BMJ Open RECAST: Study protocol for an observational study for the understanding of the increased REsilience of Children compared to Adults in SARS-CoV-2 infection

Sebastian Stricker 1,1 Niklas Ziegahn 1, Martin Karsten,2 Thomas Boeckel,3 Heike Stich-Boeckel,4 Jakob Maske,5 Evelyn Rugo,2 Anita Balazs,1 Pamela Millar Büchner,1 Chantip Dang-Heine,6 Valentin Schriever,7,8 Roland Eils,9,10 Irina Lehmann,9,11 Leif E Sander,12 Markus Ralser,13,14 Victor M Corman,15 Marcus A Mall,1,11 Birgit Sawitzki,16,17 Jobst Roehmel 1,16

ABSTRACT

Introduction The SARS-CoV-2 pandemic remains a threat to public health. Soon after its outbreak, it became apparent that children are less severely affected. Indeed, opposing clinical manifestations between children and adults are observed for other infections. The SARS-CoV-2 outbreak provides the unique opportunity to study the underlying mechanisms. This protocol describes the methods of an observational study that aims to characterise age dependent differences in immune responses to primary respiratory infections using SARS-CoV-2 as a model virus and to assess age differences in clinical outcomes including lung function.

Methods and analysis The study aims to recruit at least 120 children and 60 adults that are infected with SARS-CoV-2 and collect specimen for a multiomics analysis, including single cell RNA sequencing of nasal epithelial cells and peripheral blood mononuclear cells, mass cytometry of whole blood samples and nasal cells, mass spectrometry-based serum and plasma proteomics, nasal epithelial cultures with functional in vitro analyses, SARS-CoV-2 antibody testing, sequencing of the viral genome and lung function testing. Data obtained from this multiomics approach are correlated with medical history and clinical data. Recruitment started in October 2020 and is ongoing.

Ethics and dissemination The study was reviewed and approved by the Ethics Committee of Charité – Universitätsmedizin Berlin (EA2/066/20). All collected specimens are stored in the central biobank of Charité – Universitätsmedizin Berlin and are made available to all participating researchers and on request.

Trial registration number DRKS00025715, pre-results publication.

STRENGTHS AND LIMITATIONS OF THIS STUDY

⇒ Sample collection from children and adults with primary SARS-CoV-2 infection at multiple time points, however, samples from severely ill patients are not included.
⇒ Mass cytometry, single-cell RNA sequencing and mass spectrometry-based serum and plasma proteomics display the local and systemic immune response.
⇒ Air–liquid interface cultures reproduce in vivo conditions and will be used for functional studies.
⇒ Analysis of clinical data and lung function testing complement the multiomics approach.

INTRODUCTION

In December 2019, the novel coronavirus SARS-CoV-2 emerged as cause of acute pneumonia.1 2 By August 2021, more than 200 million people were infected with SARS-CoV-2.3 Soon after the beginning of the pandemic, it became obvious that children have an increased resilience against the primary infection. The course of disease in children is more likely to be milder and severe or even fatal courses remain extremely rare.4–8 Various hypotheses to explain the reduced susceptibility and mortality of children are currently discussed, including reduced virus entry via ACE-2 in children,9 preactivated components of the immune system, such as cross-reactive T cells10–13 and antibodies,14 or a more accentuated innate immunity in children15 16 (table 1). Most of these findings are complementary in the explanation of the observed phenomenon, however, some findings are in part contradictory and require further investigation. Opposing clinical manifestations between children and adults are also observed for other viral respiratory infections.15–17 This points to major changes
Table 1  Hypotheses to explain the resilience of children in SARS-CoV-2 infections

<table>
<thead>
<tr>
<th>Hypothesis</th>
<th>Proposed explanation</th>
<th>Scientific findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reduced susceptibility for SARS-CoV-2 in children</td>
<td>Reduced virus entry via ACE-2 in children</td>
<td>SARS-CoV-2 uses ACE-2 in the upper and lower airways for host cell entry(^82). The age-dependency of ACE-2 expression is controversially discussed.(^83-85) Even though an increased expression of ACE-2 renders the individual more susceptible to viral infection, ACE-2 also initiates anti-inflammatory signalling and might contribute to a milder immune response.(^86)</td>
</tr>
<tr>
<td>Age–dependent differential immune activation pattern</td>
<td>Preactivated immune components in children entail a milder immune response</td>
<td>In early childhood, infections of the upper respiratory tract are frequent. It has been proposed that previous infections with coronaviridae might contribute to a cross-reactive immunity.(^87) Pre-existing T cell reactivity to SARS-CoV-2 could affect the severity of COVID-19.(^19) Cross-reactive antibodies entail a milder immune response to SARS-CoV-2.(^14) Of note, uninfected infants do not express cross-reactive antibodies.(^88) The polyclonality and polyreactivity of IgM naturally present in children recognises SARS-CoV-2 particles.(^89)</td>
</tr>
<tr>
<td>Children possess a stronger innate immunity than adults</td>
<td>Children display a higher basal expression of pattern recognition receptors than adults and a stronger innate antiviral response.(^76)(^77) The nasopharyngeal mucosa of children exhibits a stronger innate immunity and expresses more antiviral cytokines than adults.(^90)</td>
<td></td>
</tr>
<tr>
<td>Children expose a different cytokine response on SARS-CoV-2 infection than adults</td>
<td>In COVID-19, a cytokine storm leads to acute respiratory distress syndrome.(^63)(^64) Certain cytokine patterns correlate with COVID-19 severity.(^91) Proinflammatory cytokine concentrations might be lower in children infected with SARS-CoV-2 than in adults.(^82)</td>
<td></td>
</tr>
<tr>
<td>Coinfections lead to a milder immune reaction, for example, because of virus competition or primed immune components</td>
<td>In COVID-19, coinfection with other pathogens is not rare, especially in children.(^93)(^95)</td>
<td></td>
</tr>
</tbody>
</table>

in the general immune response pattern during ageing. In the past, comparative immune response analyses to primary infections in various age groups were difficult to perform, as many adults had already been exposed to the pathogens. The SARS-CoV-2 outbreak provides the unique opportunity to study the age-dependent changes in immune responses in a controlled manner.

To understand the mechanisms behind the lower susceptibility of children compared with adults to develop severe COVID-19 disease, we have established the observational study RECAST (increased REsilience of Children compared to Adults in SARS-CoV-2 infecTion) focusing on the differences in the clinical presentation, lung function and the immune response to SARS-CoV-2 in children compared with adults.

The complexity of immune responses requires a multilevel approach to display changes on various layers, including local immune cell composition, cytokine signalling and systemic response. It can be assumed that the combination of several mechanisms leads to the largely different phenotypes. At the same time, modern techniques allow to engage on an exploratory approach analysing simultaneously the involvement of canonical and non-canonical immune response patterns. The multiomics approach presented here allows deeply detailed characterisation of the various layers of age dependent specific immune responses. Therefore, we believe that the presented study design will contribute to a further understanding even beyond COVID-19.

To meet these requirements adequately, we chose a multiomics approach, including: (1) single-cell RNA sequencing (scRNAseq) of peripheral blood mononuclear cells (PBMCs) and nasal epithelial cells, (2) mass spectrometry-based serum and plasma proteomics, which has been used to identify prognostic marker signatures for SARS-CoV-2 disease severity and devise risk-adapted treatment strategies,\(^18\) (3) mass cytometry (cytometry by time-of-flight, CyTOF) of whole blood samples and nasal cells, which has been used to elucidate the role of T cell cytotoxicity in COVID\(^19\) and to identify a dysregulation of the myeloid cell compartment as hallmark of severe COVID,\(^20\) (4) highly differentiated nasal epithelial cultures and functional in vitro analyses, which have been used to display age-related differences in the nasal...
epithelium, antibody testing and sequencing of the viral genome. Obtained data are complemented with anamnestic and clinical information, lung function testing, including spirometry and multiple breath washout (MBW), which is a standardised method which allows to assess the ventilation homogeneity of the lungs already in preschool children as well as smell and taste. Longitudinal sampling allows monitoring of the immune response over the course of disease and beyond. Due to the maturation of the immune system during childhood, age-specific immune response pattern against SARS-CoV-2 can be expected. Thus, participants of all age groups are enrolled. Participant recruitment began in October 2020 and is ongoing. Of particular interest is the recruitment of children infected with various SARS-CoV-2 virus variants of concern.

Study objectives
This study aims to characterise and compare primary infections with SARS-CoV-2 in children and adults, and to identify age-related determinants of disease course and prognosis. The immune system is not only highly complex, but immune response patterns also vary depending on individual predisposition; moreover, it also matures throughout the ageing of an individual. Furthermore, pivotal interest for the interpretation of these data is the correlation between immune response pattern and clinical outcome.

METHODS AND ANALYSIS
Study design
RECAST is a prospective observational cohort study at Charité – Universitätsmedizin Berlin in Berlin, Germany. It is a substudy of the Pa-COVID study of the Charité, aiming to characterise the disease course of patients suffering from COVID-19.

Data are collected longitudinally from patients with confirmed COVID-19 at three time points, directly after the diagnosis and at follow-up visits after 2 weeks and 4–6 months, and from healthy age-matched controls.

Recruitment started in October 2020 and is planned to end in October 2023.

Study population
Inclusion criteria
Main inclusion criteria for the index person are a primary acute SARS-CoV-2 infection in a minor (<18 years of age) with positive PCR or antigen testing (both will be confirmed by PCR testing).

Exclusion criteria
Subjects with pre-existing conditions affecting the immune response, such as diseases requiring chemotherapy or syndromes with immunodeficiency and subjects with concomitant medication that affects the immune response, such as systemic steroids, biologicals or investigational therapeutics targeting SARS-CoV-2, are excluded.

Study procedures
Patient identification and recruitment
A network of participating paediatric outpatient practices (n=20) has been established as sentinels to provide access to a pool of >25 000 paediatric patients. All children who tested positive for SARS-CoV-2 by PCR or antigen testing as well as their household members are eligible for inclusion.

Healthy controls are recruited from clinical routine diagnostic settings if the diagnostic screening for SARS-CoV-2 was negative.

Medical history, clinical assessment and functional testing
Assessed data include epidemiological and demographic parameters, medical history and potential risk factors, clinical course—including all diagnostic results of the present medical attendance—and household and family constellation. A complete list of all items is attached in online supplemental appendix table E1.

Data are collected at first contact and during the follow-up visits. Symptoms of post-COVID-19/long-COVID are documented and symptoms of myalgic encephalomyelitis/chronic fatigue syndrome are assessed with the Canadian consensus criteria, Chalder Fatigue Scale and PedsQL Multidimensional Fatigue Scale.

Loss of smell and taste are assessed with the ‘U-Sniff’ test, a 12-item odour identification, the ‘Sniffin’ Sticks’ olfactory threshold test and taste samples for sweet, sour, salty and bitter tastes in children aged 6 years or older.

For adults, health status and quality of life are assessed with the St George’s Respiratory Questionnaire and health status and mental health are evaluated with the 9-Item Patient Health Questionnaire (PHQ-9) and the Post-traumatic Stress Disorder Checklist for DSM-5 (PCL-5) questionnaires. For children, quality of life is assessed using the Revised questionnaire to assess Health-Related Quality of Life in children and adolescents (KINDL).

Disease severity is classified according to clinical features using the criteria outlined in the WHO COVID-19 clinical management guideline as asymptomatic, mild, moderate, severe or critical disease. Also, clinical progression is classified according to the WHO clinical progression scale. Applied classification scales are shown in tables 2 and 3.

Functional testing, including lung function testing and MBW, will be conducted at the follow-up visits after 2 weeks and 4–6 months.

Patient and public involvement
We will disseminate all findings in an appropriate and understandable manner to all participants, including children. We welcome the collaboration of participants and public in the interpretation and dissemination of all findings.
Sample collection

Samples will be collected from SARS-CoV-2 positive participants at each of three time points, directly after the diagnosis and at follow-up visits after 2 weeks and 4–6 months, and once from healthy age-matched controls.

Nasal and pharyngeal swab samples are collected for SARS-CoV-2-PCR, scRNAseq and establishment of air–liquid cell cultures. In addition, PBMCs are collected for single-cell sequencing, whole blood for mass cytometry and plasma and serum for mass-spectrometry-based proteomics, and SARS-CoV-2-specific antibody testing.

Study database

For Pa-COVID-19, a study protocol was established that harmonises clinical, molecular and immunological phenotyping assessment in COVID-19 patients. All data are added to an electronic case report form (SecuTrial). Participants included in RECAST are part of Pa-COVID-19. All participants are assigned a pseudonym consisting of a six-digit or seven-digit alphanumerical participant code. A separate log allows to match each participant and their code. Access to SecuTrial requires a username and password. All local data are secured by password.

Table 2  COVID disease severity

<table>
<thead>
<tr>
<th>Patient state</th>
<th>Descriptor</th>
<th>Score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Uninfected</td>
<td>Uninfected; no viral RNA detected</td>
<td>0</td>
</tr>
<tr>
<td>Ambulatory: mild disease</td>
<td>Asymptomatic; viral RNA detected</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Symptomatic; independent</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>Symptomatic; assistance needed</td>
<td>3</td>
</tr>
<tr>
<td>Hospitalised: moderate disease</td>
<td>Hospitalised; no oxygen therapy</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>Hospitalised; oxygen by mask or nasal prongs</td>
<td>5</td>
</tr>
<tr>
<td>Hospitalised: severe disease</td>
<td>Hospitalised; oxygen by non-invasive ventilation (NIV) or high flow</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>Intubation and mechanical ventilation, pO2/pFiO2≤150 or SpO2/FiO2≥200</td>
<td>7</td>
</tr>
<tr>
<td></td>
<td>Mechanical ventilation, pO2/pFiO2&lt;150 (SpO2/FiO2&lt;200) or vasopressors</td>
<td>8</td>
</tr>
<tr>
<td></td>
<td>Mechanical ventilation, pO2/pFiO2&lt;150 and vasopressors, dialysis or extracorporeal membrane oxygenation</td>
<td>9</td>
</tr>
<tr>
<td>Dead</td>
<td>Dead</td>
<td>10</td>
</tr>
</tbody>
</table>

Abbreviated criteria for COVID-19 disease severity according to WHO COVID-19 Clinical management guideline.44

FiO2, fractional inspired oxygen; pO2, partial pressure of oxygen.

Table 3  WHO clinical progression scale

<table>
<thead>
<tr>
<th>Disease severity</th>
<th>Definition</th>
<th>Criteria</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asymptomatic</td>
<td>Symptomatic patients meeting the case definition for COVID-19 without evidence of viral pneumonia or hypoxia.</td>
<td></td>
</tr>
<tr>
<td>Mild</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Moderate</td>
<td>Pneumonia</td>
<td>Clinical signs of (non-severe) pneumonia&lt;br&gt; Adolescent or adult: fever, cough, dyspnoea, fast breathing&lt;br&gt; Child: cough or difficulty breathing + fast breathing and/or chest indrawing&lt;br&gt; Diagnosis can be made on clinical grounds; chest imaging (radiograph, CT scan, ultrasound) may assist in diagnosis and identify or exclude pulmonary complications.</td>
</tr>
<tr>
<td>Severe</td>
<td>Severe pneumonia</td>
<td>Adolescent or adult: plus one of the following: respiratory rate &gt;30 breaths/min; severe respiratory distress; or peripheral oxygen saturation (SpO2) &lt;90% on room air&lt;br&gt; Child: plus at least one of the following: Central cyanosis or SpO2&lt;90%; severe respiratory distress (eg, fast breathing, grunting, very severe chest indrawing); general danger sign: inability to breastfeed or drink, lethargy or unconsciousness, or convulsions.</td>
</tr>
<tr>
<td>Critical disease</td>
<td>Acute respiratory distress syndrome or sepsis/septic shock</td>
<td>Oxygenation impairment, invasive ventilation or bilevel non-invasive ventilation (NIV) or continuous positive airway pressure (CPAP) (≥ 5 cmH2O) required or Infection and ≥2 systemic inflammatory response syndrome criteria</td>
</tr>
</tbody>
</table>

Modified from WHO working group.45
Patients recruited in RECAST are grouped into six age categories (Table 4). Due to the nature of observatory studies and the lack of pre-existing data, it is not possible to predict the extent of assumptive differences. Preliminary findings suggest that for most planned analyses a sample size of 15 is sufficient. The outlined sample sizes should suffice even for comparisons between children of different age groups.

### Sample description

**Table 4  Age categories of the RECAST participants**

<table>
<thead>
<tr>
<th>Age group</th>
<th>Disease state</th>
<th>No</th>
<th>Time points</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Children</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nursery (0–3 years)</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td>Kindergarten (3–6 years)</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td>Primary school (6–12 years)</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td>Secondary school (13–18 years)</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td><strong>Adults</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;60 years</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td>&gt;60 years</td>
<td>SARS-CoV-2−</td>
<td>≥30</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>SARS-CoV-2+</td>
<td>≥30</td>
<td>3 (days 0, 14, 180)</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td></td>
<td>≥360</td>
<td>≥ 720</td>
</tr>
</tbody>
</table>

RECAST, RESilience of Children compared to Adults in SARS-CoV-2 infection.

**PLANNED ANALYSES AND OUTCOMES OF INTEREST**

We propose a multiomics workup for all patients. A synopsis of the planned analyses is depicted in Figure 1.

**SARS-CoV-2-specific PCR and antibody testing**

All participants are screened for an active SARS-CoV-2 infection with Real Time Quantitative PCRs (RT-qPCRs) targeting E and N genes. Antibody testing is conducted for all serum and saliva samples with SARS-CoV-2-specific IgG-ELISAs and IgA-ELISAs. In case of a reactive screening result, confirmatory testing with a recombinant immunofluorescence assay and a plaque reduction neutralisation test are conducted.

**Figure 1** Performed analyses in the RECAST study. The RECAST study consist of eight elements, including the anamnesis and clinical data, a comprehensive multiomics workup of the immune landscape of the blood and the upper airways and lung function testing. CyTOF, cytometry by time-of-flight; RECAST, RESilience of Children compared to Adults in SARS-CoV-2 infection.
Nasal epithelial culture and functional in vitro analyses
Conditional reprogramming allows for the generation of long-term cultures of primary airway epithelial cells. Without the need of genetic modification or clone selection, conditional reprogramming enables cell expansion, while redifferentiated cultures retain their organ-specific phenotype. We establish highly differentiated polarised in vitro air–liquid interface cultures that reproduce and allow for the analysis of physiological in vivo conditions, such as heterogeneous cell composition with preserved lineage as well as functional characteristics, including production of airway surface liquid and mucociliary clearance. For material collection, FLOQswabs (Copan, Italy) are used. Swabs are transferred into DMEM/F12 medium (Gibco, USA) and transported to our laboratory within 2 hours.

Mass cytometry of whole blood samples
Whole blood is fixed with a proteomic stabiliser for preservation of surface and intracellular markers. Blood samples are stored at −80°C until batch-based analysis. Thawed samples are stained in batches of nine patient and one anchor reference sample. On barcoding of individual samples, they are pooled and stained with metal isotype conjugated anti-human antibodies as described previously. CyTOF technology allows for the detection of more than 40 different barcodes simultaneously to identify cell populations in a high-throughput setting.

Mass spectrometry-based serum and plasma proteomics
A platform technology with semiautomated sample preparation to allow for ultra-high-throughput liquid chromatography and mass-spectrometry based analyses of the proteome has recently been established by members of our group. In a directed approach, we characterise the immune response-related serum and plasma proteome, with focus on the acute phase response and the complement system. However, the plethora of proteome signals that are generated per sample also allows for an undirected approach, delivering predictive proteome signatures. To facilitate the computation of such extensive bulk data, a deep neural network is employed.

Single-cell sequencing of nasal epithelial cell samples and PBMCs
The nasopharynx is the entry point for an infection with SARS-CoV-2 and as such of distinguished concern in the exploration of the individual immune response pattern. Using scRNAseq of nasal and bronchial samples, we were previously able to identify cell types and states that correlate with a severe disease course of COVID-19. Here, scRNAseq will be applied to define the composition and transcriptional activity of immune and epithelial cells in the nasal environment of children and adults throughout the various states of SARS-CoV-2 infection. Nasal swabs (FLOQswabs, Copan, Italy) are used for sample collection. Following sample collection, swabs are directly transferred into cold Gibco Dulbecco’s Modified Eagle Medium: Nutrient Mixture F12 (DMEM/F12) (Gibco, USA) and transported to our biosafety laboratory within 1 hour for further processing. Library preparation is performed according to manufacturer’s protocol (10×) and sequencing is performed using the Illumina NextSeq 6000 platform.

Lung function testing and MBW
SARS-CoV-2 infections cause severe lung damage in adults. In a large review with 2135 children, 45% (951 children) were classified with a severity of moderate, severe or critical, all with lung involvement per definition. There is evidence that children with acute lung injury experience the same lung pathologies as adults. To assess the extent of transient and permanent functional lung impairment, we investigate the lung function with spirometry and MBW. MBW measures the lung ventilation homogeneity. This technique is already feasible without sedation in children from 2 years of age. Spirometry depends on the cooperation of the participant and may usually be conducted with children aged 6 years or older. The technical MBW procedures are in accordance with the American Thoracic Society Technical Statement. Measurements are conducted by personnel certified according to European Cystic Fibrosis Society–Clinical Trial Network (ECFS-CTN certified) and for study measurements Exhalizer D (Ecomedics, Dürrenden, Switzerland) will be used. N2 wash-out is used as tracer gas to determine the lung clearance index (LCI) as an outcome measure. The LCI increases with lung ventilation inhomogeneity.

Biobanking
Collected material is processed and stored at the central biobank of Charité (ZeBanC, https://biobank.charite.de). Material that is not immediately used is subjected to cryopreservation.

Ethics and dissemination
All procedures in this study are in compliance with the principles of the 1964 Declaration of Helsinki and its amendments. We act in adherence to the principles of Good Clinical Practice (International Council for Harmonisation, 1996). The study was reviewed and approved by the Charité Ethics Committee (EA2/066/20). All participants enrolled give written informed consent in person, for participants minor of age the written informed consent of the legal guardian is also required.
Study procedures never interfere with the medical management of participants. Samples required for medical management always have priority. There is no direct benefit for patients participating in the study. Results from the study might improve our understanding of the disease and benefit the public health.

Data are monitored regularly. Informed consent forms are audited by a monitor appointed by the Charité Clinical Trial Management Unit. Data monitoring of collected data is performed in the course of the study. As established for Pa-COVID, 24 we reiterate the fundamental principle in this study that all contributors and researchers who have access to samples commit to unrestricted data sharing. In accordance with FAIR data principles, all data collected shall be findable, accessible, interoperable and reusable. 70

Results originating from nasal epithelial culture and functional in vitro analyses, mass cytometry, mass spectrometry-based proteomics, single-cell sequencing and lung function testing and MBW as well as clinical data will be will be disseminated separately or in context in a variety of ways including abstracts, posters and presentations at conferences and published manuscripts in peer-reviewed journals. As soon as all analyses are completed, a comprehensive review will be published to put the findings in context of each other.

Discussion
The SARS-CoV-2 pandemic has accelerated scientific research in the field of virology and related immunobiology for nearly 2 years, yet many crucial questions remain unanswered. Soon after the emergence of the virus it became apparent that, while children are just as likely to be infected with SARS-CoV-2 as adults, they are less severely affected. 4–8, 71 RECAST is an observational study that aims to elucidate the differences between children and adults in primary SARS-CoV-2 infections using a multiomics approach. Revealing age-dependent differences will help to develop better suited therapeutics and vaccination strategies beyond SARS-CoV-2 infections.

Previous multiomics approaches conducted with specimen from adult donors served to elucidate the immune response in COVID-19 62, 72 to identify predictors of severe disease courses 73–74 and to isolate possible targets for therapy. 62, 74, 75 Multiomics-based studies focusing on SARS-CoV-2 infections in children remain rare and are limited to small participant numbers and only analyse a limited number of multiomics dimensions: A study including 24 infected children analysed the single-cell transcriptional landscape in the upper airways 76; with single-cell multiomic profiling of matched nasal, tracheal, bronchial and blood samples of 19 infected children, a study characterised the immune landscape with focus on the upper airways 77; the plasma proteomic and metabolomic data of 18 infected children was analysed in another study 78; clinical characteristics and serum markers were analysed in a larger group that summarised children and young adults and T cell response in a paediatric subgroup of 11 participants was examined 79; and a study with 24 infected children analysed the T cell response and specific antibody response. 80 Even though these studies contributed greatly to a better understanding of age-related immune response patterns in COVID-19, there is still a substantial demand for research. Especially studies analysing the immune response over the whole age and severity spectrum applying a multiomics approach are needed. In addition, mechanistic investigations, revealing the causal relationship between the different immune defence layers, are missing. In RECAST, we will conduct a full multiomics workup with at least 120 infected children, a larger number of participants than in previously published multiomics studies. Moreover, we will conduct follow-up visits for 6 months to profile the development of age-specific immune response patterns over the course of time. Biobanking and long-term storage of samples will be used to perform subsequent mechanistic studies on first data collection and hypotheses formulation. The recruitment of family members, both infected and non-infected, allows to assess the effect of genetic relationships. The combination of high resolution multidimensional immunological methods with clinical endpoints in the RECAST study will enable us to contribute to the understanding of the increased resilience of children to SARS-CoV-2 infections.

Author affiliations
1Department of Pediatric Respiratory Medicine, Immunology and Critical Care Medicine, Charité Universitätsmedizin Berlin, Berlin, Germany
2Karsten, Hugo, Wagner, Paediatric Practice, Berlin, Germany
3Boeckel, Haverkamper, Paediatric Practice and Practice for Paediatric Cardiology, Berlin, Germany
4Stich-Boeckel, Paediatric Practice, Berlin, Germany
5Maske, Pan kok, Paediatric Practice, Berlin, Germany
6Clinical Study Center (CSC), Berlin Institute of Health at Charité, Berlin, Germany
7Department of Paediatric Neurology, Charité Universitätsmedizin Berlin, Berlin, Germany
8Center for Chronically Sick Children (Sozialpädiatrisches Zentrum, SPZ), Charité Universitätsmedizin Berlin, Berlin, Germany
9Center for Digital Health, Berlin Institute of Health at Charité, Berlin, Germany
10Molecular Epidemiology Unit, Berlin Institute of Health at Charité, Berlin, Germany
11German Center for Lung Research, Giessen, Germany
12Department of Infectious Diseases and Respiratory Medicine, Charité Universitätsmedizin Berlin, Berlin, Germany
13Department of Biochemistry, Charité Universitätsmedizin Berlin, Berlin, Germany
14Molecular Biology of Metabolism Laboratory, The Francis Crick Institute, London, UK
15Institute of Virology, Charité Universitätsmedizin Berlin, Berlin, Germany
16Berlin Institute of Health, Berlin, Germany
17Institute of Medical Immunology, Charité Universitätsmedizin Berlin, Berlin, Germany
18Clinical Study Center (CSC), Berlin Institute of Health at Charité, Berlin, Germany
19Institute of Medical Immunology, Charité Universitätsmedizin Berlin, Berlin, Germany
20Institute of Medical Immunology, Charité Universitätsmedizin Berlin, Berlin, Germany
21Open access

Acknowledgements
We thank participating children and their families for their close collaboration. We also thank all outpatient pediatrics who are involved in the recruitment.

Contributors
BS, JR, MAM, VMC, MR and LES initiated the project and led the BMBF grant proposal. All authors contributed to the design of the study. SS, JR, NZ, MK, TB, HS-B, JM and ER are collecting data biological material and are conducting functional testing. BS, MAM, VMC, MR, LES, IL, RE, JR and VS lead their respective research field and supervise the conduction of experiments and the interpretation of results. CD-H, PMB and AB are conducting experiments. SS and JR produced the first draft of the protocol. All authors provided critical review of the manuscript and have approved the final version.

Funding This work was supported by the German Federal Ministry of Education and Research (grant number 01KI20337).

Competing interests None declared.

Patient and public involvement Patients and/or the public were involved in the design, conduct, or reporting, or dissemination plans of this research. Refer to the Methods section for further details.

Patient consent for publication Not applicable.

Provenance and peer review Not commissioned; externally peer reviewed.

Supplemental material This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and makes no representation as to the currency, completeness, or accuracy of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

ORCID iDs

Sebastian Stricker http://orcid.org/0000-0001-9129-8997
Niklas Ziegahn http://orcid.org/0000-0003-4195-8584
Johst Roehmel http://orcid.org/0000-0002-1535-8852

REFERENCES