCC (G–I), sarcomatoid RCC (J) and cystic RCC (K and L). (A) Clear cell carcinoma, grade 1, weak cytoplasmic staining in tumour cells, $\geq 50\%$ nuclear staining, cytoplasmic staining in tumour cells ($\times 40$). (B) Clear cell carcinoma, grade 2, moderate cytoplasmic staining in tumour cells, no nuclear staining ($\times 40$). (C) Clear cell carcinoma, grade 3, strong cytoplasmic staining in tumour cells, no nuclear staining ($\times 40$). (D) Clear cell carcinoma, grade 4, moderate cytoplasmic staining and nuclear (%) staining ($\times 40$). (E) Papillary RCC, grade 2, strong cytoplasmic staining in tumour cells, no nuclear staining ($\times 40$). (F) Papillary RCC, grade 3, strong cytoplasmic staining in tumour cells, no nuclear staining, normal cortex tissue is visible adjacent to tumour area ($\times 40$). (G) Chromophobe RCC, grade 2, strong cytoplasmic staining in tumour cells, no nuclear staining. ($\times 40$). (H) Chromophobe RCC, grade 3, strong cytoplasmic staining, no nuclear staining, ($\times 40$). (I) Chromophobe RCC, grade 4, moderate cytoplasm staining and nuclear staining ($\times 40$). (J) Sarcomatoid RCC, grade 4, weak cytoplasmic staining and nuclear staining ($\times 40$). (K) Cystic RCC, grade 2, moderate cytoplasmic staining, no nuclear staining ($\times 40$). (L) Cystic RCC, grade 3, strong cytoplasmic staining, no nuclear staining ($\times 40$). Last two panels: normal kidney tissues ($\times 20$, $\times 40$). (M) Graph: correlation of TRIM59 protein levels by relative scores (both intensity and extent) with grade in RCC according to the Fuhrman nuclear grading system. Error bars show mean (\pm SE).

($p < 0.05$). All grade 1 tumours ($n=4$) stained with weak TRIM59 IHC signals in cytoplasm, but with high extent of nuclear staining; while all grade 2 and 3 tumours ($n=66$) showed moderate-to-strong cytoplasmic staining intensity of TRIM59. All grade 4 tumours ($n=5$) showed weak-to-moderate intensity in cytoplasm of TRIM59 staining. No correlation between TRIM59 IHC staining in nucleus and tumour grade was found, although low-grade RCC showed higher nuclear staining.

Therefore, by systematic IHC studies in CaP (88 patients, figure 1) and kidney cancer (75 patients, figure 2), we almost exactly repeated results from our mouse model studies on TRIM59.¹⁷ We confirmed TRIM59 as an IHC marker able to detect low-grade tumour in early tumorigenesis. We also demonstrated the correlation of tumorigenesis and tumour progression with TRIM59 upregulation until high-grade tumour.

TMA analysis of TRIM59 protein expression demonstrates that TRIM59 is a multiple tumour marker

In the basic research previously using animal GEM-CaP models,¹⁷ we have disclosed that TRIM59 upregulation is involved in two oncogene families and two signal pathways of SV40Tag/pRB/p53 and Ras/Raf/MEK/ERK. TRIM59 may function as an early signal transducer in Ras signal pathway with bridging genes in two oncogene pathways.¹⁷

While it was rarely reported that SV40 Tag oncogene induced human cancer, Ras mutations are among the most frequent alterations in human cancers (for a review see ref. ²²). We assume that TRIM59 as an early Ras signal pathway effector may possibly act as a multiple tumour marker.

We therefore further extended TRIM59 IHC studies to 35 multiple cancer TMA sections (42 tumours, 126 cores, table 1). We tested different dilutions (1/300, 1/600, 1/1200 and 1/5000) of TRIM59 antibody (see online supplementary figure S3). To further confirm the specificity and reliability of TRIM59 antibody in IHC staining, we compared IHC staining in 35 different tumour-TMA sections with positive (TRIM59 antibody at 1:1200 and 1:5000 dilutions) and negative controls (no antibody added, for details see online supplementary figure S4). As summarised in table 2, TRIM59 expression was significant and tissue-specifically upregulated in most of these 35 tumours. When comparing the relative scores (both intensity and extent) in different tumours, the highest staining was observed in breast, lung, liver, skin (squamous cell carcinoma) and endometrial cancers.

Further confirmation of TRIM59 as a tumour marker in patients with eight different tumours

Since the 35 tumour-TMA contained only limited cases in each tumour type, we selected more cases ($n=92$) of eight different tumour types with different tumour grades, which all showed upregulated expression of TRIM59. IHC staining of TRIM59 in eight tumours are shown: lung ($n=4$, figure 3A–C), breast ($n=3$, figure 3D,E),

gastrointestinal ($n=2$, figure 3F,G), female genital tract ($n=5$, figure 3H–J), bladder ($n=44$, figure 3K), prostate ($n=27$ from UWO, figure 3L), head and neck mucosal tumour (squamous cell carcinoma, SCC of mouth, tongue and larynx, $n=4$, figure 3M–O) and parotid gland ($n=3$, figure 3P,Q). Normal areas in lung, breast, colon, endometrial, bladder, larynx and parotid tissues showed very weak or completely negative staining (figure 3).

Since some tumours (eg, prostate) showed mostly cytoplasmic and no nuclear TRIM59-IHC staining, as a comparative study, we assessed their relative scores (combine both intensity and extent scores, see Materials and methods). More tumours from kidney (RCC, $n=75$) and prostate cancer ($n=27$) were included as references and all were assessed by relative scores simultaneously, since we already analysed a large cohort of these patients. Figure 3R shows the comparison of the mean of IHC-TRIM59 relative scores. The highest relative scores were found in SCC of the parotid, mouth, larynx and tongue, followed by lung, breast and female genital tract cancers.

The comparison of relative scores on low and high grades separately was done (data not shown). Cases of grade 1 lung cancer (bronchoalveolar, adenocarcinoma, squamous cell carcinoma (SCC) and large cell carcinoma) and breast cancer (invasive lobular and invasive mammary carcinoma) all showed the strongest staining as compared with other tumours. In endometrial cancer, the TRIM59 relative scores were moderate in grade 1 and moderate to strong in grade 2. The three tumours of SCC from mouth, tongue and larynx with different grades (figure 3M–O) also showed high relative scores (both intensity and extent).

As a comparative study (figure 3R, table 2), we tested 44 bladder cancer cases with 38 low-grade and 6 high-grade tumours. The mean value of relative scores was 1.6, that is, weak to moderate in bladder cancer cases. In 27 prostate cancer cases (from UWO only) tested, the relative scores of TRIM59 (cytoplasmic staining only) from PIN through Gleason scores of 10 were actually relatively weak (figure 3R), although in Gleason score 4, 6 and 8 were weak to moderate separately.

So far, we identified that TRIM59 upregulation is ‘tumour-specific’. First, we demonstrated the correlation of TRIM59-enhanced IHC signals with tumorigenesis and progression, which were statistically significant in this report with 291 cases and 37 tumour type analyses. Second, although TRIM59 is a normal gene involved in CDC (cell cycle division) regulation from G1 to S-phase and involved in DNA S-phase and cell growth,¹⁷ we demonstrated that in normal or non-tumour areas in all tested 37 different kinds of cancers, TRIM59 IHC staining signals were mostly negative or very low (figures 1–3). By moderating antibody dilutions and testing various blocking reagents (see online supplementary figure S4), we demonstrated that TRIM59 induces tumorigenesis/oncogenesis only when it is abnormally upregulated.

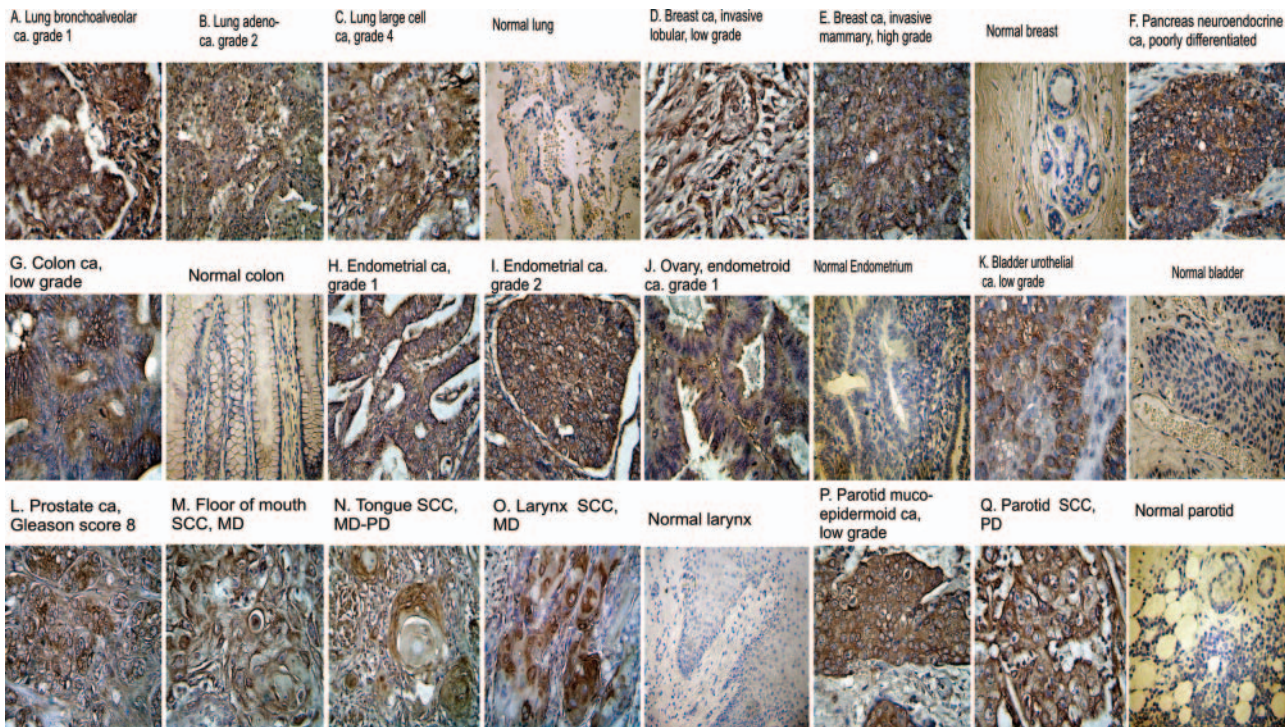


Figure 3 Comparison of TRIM59 expression as a multiple-cancer marker in eight types of tumours in breast, lung, parotid, gastrointestinal, female genital tract, bladder, head and neck mucosal tumour and prostate cancer. Negative TRIM59 staining in normal tissues was shown for each tumour, respectively (×20). (A) Lung bronchoalveolar carcinoma, grade 1, strong cytoplasmic and nuclear staining of tumour cells, (×40). (B) Lung adenocarcinoma, grade 2, moderate cytoplasmic staining of tumour cells, no nuclear staining (×40). (C) Lung large cell carcinoma, grade 4, moderate cytoplasmic and nuclear staining of tumour cells (×40). (D) Breast cancer, invasive lobular, low-grade, moderate to strong cytoplasmic and nuclear staining of tumour cells (×40). (E) Breast cancer, invasive mammary, high-grade, moderate cytoplasmic staining of tumour cells, no nuclear staining (×40). (F) Pancreas neuroendocrine carcinoma, poorly differentiated, strong cytoplasmic staining, no nuclear staining of tumour cells (×40). (G) Colon carcinoma, low-grade, weak-to-moderate cytoplasmic staining of tumour cells, no nuclear staining (×40). (H) Endometrial carcinoma, grade 1, weak-to-moderate cytoplasmic staining of tumour cells, no nuclear staining (×40). (I) Endometrial carcinoma, grade 2, moderate-to-strong cytoplasmic staining, no nuclear staining (×40). (J) Ovary, endometrioid carcinoma, grade 1, weak cytoplasmic staining, no nuclear staining (×40). (K) Bladder urothelial carcinoma, low-grade, moderate cytoplasmic staining, no nuclear staining (×40). (L) Prostate adenocarcinoma, Gleason score 8, moderate cytoplasmic staining, no nuclear staining (40×). (M) Floor of mouth, SCC moderately differentiated, strong cytoplasmic staining, no nuclear staining (40×). (N) Tongue cancer, SCC moderately to poorly differentiated, strong cytoplasmic staining, no nuclear staining (40×). (O) Larynx, SCC moderately differentiated, strong cytoplasmic staining, no nuclear staining (40×). (P) Parotid, mucoepidermoid carcinoma, low-grade, strong cytoplasmic staining, no nuclear staining (×40). (Q) Parotid, metastatic SCC, poorly differentiated, moderate-to-strong cytoplasmic staining, no nuclear staining (×40). (R) Graphic comparison of relative scores (both intensity and extent) of TRIM59 IHC signals in eight different tumour types. fGT, female genital tract. Error bars show means (±SE) of the relative scores in all grades analysed and compared.

In most of human cancers tested in this clinical IHC study, strong TRIM59 expression in tumour were identified in epithelial cancers: lung, breast, skin, which all associated with epithelium originations (very rare from mesenchymal tumours, for details see table 2).

Furthermore, we also confirmed that TRIM59 expression involved in multiple tissue expression even in embryo development. We carried out IHC of mouse embryo sections by double-staining TRIM59. Cytokeratin (keratin), a family of proteins that are primarily found in epithelial cells was used as reference. Online supplementary figure S5 illustrated confocal microscope images of IHC staining of TRIM59 in different organs/tissues of mouse embryo (14.5 days postconception). TRIM59 was highly expressed in

cytokeratin-expressing cells in the lung (first row), skin (second row), and kidney (not shown) of mouse embryos. TRIM59 staining in mouse embryos revealed the same pattern of the epithelium origin as in human tumours, which the TRIM59 gene were found highly upregulated in those tumour types as well (see table 2).

Given our previous experiments suggesting TRIM59 functions in the Ras pathway, we tested if TRIM59 upregulation was correlated with the BRAF, an early signal effector Ras signal pathway (for a review see ref. ²²). We selected 24 RCC patients, which previously were confirmed with upregulation of TRIM59 expression. Three antibodies were used: B-Raf Antibody (mAb) and Raf-1 Antibody (Ab-259) testing the total Raf protein and

R. Comparison of relative scores of TRIM59 IHC staining in eight different tumor types

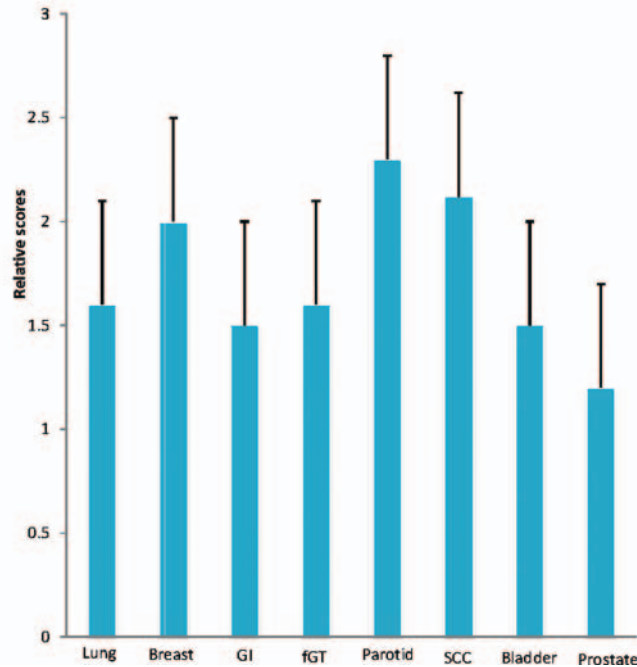


Figure 3 Continued.

Raf-1 Antibody (Phospho-Ser²⁵⁹ pAb) testing the activated phosphorylated B-Raf. Some of the serial slides were stained in parallel on each patient by different antibodies. As shown in online supplementary figure S6A first two columns, in all 12 clear cell carcinoma of RCC samples, there were no or very weak B-Raf IHC signals in all three used Raf antibodies. In papillary RCC (see online supplementary figure S6B) and chromophobe RCC samples (see online supplementary figure S6C) (15 samples of 24 samples or 62% of all RCC samples tested), there were higher IHC signals in all three antibodies used for staining in cancer areas specifically, showing higher intensity and extent than clear cell carcinoma (see online supplementary figure S6A). Nuclear signals were found only by B-Raf P-Ser antibody in papillary and chromophobe tumours. Online supplementary table S1 summarises the results. It is intriguing that in those TRIM59 upregulated kidney cancers (RCC), neither total nor phosphorylated BRaf were detected in clear RCC (as a control), but were all highly positive in other two RCCs (papillary and chromophobe) tumours. We could not definitely confirm that TRIM59 was acting along the Ras pathway in all cases where it was detected.

CONCLUSIONS

This is the first report on a possible 'ubiquitous' tumour marker. Ras mutations are among the most frequent alterations in human cancers that lead to approximately 30% of all human cancers with expression of

constitutively active Ras proteins (for a review see refs. ²²⁻³⁰). In this report, we have demonstrated that there are more than mutation issues of the Ras signal pathway in tumorigenesis and progression, since TRIM59 is upregulated as a novel proto-oncogene in a variety of human cancers.

Detection and treatment of cancer at the earliest stage are critical for patient survival. This investigation demonstrated a novel TRIM59 gene as a multiple tumour marker for early diagnosis of tumorigenesis. As a multiple biomarker associated with epithelium origin (very rarely arisen from mesenchymal tissue, see table 2) TRIM59 may be used as an EMT (epithelium–mesenchymal transition) specific cobiomarker. The TRIM59 antibody may also be used for molecular-targeted imaging as a new diagnostic marker, for example, in the context of targeted microbubble ultrasound destruction technology,²³ before the protocol of a serum TRIM59 test can be established.

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Competing interests None.

Provenance and peer review Not commissioned; externally peer reviewed.

Data Sharing Statement Extra data are available by emailing correspondent author JWX.

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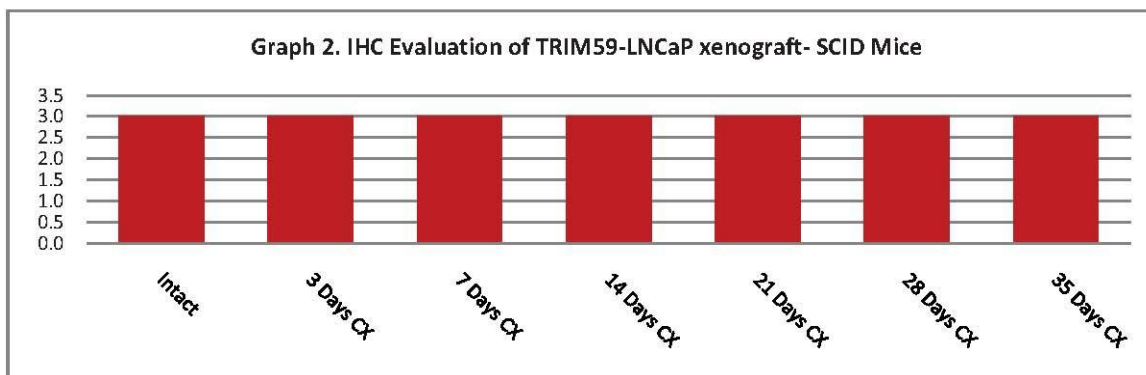
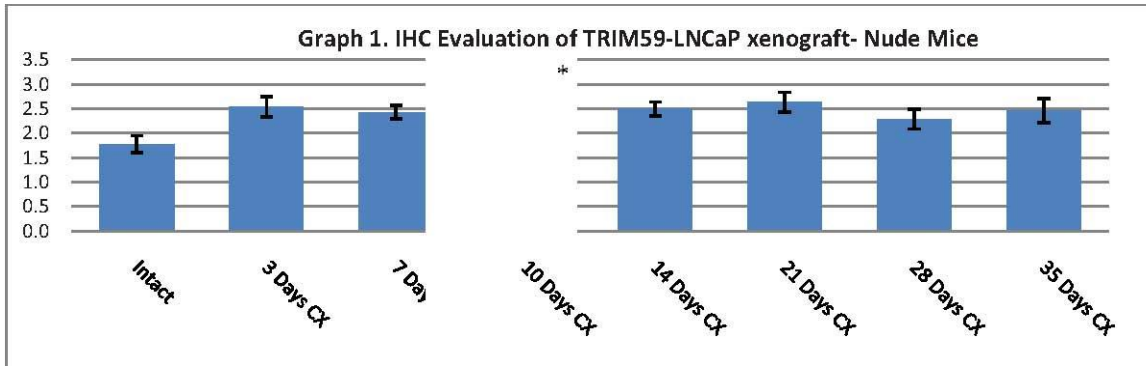
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Supplemental Fig.2 IHC evaluation of androgen responsiveness of TRIM59 in the LNCaP xenograft TMA.

The LNCaP xenograft TMA (tissue microarray) were prepared from xenografts generated by an androgen-dependent human prostate cancer cell line LNCaP in nude SCID (Severe Combined Immuno Deficiency) mice. The TMA is a tumor sample array comprising samples of time points (from day 1 to day 35) of the xenografts taken before and after castration (CX) of the implanted LNCaP tumors in nude (SCID) mice.

Summary of results: In the nude mice implanted with xenograft induced by androgen-dependent human prostate cancer cell line LNCaP, TRIM59 is significantly up-regulated after 10 days castration. After 35 days after the castration resistance stage, TRIM59 expression does not up-regulate but not significantly (shown in graph 1, Fig. A). Similarly, in the SCID mice TRIM59 shows strong immunoreactivity but does not change in all time points of castration (shown in graph 1, Fig. B).

Conclusion: TRIM59 is not androgen-responsive in castration test in TMA of human prostate cancer xenograft in nude and SCID mice test.



Suppl Fig. 2

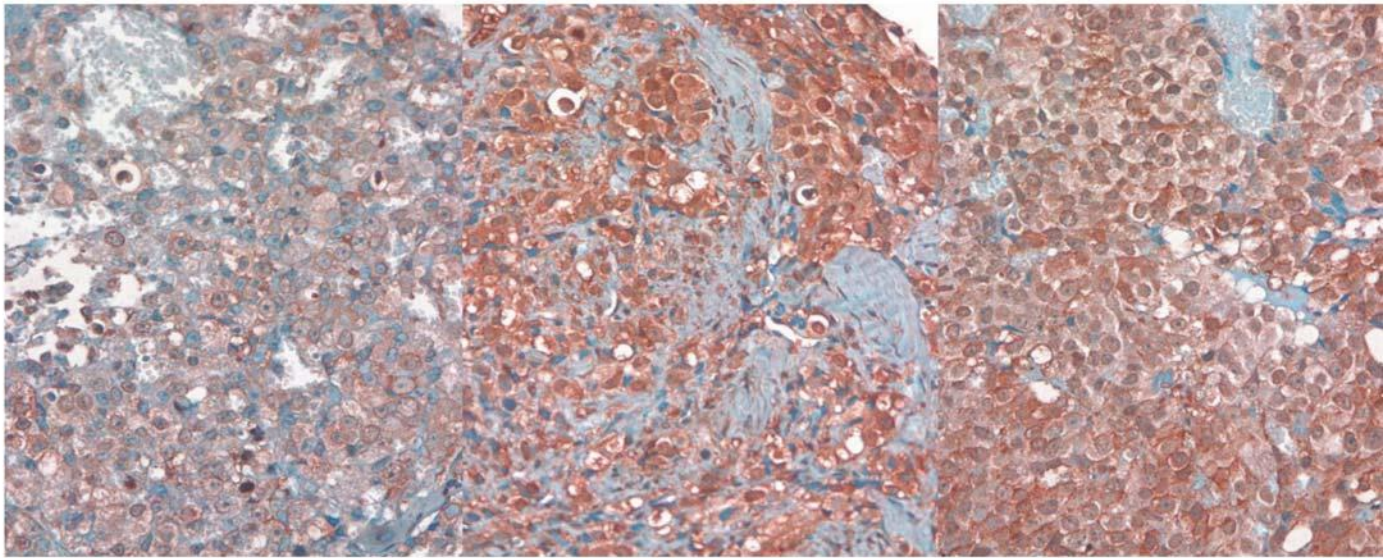
TRIM59 SCID-LNCaP xenograft TMA

A Day after castration in nude mice

No castration

10 days

35 days

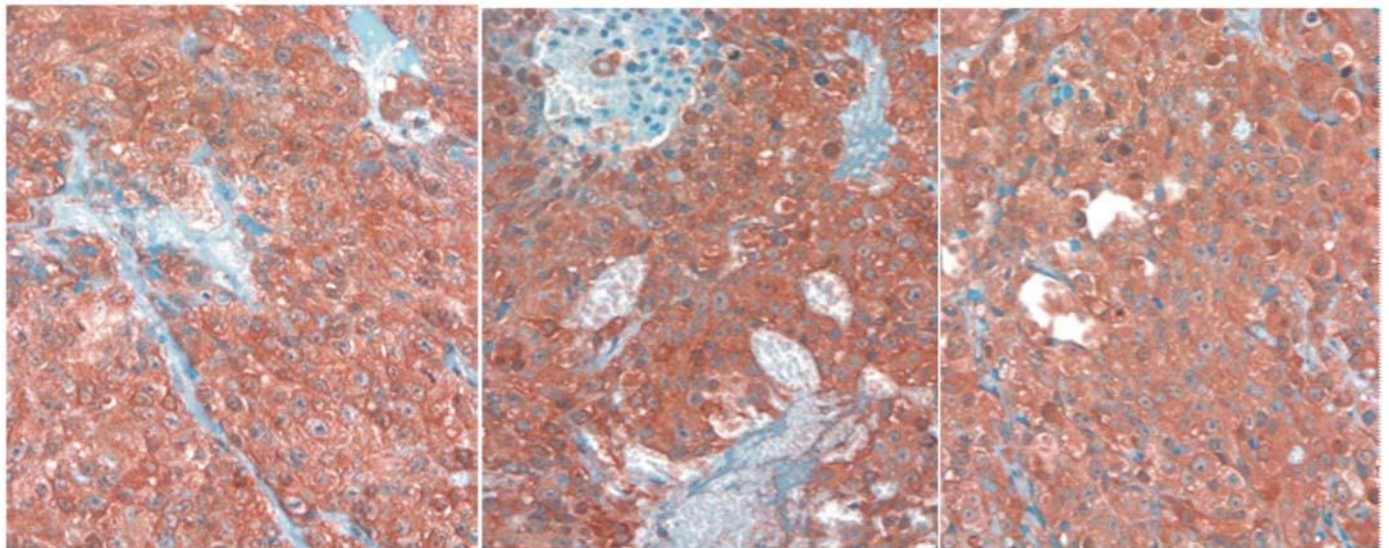


B Day after castration in SCID mice

No castration

14 days

35 days



Supplemental Fig. 3 Test of the diluted TRIM59 antibody for moderate IHC staining

Results: The following IHC staining showed results of the moderated TRIM59 IHC signals, when the first antibody TRIM59 diluted to 1:300, 1:600 and 1:1200. We assessed these IHC results with the negative control (without TRIM59 antibodies added) in five different tissues: ovary clear cell carcinoma, endometroid adenocarcinoma, lung squamous cell carcinoma, hepatocellular carcinoma and follicular lymphoma.

When comparing the first three columns with the negative control in Fig. s-3. 1:1200 produced less background than concentrated TRIM59 antibodies. However, the TRIM59 IHC staining still remain stronger than the normal non-tumor areas even at 1:300. In order to detect weak TRIM59-IHC staining signals, we also tried 1:300 dilution and compared with negative controls (no antibody added).

We also tested additional avidin/biotin blocking agent (from Vector Lab) to decrease the background staining, and similar results were obtained.

Legend of Figures:

(A).Ovary, clear cell carcinoma, (20X), positive, moderate to strong cytoplasmic staining in 1:300 dilution, weak to moderate cytoplasmic staining in 1:600 and 1:1200 dilution of first antibody, negative control, no staining. (B). Endometroid adenocarcinoma, (20X), positive, moderate to strong cytoplasmic staining in 1/300 dilution, weak to moderate cytoplasmic staining in 1:600 and 1:1200 dilution of first antibody, negative control, no staining (C). Lung squamous cell carcinoma (20X), positive, strong cytoplasmic staining in 1/300 and 1:600 dilution, moderate to strong cytoplasmic staining in 1:1200 dilution of first antibody, negative control, no staining. (D). Hepatocellular carcinoma, (40X), positive, strong cytoplasmic staining in 1/300 and 1:600 dilution, moderate to strong cytoplasmic staining in 1:1200 dilution of first antibody, negative control, no staining. (E).Follicular lymphoma, (20X),positive, weak cytoplasmic staining in 1:300, very weak cytoplasmic staining in 1:600 dilution , no staining with 1:1200 dilution, negative control, no staining

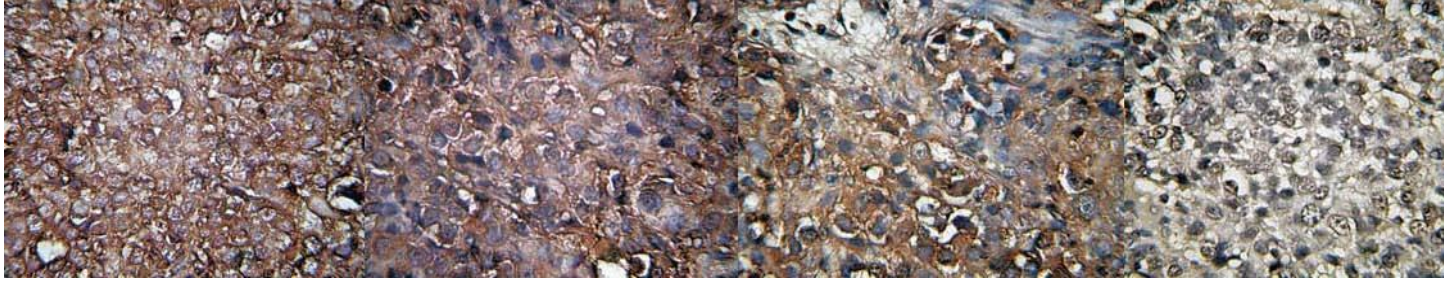
Supplemental Fig. 3 Titration of the diluted TRIM59 antibody in TMA (5 tumors) for moderate IHC staining

Dilutions of TRIM59 first antibody

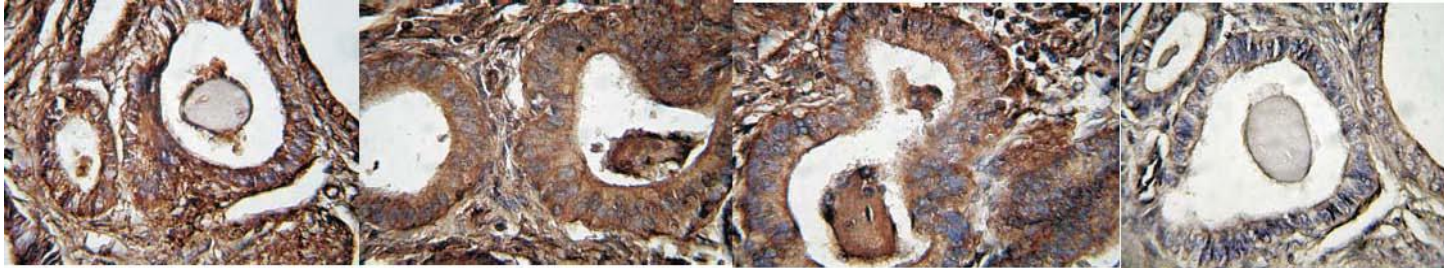
1/600 1/1200 Negative control

1/300

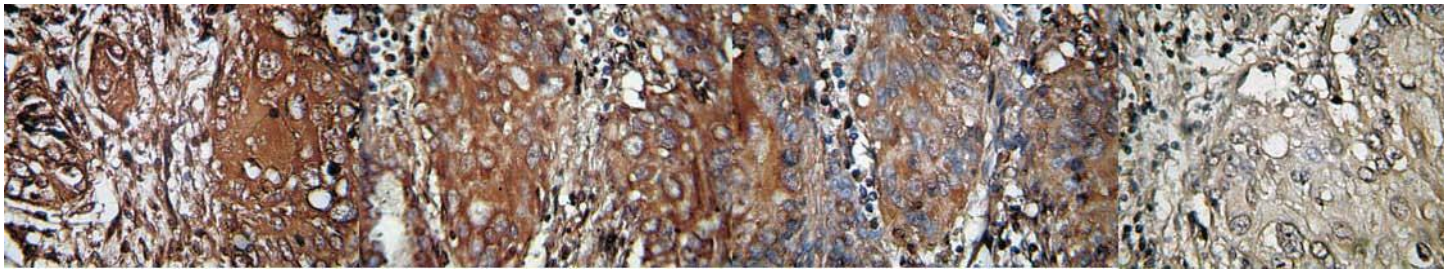
A. Ovary, clear cell carcinoma



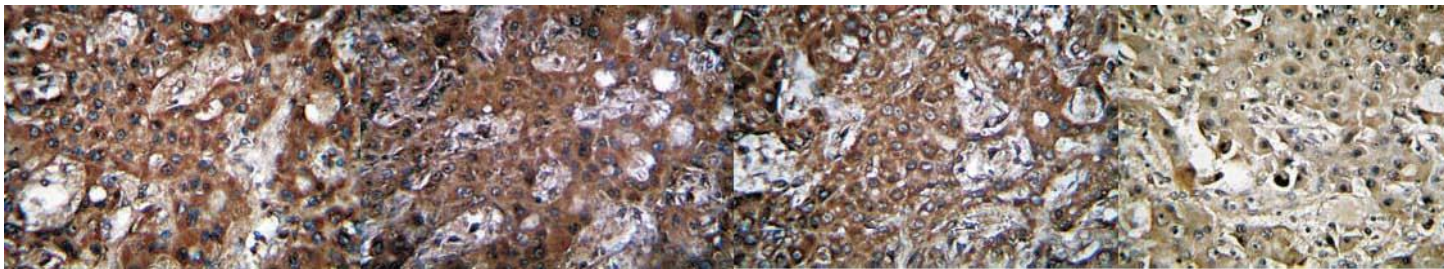
B. Endometrioid adenocarcinoma



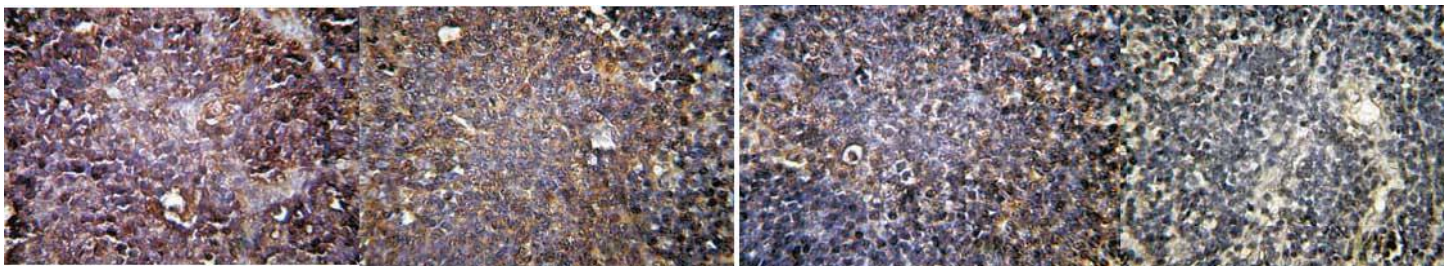
C. Lung SCC



D. Hepatocellular carcinoma



E. Follicular lymphoma



Supplemental Fig. 4 Comparison of TRIM59 protein levels in immunohistochemistry

(IHC) staining in human multiple tumor TMA (tissue microarray) with negative controls

Summary of results:

We performed IHC staining of 35 tumors TMA with both TRIM59 antibody (positive) and without any antibody (negative) in serial consecutive slides for each tissue and in all three cores for each patient. The comparison was performed histologically by cell and tissue type, since there were some differences even in serial slides in some area of TMA, tissues in each core, which were not identical, and in some cores tissues were missing during IHC process.

The following results in most (25 panels marked as #) of 35 tumor TMA confirmed results of Table 2 of the text on TRIM59-TMA. **Strong cytoplasmic /nuclear staining by**

TRIM59 antibody in tumor area:

#3 Skin (squamous cell carcinoma), #4 Skin (basal cell carcinoma), #10 Breast ductal adenocarcinoma; #12 Thyroid medullary carcinoma; #13 Lung squamous cell carcinoma; #14. Lung bronchoalveolar carcinoma; #15 Lymph node with metastatic carcinoid; #16 Lung mesothelioma; #17 Hepatocellular carcinoma, #18 Metastatic carcinoid liver; #19 Endometrial carcinoma. **Moderate cytoplasmic /nuclear staining by TRIM59 antibody in tumor area:**

#1 Kidney (Clear cell carcinoma), #2 Adrenal gland (cortical carcinoma), #7 Omentum (serous adenocarcinoma), #9 Cervix adenocarcinoma; **Weak cytoplasmic /nuclear staining by TRIM59 antibody in tumor area featured less different with controls:**

#5 Skin (melanoma), #6 Uterus (leiomyosarcoma), #8 Ovary, serous adenocarcinoma, #11 Bladder (urothelial carcinoma); #20 Small bowel, marginal zone lymphoma; #21 Spleen, Hodgkin's lymphoma; #22 Stomach, malt cell lymphoma; #23 Thymoma; and # 25. Lymph node follicular lymphoma. **Negative cytoplasmic /nuclear staining by TRIM59 antibody in tumor area:** #24 Appendix (goblet cell carcinoid)

Legend of panels:

1. 1. Kidney (Clear cell carcinoma), positive IHC for TRIM59 expression moderate cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).
2. 2. Adrenal gland (cortical carcinoma), positive IHC for TRIM59 expression moderate cytoplasmic staining with no nuclear staining; negative control in negative control, weak background staining in cytoplasmic of few cells (20X, 40X).
3. 3. Skin (squamous cell carcinoma), positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).
4. 4. Skin (basal cell carcinoma), positive IHC for TRIM59 expression strong cytoplasmic

staining with no nuclear staining; negative control no staining (20X, 40X).

5. 5. Skin (melanoma), positive IHC for TRIM59 expression very weak cytoplasmic staining and nuclear staining; negative control no staining (20X, 40X).

6. 6. Uterus (leiomyosarcoma), positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

7. 7. Omentum (serous adenocarcinoma), positive IHC for TRIM59 expression moderate cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

8. 8. Ovary (serous adenocarcinoma), positive IHC for TRIM59 expression weak to moderate cytoplasmic staining and no nuclear staining, negative control no staining (20X, 40X).

9. 9. Cervix adenocarcinoma, positive IHC for TRIM59 expression moderate to strong cytoplasmic staining; negative control no staining (20X, 40X).

10. 10. Breast ductal adenocarcinoma, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

11. 11. Bladder (Urothelial carcinoma), positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

12. 12. Thyroid medullary carcinoma, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

13. 13. Lung squamous cell carcinoma, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

14. 14. Lung bronchoalveolar carcinoma, positive IHC for TRIM59 expression strong cytoplasmic and nuclear staining; negative control no staining (20X, 40X).

15. 15. Lymph node metastatic carcinoid, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

16. 16. Lung mesothelioma, positive IHC for TRIM59 expression strong cytoplasmic and nuclear staining; negative control no staining (20X, 40X).

17. 17. Hepatocellular carcinoma, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

18. 18. Liver metastatic carcinoid, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

19. 19. Endometrial carcinoma, positive IHC for TRIM59 expression strong cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

20. 20. Small bowel, marginal zone lymphoma, positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

21. 21. Spleen, Hodgkin lymphoma, positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

22. 22. Stomach, malt lymphoma, positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining.

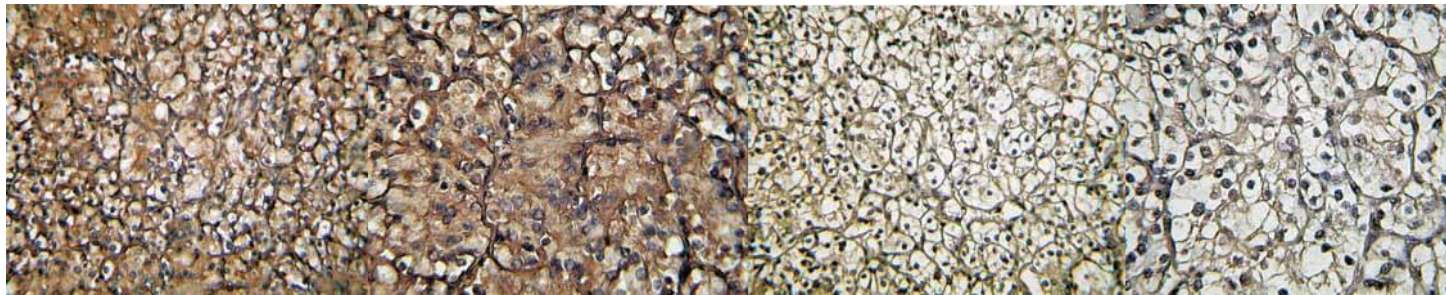
23. Thymoma, positive IHC for TRIM59 expression weak cytoplasmic staining and no nuclear staining; negative control no staining (20X, 40X).

1. 24. Appendix (Goblet cell carcinoid), IHC for TRIM59 expression no staining, negative control no staining (20X, 40X).

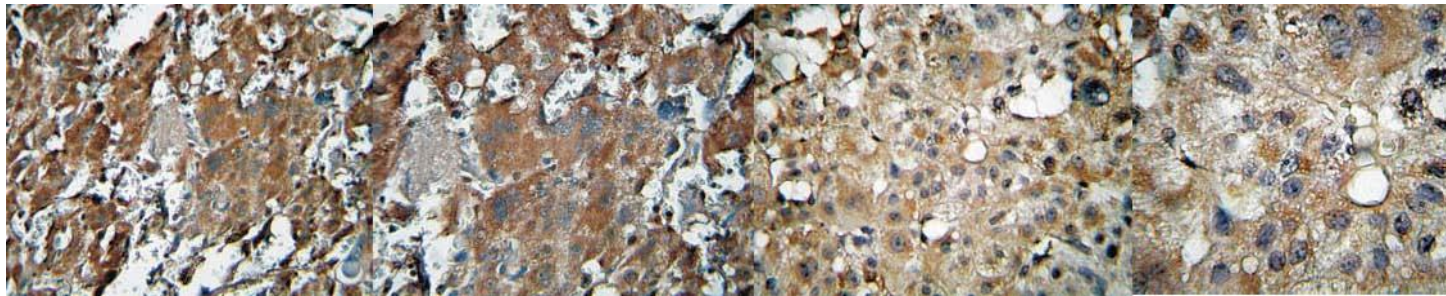
2. 25. Follicular lymphoma, positive IHC for TRIM59 expression weak cytoplasmic staining, negative control no staining (20X, 40X).

Suppl.Fig. 4 Positive and negative controls of immunohistochemistry (IHC) staining by TRIM59 antibody in 25 tumors listed from multiple-cancers TMA

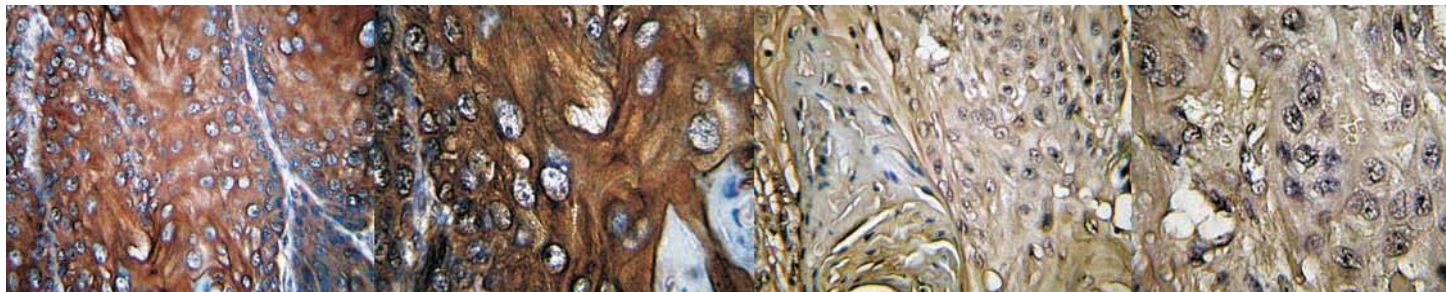
1-Kidney, clear cell carcinoma, positive (moderate) Negative



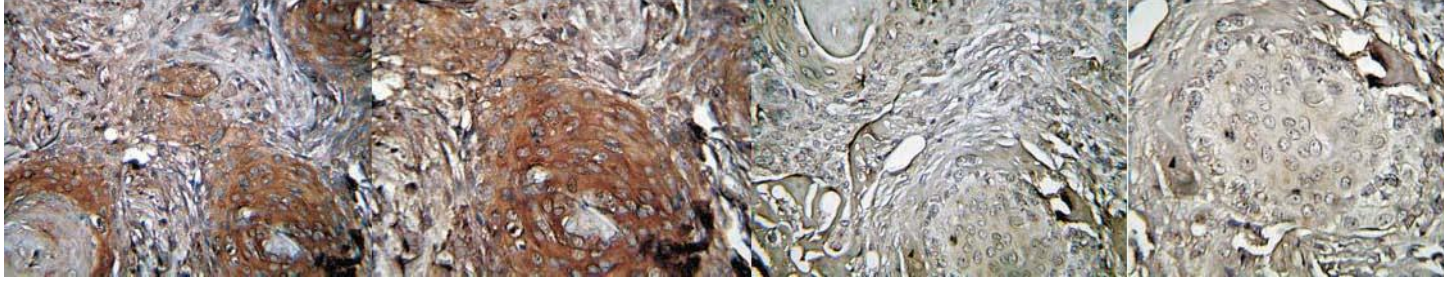
,2-Adrenal gland, cortical carcinoma, positive (moderate) Negative



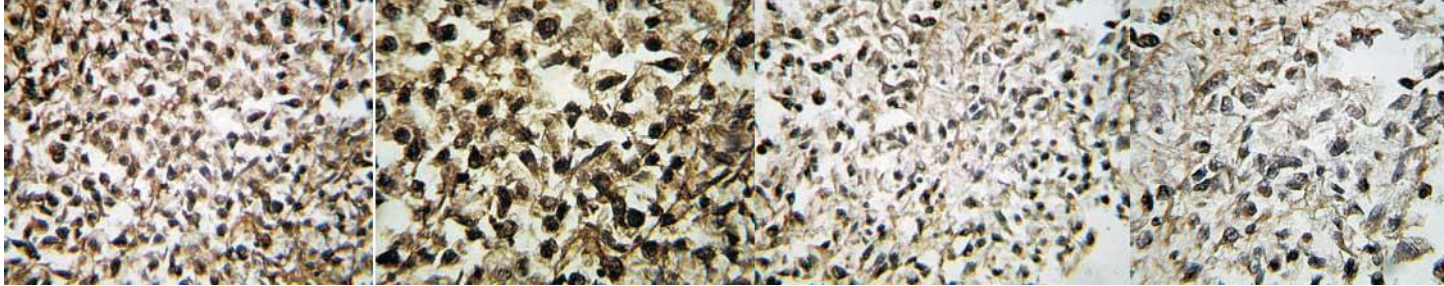
3-Skin, SCC, positive (strong) Negative



4-Skin, BCC, positive(strong) Negative



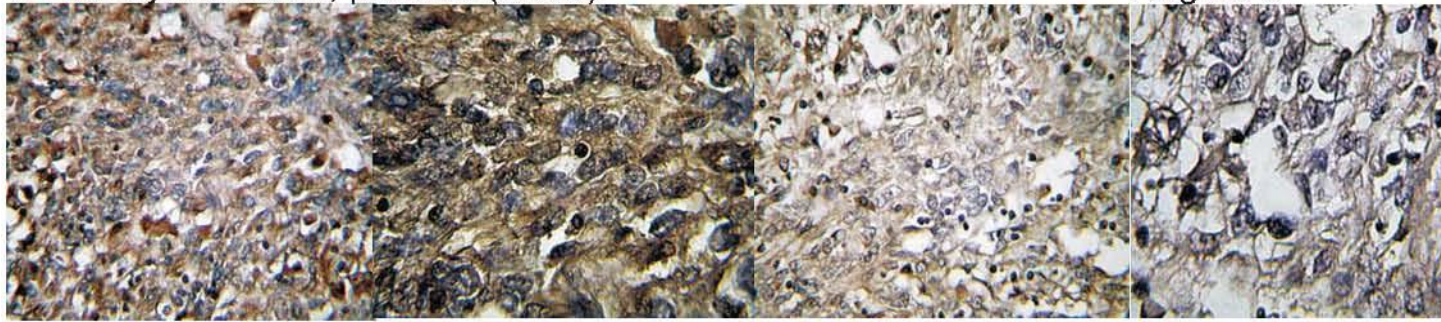
5-Skin, melanoma, positive (weak) Negative



x20 x40 x20 x40

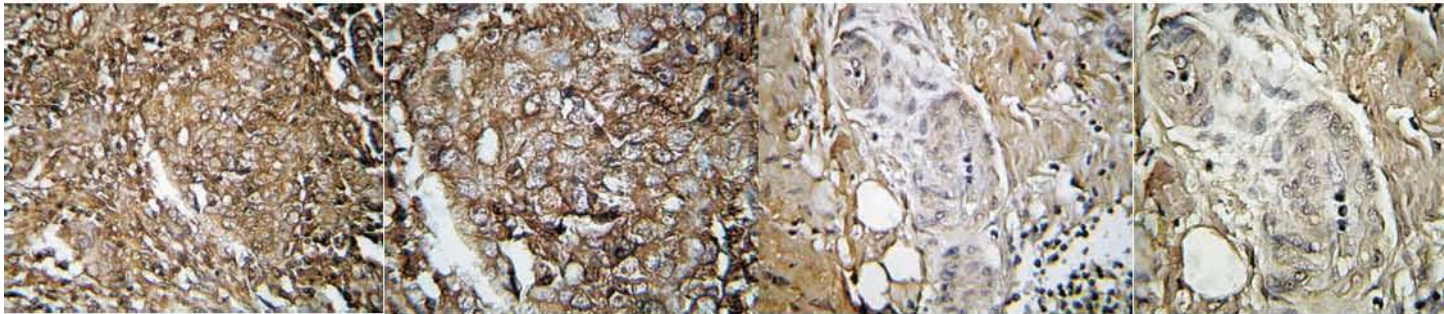
6-Leiomyosarcoma, positive (weak)

Negative



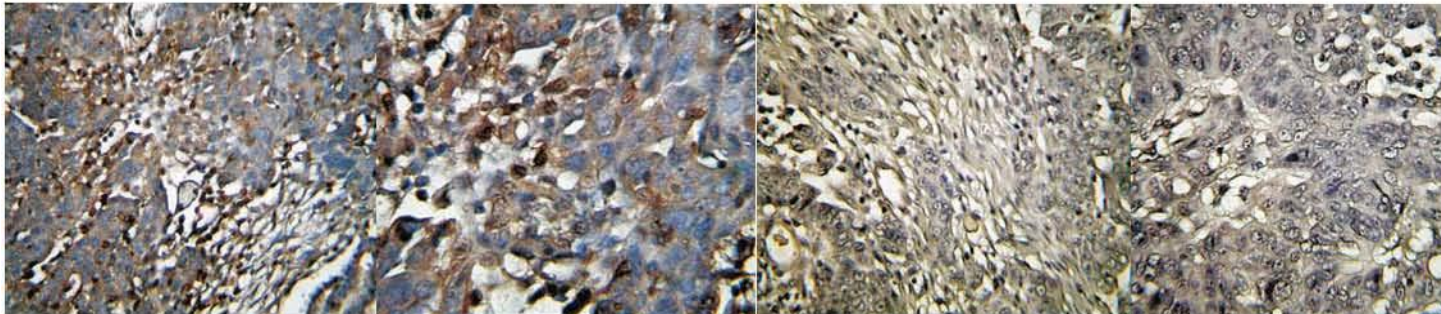
7-Omentum, serous adenocarcinoma, positive (moderate)

Negative



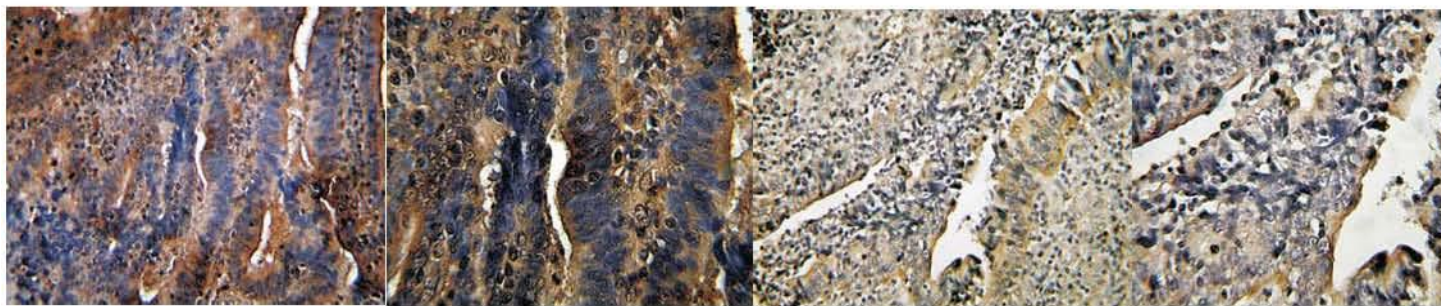
8-Ovary, serous adenocarcinoma, positive (weak)

Negative



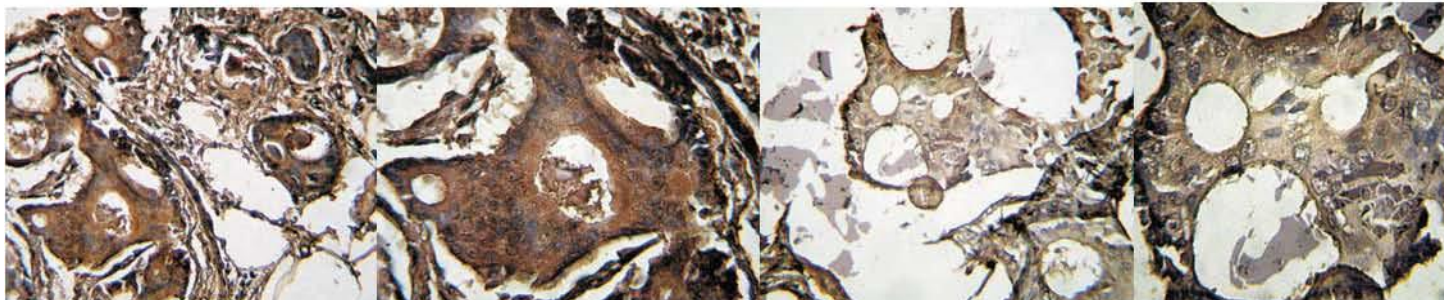
9-Cervix adenocarcinoma, positive (moderate)

Negative



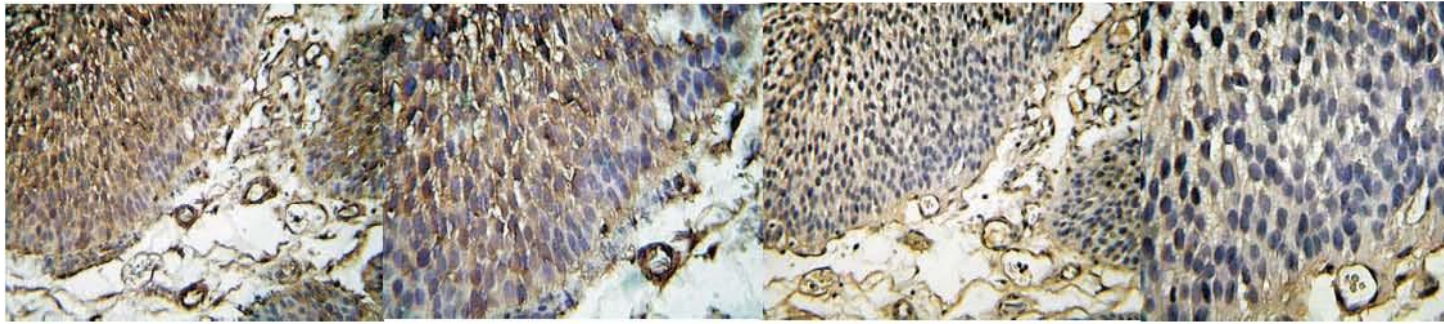
10-Breast ductal adenocarcinoma, positive (strong)

Negative



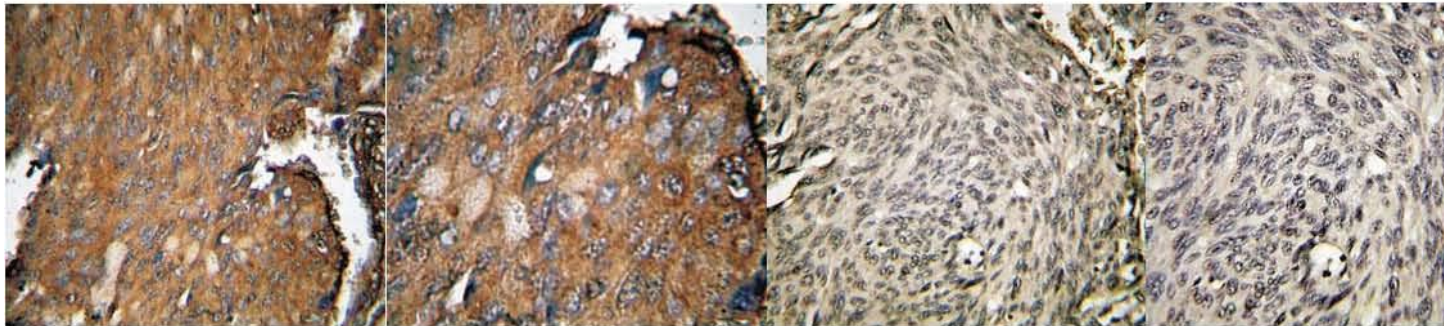
11- Bladder, Urothelial carcinoma, positive (weak)

Negative



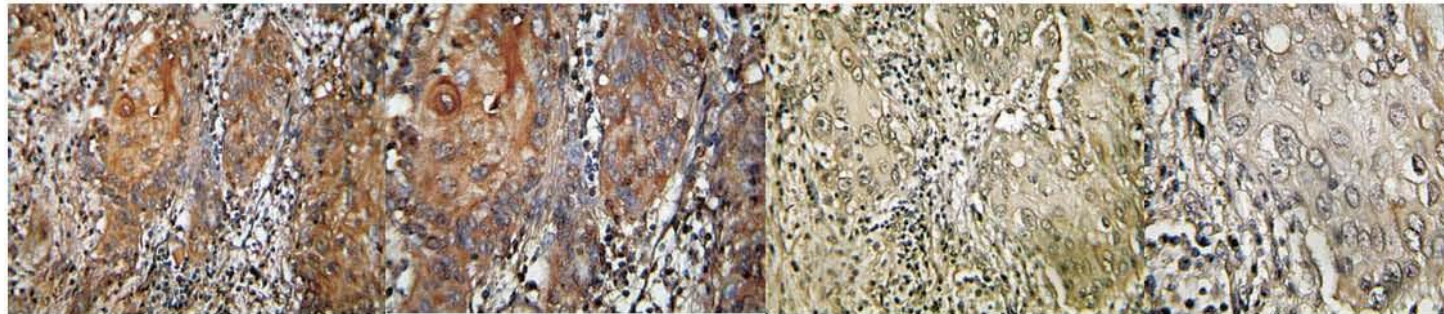
12-Thyroid, medullary carcinoma, positive (strong)

Negative



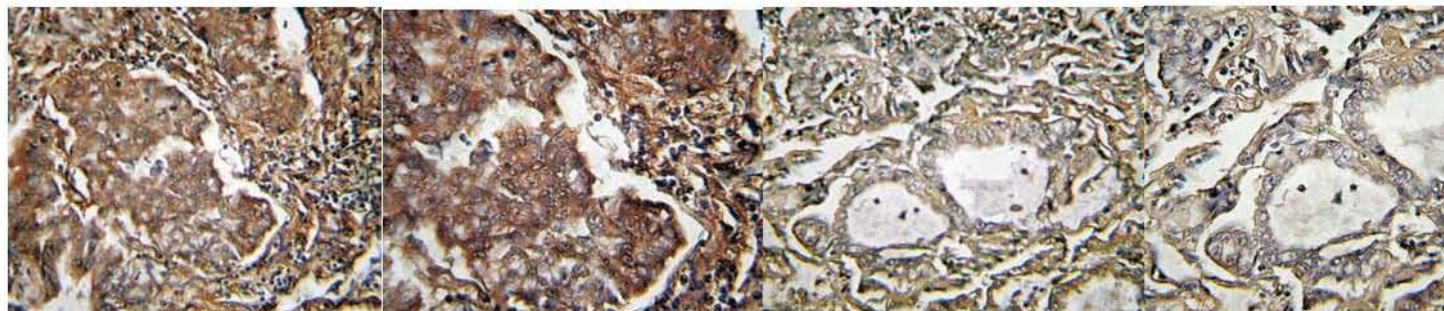
13-Lung SCC, positive (strong)

Negative



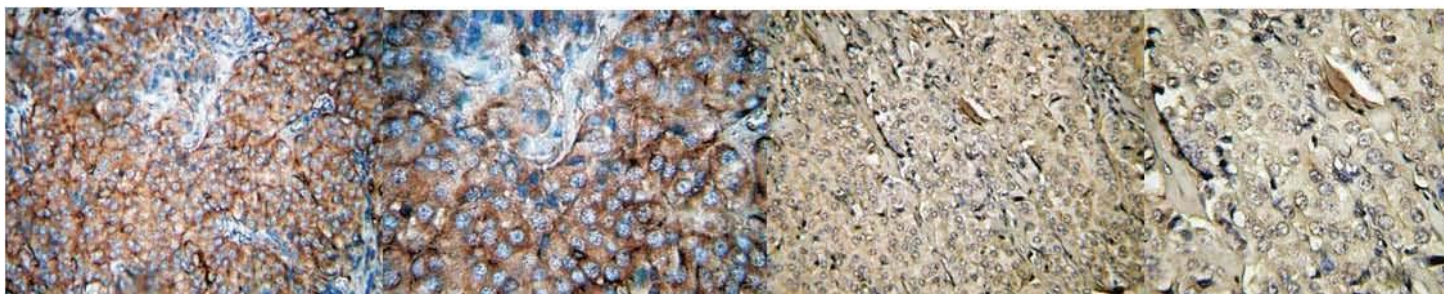
14- Lung, bronchoalveolar carcinoma, positive (strong)

Negative



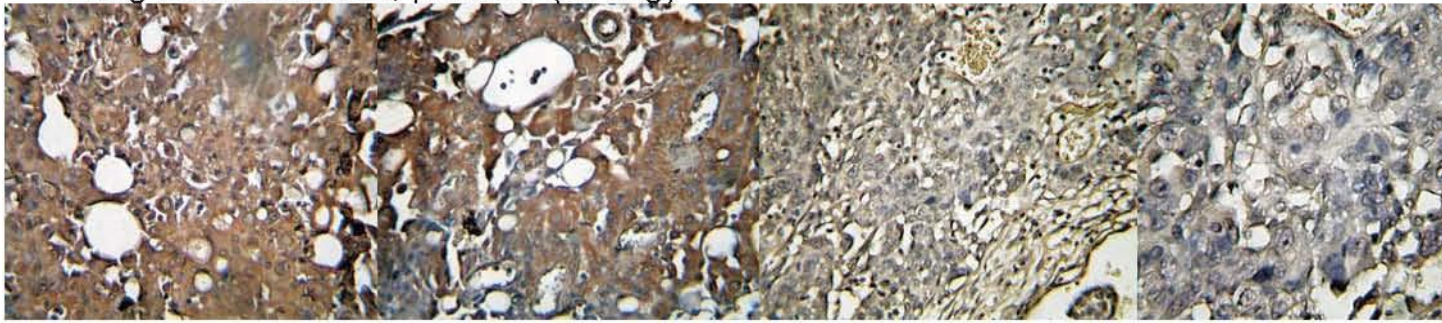
15-Lymph node metastatic carcinoid, positive (strong)

Negative



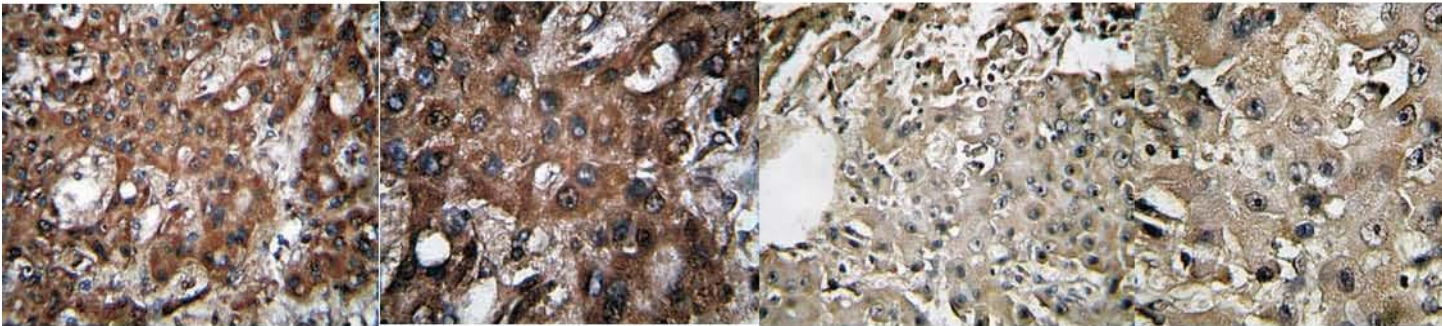
16- Lung mesothelioma, positive (strong)

Negative



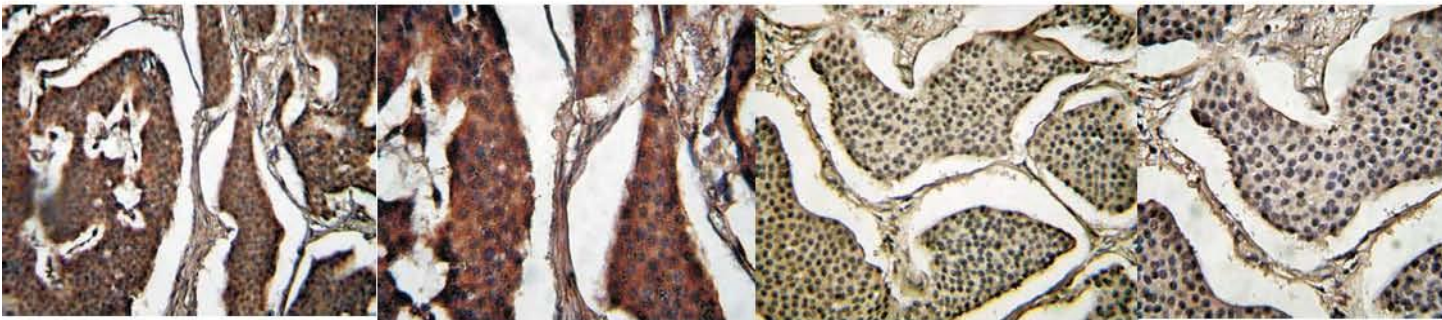
17- Hepatocell carcinoma, positive (strong)

Negative



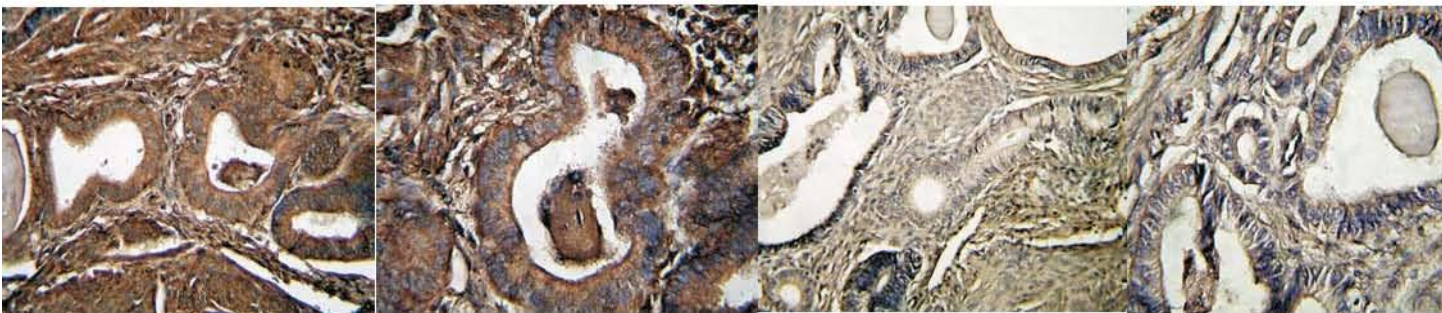
18- Liver metastatic carcinoid, positive (strong)

Negative



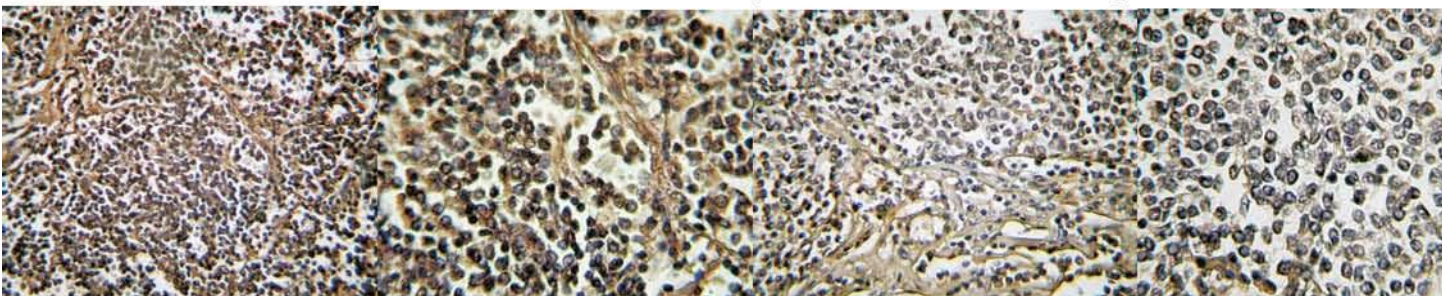
19- Endometrial adenocarcinoma, positive (strong)

Negative



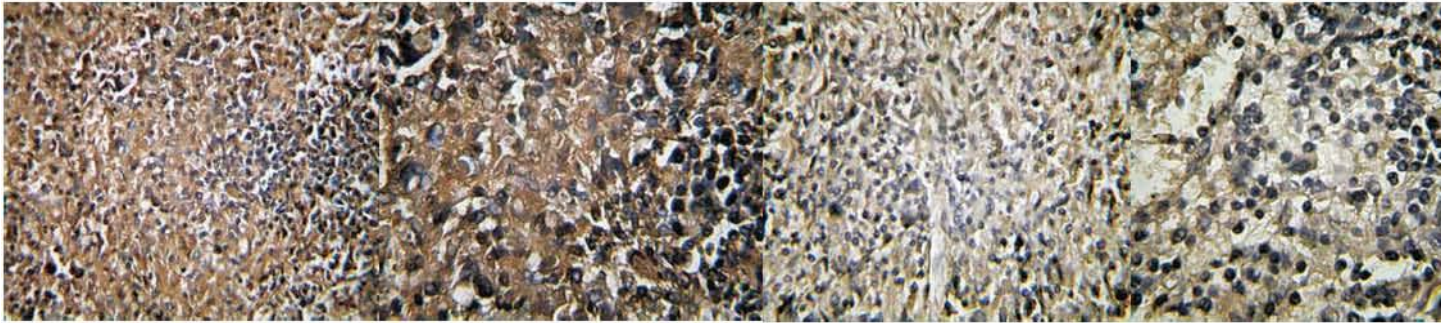
20- Small bowel marginal lymphoma, positive (weak)

Negative



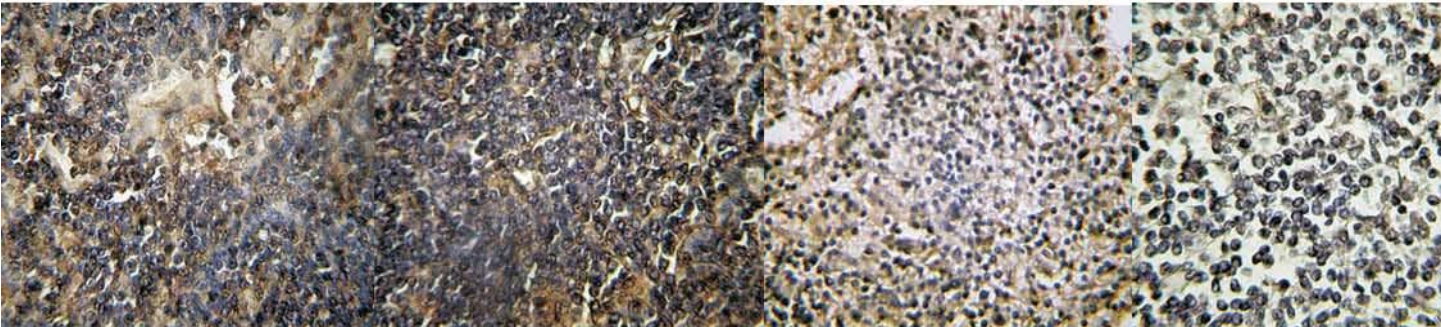
21- Spleen hodgkin lymphoma, positive (weak)

Negative



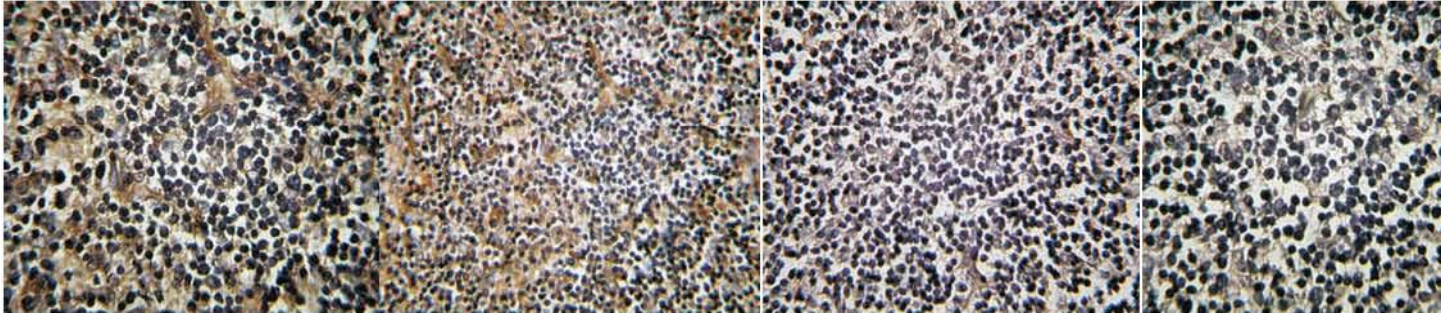
22- Stomach malt lymphoma, positive (weak)

Negative



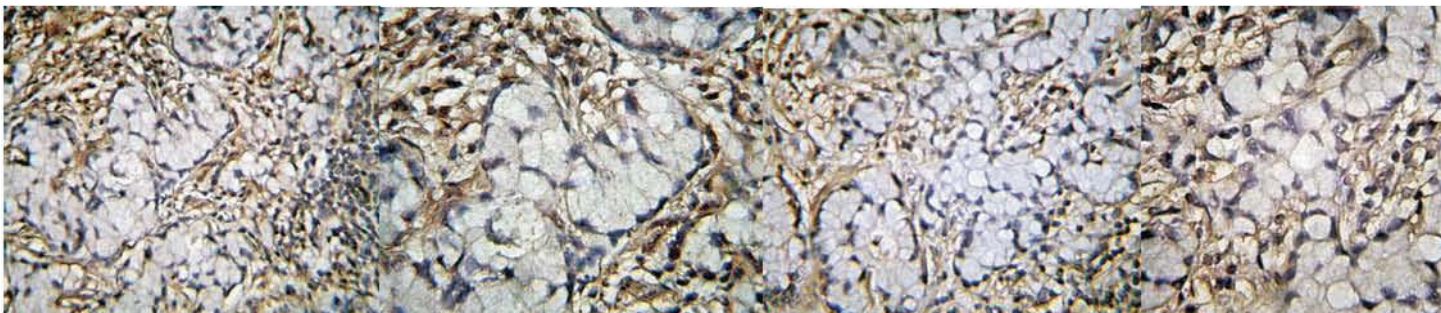
23- Thymoma, positive (weak)

Negative



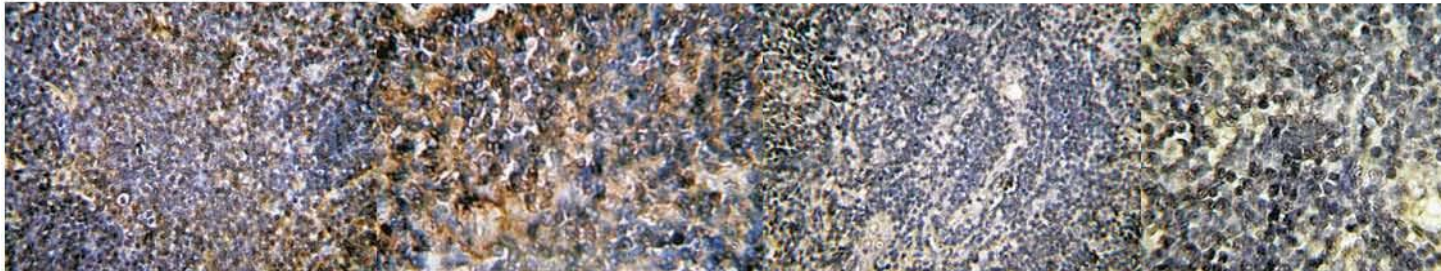
24- Appendix, Gablet cell carcinoid, positive (no staining)

Negative



25- Follicular lymphoma, positive (weak)

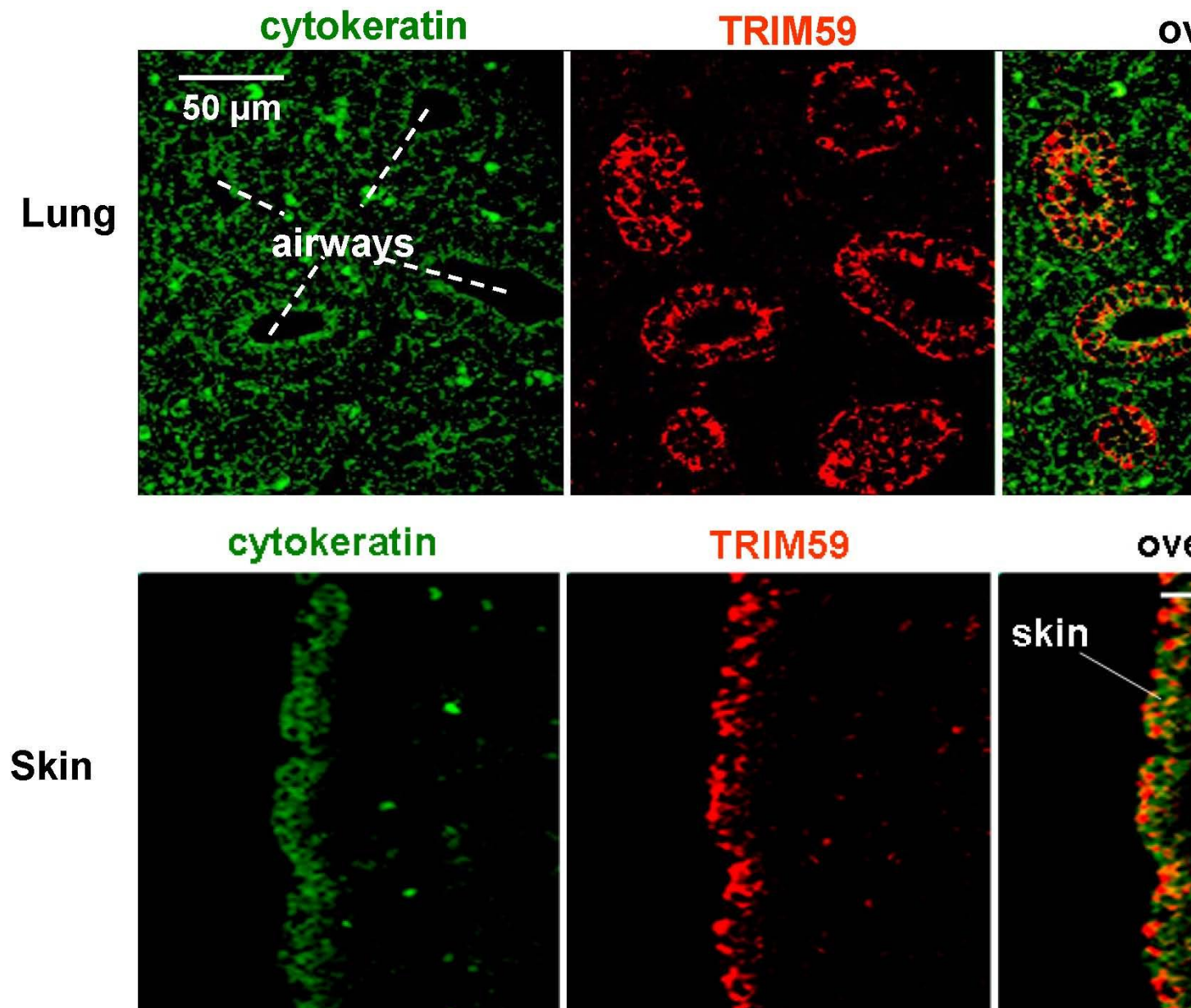
Negative



Confocal microscope imaging of
immunohistochemistry of TRIM59 protein in
mouse embryo (14.5d.p.c)

Supplemental

Fig. 5



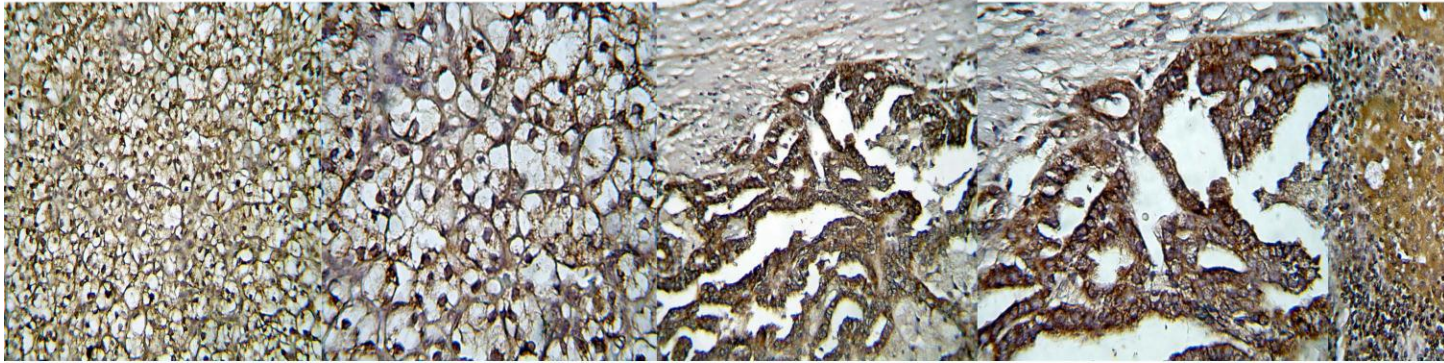
Supplemental Fig. 6

Test of Ras activation in kidney cancer patients correlated with up-

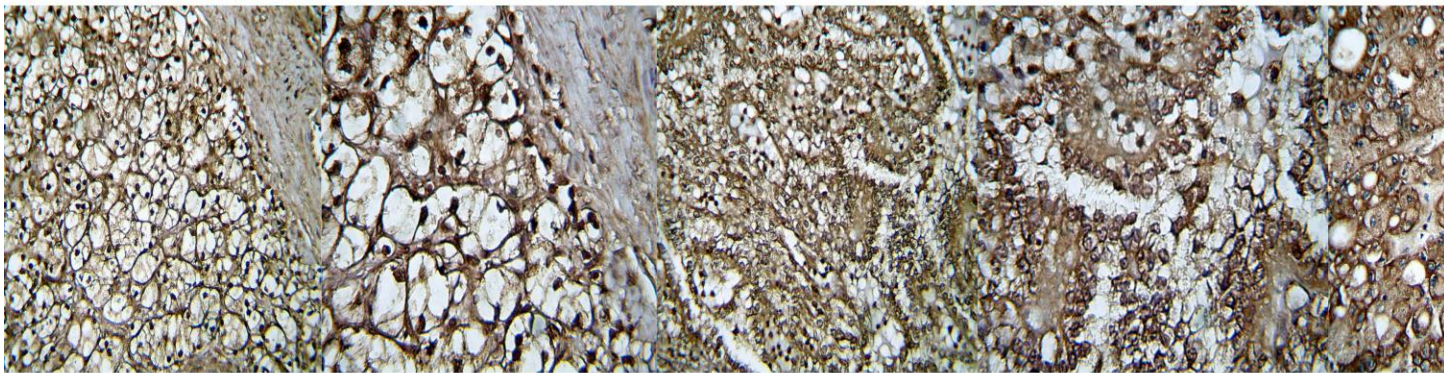
A. Clear cell carcinoma

B. Papillary RCC

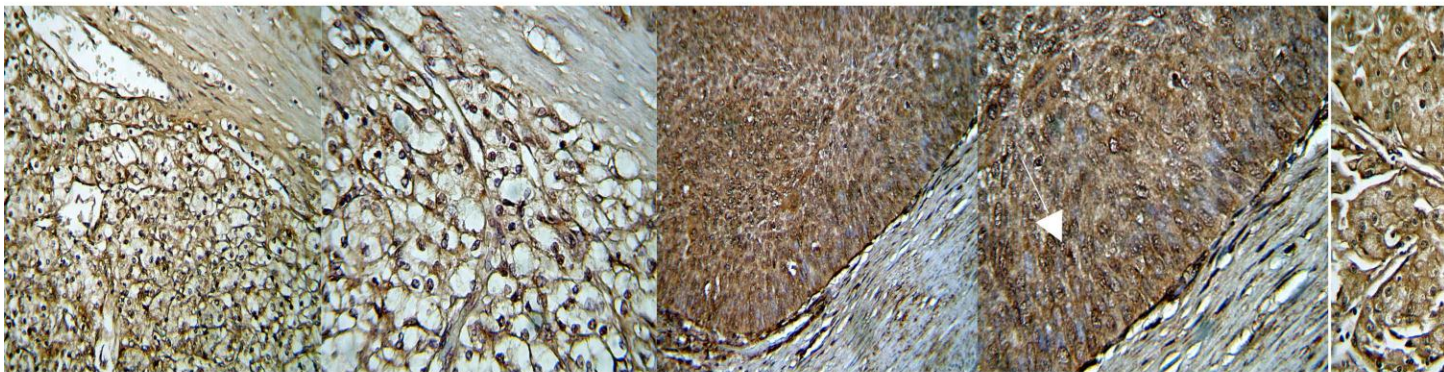
Antibody: B-Raf for total B-raf



Antibody: Raf 1 for total Raf



Antibody: Phosphor-Ser- 259 Raf 1



X20

X40

X20

X40

Supplemental Table 1 Correlation of TRIM59 up-regulation with B-Raf hyper-phosphorylation

Antibody used	RCC types	Patient number	Intensity score	Nuclear staining
B-Raf (total B-raf)	Clear cell Carcinoma	6	0-1	No
	Chromophobe	3	2-3	No
	papillary	3	2-3	No
Raf-1 (total B-raf)	Clear cell Carcinoma	3	0	No
	Chromophobe	2	2-3	No
	papillary	1		No
B-Raf p-Ser ²⁵⁹ (phosphorylated B-Raf)	Clear cell Carcinoma	3	0-1	No
	Chromophobe	1	2-3	20%
	papillary	2	2-3	50%

Since BRAF is the most important early effector in the Ras signal pathway, and Raf mediate phosphorylation is the main effector recruited by GTP-bound Ras to activate the MEKMAP kinase pathway, we performed IHC studies in these RCC sections by probing Raf activation /up-regulation and hyper-phosphorylation. Three antibodies were used: B-Raf Antibody (mAb) and Raf-1 Antibody (Ab-259) testing for total Raf protein and Raf-1 Antibody (Phospho-Ser²⁵⁹ pAb) testing activated phosphorylated B-Raf. Some of serial slides were stained parallelly on each patient by different antibodies.

Supplemental Fig. 1 Characterization of TRIM59 antibodies constructed and used for human tissue studies.

Summary of results: The antibody TRIM59#72 used for mouse and human studies were from C-terminal 150 a.a. peptide, which is TRIM59 specific, while N-terminal sequence of TRIM59 contains mostly the common function domains of the 70 member TRIM59 family. In Western blots on both mouse and human cell lysates, #72 recognized only one band, while #71 showed more bands. We demonstrated TRIM59#72 can be used for human tissues for immunohistochemistry and Western blots.

(A). Diagram of a novel TRIM family member TRIM59 showing the gene structure (upper line), functional domains of RBCC family. Two antibodies (TRIM59#71 and #72) are shown by arrows. (B). 10 % SDS-PAGE showing purified GST-TRIM59 protein #71 and #72 protein (shown by arrows). GST protein showed as a control (26kda, by arrow). (C-D). Characterization of polyclonal antibodies (#71 and #72) of mouse TRIM59 in mouse cell and tissue lysates. Similar identical Western blots prepared for total cellular IMAC column purified proteins and TRIM59#72 antibody-affinity column purification products from NIH3T3 cells, prostate tissue of wild type (WT), KIMAP and TGMAP prostate tumor samples. TRIM59 protein was shown by an arrow. (D) Characterization of polyclonal antibodies against mouse TRIM59. Similar identical Western blots prepared for total cellular IMAC column purified proteins and TRIM59#72 antibody-affinity column purification products from NIH3T3 cells, prostate tissue of wild type (WT), KIMAP and TGMAP prostate tumor samples. Using a TRIM59#72 affinity column purified TRIM59 protein, both TRIM59#71 and TRIM59#72 recognized the same major band (53kDa) in Western blots. (E) Western blots test of levels of TRIM59 protein in human prostate cancer cultured cells PC3, DU145, LNCaP and human kidney cell HEK293. A unique 53 kDa band was shown exactly as in mouse cell and tissue lysates (C).

R. Comparison of relative scores of TRIM59 IHC staining in eight different tumor types

