ABSTRACT

Introduction Type 2 diabetes (T2D) is characterised by elevated plasma glucose, free fatty acid (FFA) and insulin concentrations, and this metabolic profile is linked to diabetic cardiomyopathy, a diastolic dysfunction at first and increased cardiovascular disease (CVD) risk. Shifting cardiac metabolism towards glucose utilisation has been suggested to improve cardiovascular function and CVD risk, but insulin treatment increases overall glucose oxidation and lowers lipid oxidation, without reducing CVD risk, whereas SGLT2 inhibitors (SGLT2i) increase FFA, ketone body concentrations and lipid oxidation, while decreasing insulin concentrations and CVD risk. The aim of the present study is to elucidate the importance of different metabolic profiles obtained during treatment with a SGLT2i versus insulin for myocardial function in patients with T2D.

Methods and analyses Randomised, crossover study, where 20 patients with T2D and body mass index >28 kg/m² receive 25 mg empagliflozin daily or NPH insulin two times per day first for 5 weeks followed by a 3-week washout before crossing over to the remaining treatment. Insulin treatment is titrated to achieve similar glycaemic control as with empagliflozin. In those randomised to insulin first, glycaemia during an initial empagliflozin run-in period prior to randomisation serves as target glucose. Metabolic and cardiac evaluation is performed before and at the end of each treatment period.

The primary endpoint is change (treatment—washout) in left ventricular peak filling rate, as assessed by cardiac MRI with and without acute lowering of plasma FFAs with acipimox. Secondary and explorative endpoints are changes in left atrial passive emptying fraction, left ventricular ejection fraction, central blood volume and metabolic parameters.

STRENGTHS AND LIMITATIONS OF THIS STUDY

⇒ Comparison with NPH insulin, which has opposite metabolic effects to empagliflozin, provides a strong basis for detecting metabolic effects on cardiac function.

⇒ Repeated cardiac MRI during depletion of plasma free fatty acids (FFAs) with acipimox during treatments and washouts allows for dissection of the individual roles of hyperglycaemia, hyperinsulinaemia and elevated FFAs on cardiac function in type 2 diabetes (T2D).

⇒ A crossover design is vulnerable to dropout, but provides greater statistical power.

⇒ Effects of metabolic changes on cardiac function are limited to the 5-week intervention period, which excludes effects arising from longer-term treatment.

INTRODUCTION

Type 2 diabetes (T2D) is characterised by hyperglycaemia, hyperinsulinaemia, increased free fatty acids (FFAs) and impaired tissue glucose uptake and oxidation.1 T2D is associated with an increased cardiovascular morbidity, and the more dysregulated the metabolic state, the greater...
the cardiovascular risk. T2D develops when insulin secretion can no longer compensate for the ambient insulin resistance, and therefore previous treatments has focused on increasing insulin signalling by either exogenous insulin administration, stimulation of endogenous insulin secretion or enhancing insulin sensitivity.

Diabetic cardiomyopathy (DCM) is an early ‘silent’ complication to T2D, independent of hypertension and/or coronary heart disease. It is characterised by left ventricular (LV) hypertrophy and diastolic dysfunction and has been linked to the increased cardiovascular risk in T2D. DCM may be accurately described by measuring left ventricular peak filling rate (LVPFR) and left ventricular ejection fraction (LVEF) using cardiac MRI (CMR). Both diastole and systole are energy requiring processes and sensitive to changes in energy availability. Interestingly, cardiac metabolism in patients with T2D is altered and depends more on lipid oxidation and less on glucose oxidation compared with non-diabetic controls. It has been argued that glucose oxidation is a better source of energy for the heart than lipid oxidation, especially during stress, such as myocardial ischaemia, because this yields more adenosine triphosphate unit oxygen. However, manipulating cardiac metabolism towards glucose oxidation, by administering glucose–insulin (–potassium) infusions in patients with hyperglycaemia and myocardial infarction has been attempted, but did not improve survival in neither diabetic nor non-diabetic patients. In intensive care unit patients, strict glycemic control using insulin has been associated with increased mortality, and in patients with T2D and increased cardiovascular disease (CVD) risk, intensive glycemic control has not reduced CVD risk compared with conventional glycemic control and in the ACCORD Study, which involved aggressive insulin treatment resulted in excess mortality. Thus, insulin treatment does not prevent cardiovascular events in patients with T2D nor improve prognosis when such occur.

SGLT2 inhibition (SGLT2i), on the other hand, is a newer treatment principle in T2D, which has proven effective in attenuating the risk of myocardial infarctions, worsening of heart failure, cardiovascular mortality and all-cause mortality in patients with T2D. SGLT2i increases renal glucose excretion thereby lowering plasma glucose (PG) and insulin levels and increasing glucagon release, lipolysis and ketogenesis. Additionally, tissue glucose uptake and oxidation is reduced and hepatic glucose production is increased. The exact cardioprotective mechanisms of SGLT2i are not yet understood, but has been proposed to be linked to improved haemodynamics, inhibition of myocardial Na+/H+ exchange or reductions in inflammatory activity.

An early and interesting hypothesis proposed that changes in cardiac metabolism may be responsible for the cardioprotective effect of SGLT2i. The lowered glucose and insulin concentrations, persistent hyperketonaemia and elevated FFAs, caused by SGLT2i treatment, leads to reduced glucose uptake, increased ketone body uptake and oxidation and unchanged uptake of FFAs in the heart while overall lipid oxidation is increased. This altered energy metabolism may rapidly improve myocardial function, and during myocardial stress. The SGLT2i-induced myocardial fuel switch from glucose to fatty acids and ketone bodies has been suggested to ameliorate adverse cardiac remodelling and heart failure in non-diabetic porcine models, and it is noteworthy that eliminating the availability of FFAs to insulin resistant hearts can lead to cardiac dysfunction in rodents and in humans, suggesting an important role for lipid metabolism in cardiac function. Cardiovascular endpoint trials with SGLT2 inhibitors have shown effects within weeks after initiation of treatment, coinciding with the metabolic effects of the treatment.

Altogether, SGLT2 inhibitors ‘amplify’ some components of the dysmetabolic profile of T2D and works opposite the metabolic effects of insulin. This raises the question of how cardiac function in patients with T2D depends on lipid and glucose oxidation in the resting state and during stress, and how increasing or lowering blood glucose, FFAs, ketone bodies and insulin concentrations influence cardiac function.

**Objective**

The primary objective of the present study is to evaluate myocardial function in patients with T2D and high risk of CV events using advanced CMR scans during rest, chronotropic stress and under depletion of plasma FFAs before and after 5 weeks of empagliflozin treatment (high FFA and ketone body concentrations, high lipid oxidation and low insulin concentrations) and before and after 5 weeks of human insulin treatment titrated to yield glycaemic control similar to the empagliflozin treatment period (low FFA and ketone body concentrations, high insulin concentrations and glucose oxidation).

**Hypothesis**

We hypothesise that hyperinsulinaemia and hyperglycaemia are conditions that negatively affect cardiac function in T2D, while the availability of FFAs and ketone bodies and switching metabolism towards lipid oxidation improves cardiac diastolic and systolic function. Thus, we expect that lowering PG insulin independently, and increasing fatty acid concentrations, lipid oxidation and ketone body availability with empagliflozin treatment, improves myocardial function in patients with T2D, and that depleting plasma of FFAs during empagliflozin treatment will impair cardiac function.

**METHODS AND ANALYSES**

**Design**

This is a 20-week prospective, investigator-initiated, comparator-controlled, open-label, two-arm crossover, human study where subjects are randomised in blocks...
of 3–5 to Neutral Protamine Hagedorn (NPH) insulin twice daily or empagliflozin treatment (25 mg once daily) for 5±1 weeks, followed by 3±1 weeks washout and crossover of treatment for 5±1 weeks (figure 1). For 7 weeks preceding randomisation, but after inclusion, patients undergo a programme of 2 weeks of washout of pre-existing antihyperglycaemic treatment (except metformin), 2 weeks of empagliflozin run-in (used for glycaemic target and titration of treatment in participants randomised to insulin first, see below) followed by 3 weeks of washout. During run-in and treatment periods, participants measure blood glucose two times per day (fasting and before evening meal) and during washouts patients measure fasting blood glucose.

After the screening visit (V0) there are four study visits (V1–4)—before and at the end of each treatment period. Each visit consists of three study days—a metabolic study day (MET) and two CMR study days. Randomisation is performed at V1 after the metabolic study day.

Participants
Twenty subjects older than 18 years diagnosed with T2D, a body mass index ≥ 28 kg/m², glycated haemoglobin ≤ 9%, fasting C-peptide > 500 pmol/L and unchanged antihyperglycaemic treatment for 12 weeks prior to screening, and who are at a risk of CVD, are eligible for the study. High CVD risk is modified from the EMPA-REG protocol.46 Inclusion, exclusion and withdrawal criteria are listed in box 1.

Recruitment
Participants are recruited from the Department of Endocrinology and Cardiology at Hvidovre Hospital and are identified by reviewing laboratory results and patient files. Potential participants will be contacted by means of a recruitment letter, in which they are informed of the opportunity to participate in a scientific research project. We also will advertise for participant in local newspapers and on the internet as well as social media (eg, wwww.forsøgsperson.dk; www.sundhed.dk and www.facebook.com).

Outcomes
The primary outcome is change in myocardial diastolic function. This was chosen because first, diastole is a highly energy requiring process,11 12 and second, because diastolic dysfunction (with or without LV hypertrophy) is the notable early manifestation of DCM.47 Thus, if changes in overall energy metabolism are to affect cardiac function in patients with T2D, it may well occur in diastole at the earliest. Diastolic cardiac function can be accurately assessed using CMR by measuring LVPFR and LAPEF.48 Our primary outcome measure is change (LVPFR treatment – LVPFR washout) in LVPFR (ΔLVPFR). All endpoints are listed in table 1.

Randomisation and intervention
Participants are randomised consecutively by lottery in blocks of 3–5 to treatment with either subcutaneous NPH insulin (Insulatard) two times per day or oral empagliflozin (Jardiance) 25 mg once daily first. All patients will receive both treatments during the trial. Randomisation is performed at V1. NPH insulin is initiated at a dose of 0.2 IU/kg body weight/day and is titrated daily over phone (phone contacts, figure 1) by 0.05 IU/kg body weight/day until average blood glucose (BG) over three consecutive days is within ±1 mmol/L of the individual glycaemic target. In participants randomised to insulin first, the glycaemic target is average fasting and evening glucose concentrations during the second week of empagliflozin run-in. In patients randomised to insulin second, the glycaemic target is average fasting and preprandial evening BG values of week 3 and 4 during the first (empagliflozin) treatment period.

As previously discussed, insulin and empagliflozin represents two metabolically opposing methods for lowering PG concentrations. By titrating insulin treatment...
Box 1  Eligibility criteria

**Inclusion criteria**
- Age ≥18 years
- Body mass index ≥28 kg/m²
- Glycated haemoglobin <9% (<10% in diet or metformin treated only)
- Fasting C-peptide ≥500 pmol/L
- Unchanged glycaemic treatment for 3 months prior to inclusion
- High cardiovascular risk as one of the following:
  - Previous myocardial infarction, stroke or peripheral arterial disease more than 2 months prior to informed consent
  - Evidence of multivessel coronary arterial disease (CAD) but without prior myocardial infarction, if more than 50% stenosis is present, if revascularised (Coronary Artery Bypass Graft (CABG) or Percutaneous Coronary Intervention (PCI)) more than 2 months prior or if one vessel is vascularised and the other with 50% stenosis
  - Single vessel CAD without prior myocardial infarction if more than 50% stenosis is present, not revascularised and positive stress test for ischaemia

**Exclusion criteria**
- Insulin treatment within 3 months from informed consent
- Type 1 diabetes
- Psychiatric disorder or mental retardation
- Drug or alcohol abuse within 3 months from informed consent
- Poor compliance
- Anaemia (haemoglobin <103.1 g/L) or other blood dyscrasias causing haemolysis or unstable erythrocytes
- Indication of liver disease (Alanine transferase or alkaline phosphatase elevation above three times the upper normal limit)
- Impaired renal function (estimated glomerular filtration rate <45 mL/min/1.73 m²)
- Antiobesity medication within 3 months from informed consent
- Systemic steroid treatment within 6 months from informed consent
- Poor compliance
- Bariatric surgery or other gastrointestinal conditions that may compromise gastrointestinal absorption
- Peptic ulcer—verified endoscopically
- Any form of surgery within 3 months of informed consent
- Acute myocardial infarction, stroke or peripheral arterial disease within 2 months of informed consent
- Persistent or permanent atrial fibrillation

Inability to undergo experimental procedures including exclusion criteria for cardiac MRI scanning:
- Implantable cardioverter defibrillator/pacemaker
- Ferromagnetic clips
- Claustrophobia

Contraindication to glycopyrrolate infusion:
- Known closed-angle glaucoma known severe prostate hyperplasia
- Tachycardia (HR >100 at rest)
- Known bladder atony
- Cardiac insufficiency or non-congenital pylorus stenosis—verified endoscopically
- Known gastroparesis

Contraindications to adenosine:
- Second or third degree atioventricular block
- Severe hypotension (blood pressure ≤90/50 mm Hg)
- Long QT syndrome
- Unstable angina pectoris
- Decompensated heart failure

Continued

Box 1  Continued

- Sinus node dysfunction
- Chronic obstructive pulmonary disease or asthma bronchiale (forced expiratory volume in one second ≤50% of expected)
- Allergy towards any of the drugs or diagnostics used in the protocol (insulin, empagliflozin, acipimox, glycopyrrolate, adenosine, gadolinium contrast enhancer)
- Any condition, which in the opinion of the investigator, may jeopardise subject safety or compliance with the protocol.

**Withdrawal criteria**
- Subjects may withdraw from the study without any notice or reason
- Pregnancy discovered during the experiment
- Unacceptable adverse reactions or reactions associated with the planned experiments, including severe glycaemic dysregulation during washout periods

to match the glycaemic control found with empagliflozin in the same participants, the result is two distinct metabolic phenotypes: one with hyperinsulinaemia and suppressed levels of FFAs (NPH insulin treatment), and one with reduced insulin levels and increased levels of FFAs (empagliflozin treatment)—but both with the same levels of glycaemic control (figure 2). NPH insulin has been chosen over more modern human insulin analogues, as it is not albumin bound and can be measured in an ordinary insulin assay.

**Safety considerations**
During washout periods, blood glucose concentrations will increase—that is a separate point of the study, but severely dysregulated diabetes is an exclusion criterium to ensure participant safety. The risk of severe hyperglycaemia is reduced in several ways in the study:
- Existing metformin treatment is continued throughout the whole study as background antglycaemic treatment.
- In case of fasting BG concentrations of more than 13 mmol/L, patients are instructed to contact study personnel.
- Phone contacts by study investigator are planned in the second week of washout periods to follow-up on the patient and enquire to hyperglycaemic events or other adverse events.
- As soon as the final day (CMR with acipimox) of a washout visit (visit 1 or 3) is completed, antglycaemic treatment according to study drug sequence is commenced to minimise time spent in hyperglycaemia.

In case of fasting BG >13 mmol/L, the patient will be contacted daily for two additional days. If average fasting BG over the 3 days >13 mmol/L that triggers an extra safety visit, where fasting PG is measured. If PG >13 mmol/L on the day of the extra visit, then the patient is withdrawn from the study and antihyperglycaemic treatment is initiated.

### Screening visit (V0)

Once oral and written informed consent is obtained by the study investigator, the screening procedure follows. Medical history is recorded, screening blood samples drawn and an ECG, recording of blood pressure, pulse rate and registration of anthropometric data are performed, and patients are screened according to inclusion and exclusion criteria. A standard transthoracic echocardiography is performed, and oxygen consumption (VO₂)max is estimated (table 2).

### Study visits

All study visits consist of three study days—a metabolic study day and two CMR study days (table 3).

### The metabolic study day

The metabolic study is conducted at the Department of Endocrinology, Hvidovre Hospital, to document the metabolic effects of each study drug.

Participants meet in the morning after an overnight fast. Anthropometric data, blood pressure, pulse rate and an ECG are recorded, and two catheters, one in each arm are inserted for infusion of tracers and for repeated drawing of arterialised blood samples, respectively. Baseline and safety blood samples are taken (table 4), the participant empties bladder and the investigational drug (V2, V4) and the participants’ usual medications are administered at 08:00 hours. Body composition is determined by dual energy X-ray absorptiometry scan.

#### Basal metabolism

Primed infusions of stable glucose ([6,6-D₂]-glucose) and glycerol ([1,1,2,3,3-D₅]-glycerol) tracers are initiated (T=−180 min). Blood is sampled at −30 to −15 and −2 min to characterise glucose, lipid and amino acid metabolism. The patient empties bladder, urine is weighed and samples are taken for determination of tracer concentrations and urinary nitrogen excretion, and the 5-hour oral glucose tolerance test (OGTT) is initiated.

#### Five-hour OGTT

The patient ingests anhydrous glucose (72 g) with added [U-13C₆]-glucose tracer (3 g) dissolved in 250 mL of water over 5 min (T=0 min). Intravenous tracer infusions continue unchanged. Blood is sampled regularly for 5 hours for characterisation of postprandial glucose, lipid and amino acid metabolism (tables 3 and 4). The patient empties bladder regularly during and at the end of the OGTT. Urine is sampled for nitrogen excretion and tracers/tracees.

Fat and muscle biopsies: biopsies are obtained during the basal (T=−60 min) and the maximally insulin stimulated (T=60 min) state. Muscle biopsies are considered proxies for cardiomyocyte metabolic status. Overall, 30 min ventilated hood indirect calorimetry (Vyaire Vyntus CPX) is performed during the basal period (prior to biopsies) t=−90 min and postprandially at t=60 min for determination of fasting and postprandial energy expenditure and respiratory quotient.

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**Table 1** Endpoints

<table>
<thead>
<tr>
<th>Primary end point</th>
<th>Change in left ventricular peak filling rate (ΔLVPFR)</th>
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<tbody>
<tr>
<td>Secondary endpoints</td>
<td>Change in left atrial passive emptying fraction (ΔLAPEF)</td>
</tr>
<tr>
<td></td>
<td>Change in left ventricular ejection fraction (ΔLVEF)</td>
</tr>
<tr>
<td>Explorative endpoints include</td>
<td>Cardiovascular:</td>
</tr>
<tr>
<td></td>
<td>Change in oxygen consumption max and exercise tolerance test variables</td>
</tr>
<tr>
<td></td>
<td>Change in central blood volume and haematocrite</td>
</tr>
<tr>
<td></td>
<td>Change in heart rate variability</td>
</tr>
<tr>
<td></td>
<td>Change in left ventricular volume</td>
</tr>
</tbody>
</table>

| Metabolic: |
| Basal and postprandial AUC free fatty acids and glycerol turnover |
| Endogenous glucose production and tissue glucose disposal (metabolic clearance of glucose) |
| Fasting and postprandial energy expenditure and respiratory quotient Glucagon–insulin ratio |
| Insulin sensitivity (AUC glucose metabolic clearance/AUC insulin concentration) |
| Beta-cell function (prehepatic insulin secretion rate, correlated to ambient glucose) |

AUC, area under the curve.

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**Figure 2** Metabolic profile of the two study drugs. Schematic representation of the metabolic changes expected with the two study drug treatments in a patient randomised to insulin first. Insulin treatment is characterised by low glucose, low free fatty acids (FFAs) and high insulin concentrations; empagliflozin treatment by low glucose, high FFAs and low insulin.
Exercise test (50% VO$_2$max)

At T=300 min, the participant is exercised at 60 W for 4 min after which work load is increased until oxygen consumption is 50% of estimated VO$_2$max. Pulse rate is recorded with a chest mounted pulse rate monitor and oxygen consumption is 50% of estimated VO$_2$max. Pulse rate is steady for 2 min, oxygen consumption and pulse rate are recorded, and the test is stopped. VO$_2$max is estimated by linear extrapolation to the theoretical maximal pulse rate (220-age).  

Ad libitum meal test

SGLT2 inhibition is associated with a lower weight reduction than predicted from the urinary energy loss. SGLT2 inhibition does not change resting energy expenditure or blunt the thermogenic effect of feeding, suggesting that energy intake is increased. Therefore, the metabolic study day is ended with an ad libitum meal, consisting of thoroughly mixed pasta bolognese (fixed nutrient composition and energy content). Patients are placed in a quiet corner and instructed to eat until full. Two glasses of water (total 300 mL) are allowed with the meal. The meal is weighed before and after serving and ad libitum meal intake defined as the difference. Throughout the day, patients are asked to score their hunger, satiety and sensation of fullness on a visual analogue scale.

Cardiac evaluation

Two CMR days are performed during each visit (V1–V4). In addition, diurnal blood pressure and Holter monitoring are performed. CMR is conducted at the Department of Cardiology, Rigshospitalet, Copenhagen, whereas Holter monitoring and diurnal blood pressure monitoring are performed at the Department of Cardiology, Hvidovre Hospital. Participants meet fasting and morning medication, including investigational medicinal product (V2 and V4), is administered. Anthropometric data are recorded, and two intravenous catheters are inserted into an antecubital and the contralateral dorsal hand vein for infusion of adenosine, gadolinium contrast and glycopyrrolate and for blood sampling respectively (table 4). Prior to CMR, a transthoracic echocardiography is performed (table 2).

CMR is performed on a 1.5 Tesla scanner (Siemens Aera; Siemens; Erlangen; Germany) with the patient lying supine on the back, using an 18-channel cardiac coil with continuous ECG gating.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>Screening procedures</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood samples</td>
<td>Haematology (haemoglobin, thrombocytes, haematocrit, leucocytes), liver and renal function tests (creatinine, eGFR (Cockroft-Gault formula), alkaline phosphatases, alanine aminotransferases, lactate dehydrogenase, bilirubin, amylase, sodium, potassium), fasting P-glucose, C-peptide, glycated haemoglobin, TSH, urinary albumin/creatinine mass ratio and in fertile women, u-hCG.</td>
</tr>
<tr>
<td>Echocardiography</td>
<td>Parasternal long axis view, parasternal short axis view at aortic, mitral and apex levels, apical four-chamber view, left ventricular ejection fraction (LVEF), E/E', E', LVEDV/BSA.</td>
</tr>
<tr>
<td>Estimation of oxygen consumption (VO$<em>2$)$</em>{max}$</td>
<td>Maximum oxygen uptake is estimated using Åstrøm’s two-point test performed on a cycle ergometer during indirect calorimetry. From measurements of VO$_2$ at two submaximal pulse rates VO$_2$max is estimated by linear extrapolation to the theoretical maximal pulse rate (220-age).</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Table 3</th>
<th>Visit overview</th>
</tr>
</thead>
<tbody>
<tr>
<td>Metabolic study day</td>
<td>Cardiac MRI (CMR)</td>
</tr>
<tr>
<td>▶ Dual energy X-ray absorptiometry scan and fasting safety and efficacy blood samples</td>
<td>▶ Fasting blood samples, before and after CMR.</td>
</tr>
<tr>
<td>▶ Determination of 3-hour basal metabolism.</td>
<td>▶ Echocardiography</td>
</tr>
<tr>
<td>▶ Infusion of glucose and glycerol tracers</td>
<td>▶ CMR rest</td>
</tr>
<tr>
<td>▶ Basal muscle and fat biopsies</td>
<td>▶ Without enhancement</td>
</tr>
<tr>
<td>▶ Basal energy expenditure and determination of respiratory quotient</td>
<td>▶ With enhancement and adenosine infusion</td>
</tr>
<tr>
<td>▶ 5-hour oral glucose tolerance test (OGTT)</td>
<td>▶ CMR stress</td>
</tr>
<tr>
<td>▶ With oral glucose tracer</td>
<td>▶ Unenhanced repeated during pharmacological chronotropic stress with glycopyrrolate infusion</td>
</tr>
<tr>
<td>▶ Continued intravenous glucose and glycerol tracer</td>
<td>▶ 24-hour ambulant blood pressure</td>
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</tbody>
</table>
Cine 2-chamber, 3-chamber and 4-chamber images, complete transverse and short axis cine stacks covering the whole heart are acquired. All images are obtained during end expiratory breath holds.

Myocardial perfusion images during rest and stress are obtained at the basal, mid-ventricular and apical cardiac short-axis level. Rest perfusion images of the myocardium are acquired using an intravenous bolus of gadolinium contrast (Gadovist, Bayer AG, Germany) 0.075 mmol/kg bodyweight. The time of gadolinium contrast entry into the right and the left ventricle is accurately determined, and this transit time of gadovist multiplied by cardiac output is used to calculate the pulmonary and central (pulmonary+cardiac) blood volume.

Myocardial stress perfusion images are obtained with an intravenous dose of 0.075 mmol/kg of gadovist during and 10 min after an intravenous adenosine (140 µg/min) administrated for maximum 4 min). This is followed by evaluation of cardiac function during chronotropic stress, where short axis cine stack will be reacquired 10 min after the administration of intravenous glycopyrrolate (4 µg/kg, max. 400 µg, given as a bolus). This approach has been shown to unmask subclinical diastolic dysfunction as has been demonstrated in normal healthy elderly.48 49

**Cardiac MRI, acipimox**

CMR scans will follow the same procedure as described above, but participants are instructed to ingest 250 mg acipimox two times per day, 4 hours before and right before the scan, to determine myocardial function. This repeated administration of acipimox is required for adequate suppression of hormone sensitive lipase activity and depletion of plasma FFAs.50 This has been shown to gradually impair cardiac function,42 and is done to disclose any coupling between FFA availability and cardiac function.

**CMR image analysis**

It is performed using Circle42 (Circle Cardiovascular Imaging, Calgary Canada, V.5.5.1). LV volumes, LV mass, LVEF and LVFPFR are determined by tracing of the endocardial and epicardial contours in end-diastolic and endystolic phases. The papillary muscles are excluded from the myocardium. On native and postcontrast T1-mappings, endocardial and epicardial borders are traced, and the mean extravascular volume (ECV) is calculated from areas outside late gadolinium enhancement (LGE) lesions. For determination of the ECV within an LGE lesion, myocardium without LGE in the segment is excluded. Myocardial perfusion scans are inspected for perfusion defects. Regions with infarctions, subendocardial perfusion defects or dark-rim artefacts will be excluded. Blinded to clinical data, the analyses will be reviewed and finalised by two CMR specialists.

**Diurnal blood pressure and Holter monitoring**

Between study days during each visit, diurnal blood pressure is recorded (ScottCare, ABP 320, Cleveland, Ohio, USA) for 24 hours with 15 min intervals between 6:00 and 22:00 and 60 min intervals during night-time. Cardiac rhythm is evaluated with Holter monitoring (SCOTTCARE, CHROMA, model RZ153C, Cleveland, Ohio, USA) for 48 hours.

**Patient and public involvement**

No patient involved.

**ANALYSES**

**Blood and tissue samples**

Subcutaneous fat and muscle biopsies: local analgesia is applied before sampling with a Bergströms cannula. Samples are immediately frozen in liquid nitrogen and stored at −80°C. Blood and urine samples: samples are spun, aliquoted and stored at −20°C (GLP-1, PYY, Glucagon) or −80°C for later analysis. Bedside PG measurements are performed using the glucose oxidase technique (YSI model 2300 STAT Plus; YSI,Yellow Springs, Ohio, USA). Home blood glucose measurements are carried out on Contour XT (Ascensia Diabetes Care Holdings AG, Basel, Switzerland). Safety blood and urine samples are analysed on the same day at the Department of Clinical Biochemistry, Hvidovre Hospital.

**Statistical methods**

**Sample size calculation**

Measures of myocardial function are highly reproducible when assessed using CMR, and interstudy and cohort coefficients of variation are in the range of 3%−5%.51–53 Using the same CMR protocol as the present, Ahtarovski et al found a mean difference of 92 mL/s in LVFPFR between healthy young (585±62 mL/s) and healthy elderly subjects (493±55 mL/s).53 We assume that patients with T2D have LVFPFR corresponding to healthy elderly subjects, and we assume that empagliflozin treatment improves LVFPFR by 30 mL/s (ΔLVFPFR=30 mL/s) from baseline and that insulin treatment does not improve LVFPFR (ΔLVFPFR=0 mL/s).

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**Table 4** Blood samples on metabolic and cardiac MRI (CMR) study days include

<table>
<thead>
<tr>
<th>Metabolic study day</th>
<th>Blood samples: glucose, insulin, C-peptide, glucagon, free fatty acids (FFAs), triglycerides, total amino acids and ketone bodies (beta-hydroxybuturate), tracers/tracees, gut hormones Glycated haemoglobin, urate, blood urea nitrogen, cortisol are sampled at baseline</th>
</tr>
</thead>
<tbody>
<tr>
<td>CMR</td>
<td>Markers of cardiac function, including pro-atrial natriuretic peptide (pro-ANP) and pro-brain natriuretic peptide (pro-BNP), glucose, insulin, C-peptide, glucagon, FFAs, triglycerides, ketone bodies, haematocrit are drawn before and after CMR.</td>
</tr>
</tbody>
</table>
Conservatively, setting the SD of between treatment differences of ΔLVFPR at 30 mL/s, a number of 20 patients would be adequate to determine a 30 mL/s difference between the two treatments with a power of 93% and a two-sided significance level of 0.01, when evaluating data with the paired student’s t-test. In case of a 20% dropout rate, power would still be acceptable (83%, p = 0.01).

Statistical analysis plan

The primary and secondary endpoints are analysed assuming no period effect or treatment–period interaction. This assumption is reasonable, given results from similar studies, where no such interactions or effects have been reported.84 Normally distributed data are presented using standard descriptive statistics, and reported as mean (SD) for normally distributed and median (Q1;Q3) for non-normally distributed data. Likewise, comparisons of normally distributed data are done using the paired Student’s t-test for all completers, whereas Wilcoxon’s paired signed rank test will be used if data are non-normally distributed.

Timeframe

Screenings are performed from January 2018. Last patient, last visit is expected second half of 2021 after which the study will be unregistered with the Danish Medicines Agency and the Capital Region Municipal Ethical Committee within 90 days. Data analyses are expected to be completed by Winter 2022. No later than 12 months after unregistering of the study, will results be made publicly available (www.clinicalregister.eu). Trial registration number: EudraCT: 2017-002101.

ETHICS AND DISSEMINATION

The study is conducted according to ICH GCP guidelines E6 (R2) and the Declaration of Helsinki and all participants will provide oral and written informed consent. The study is approved by the Danish Medicines Agency (ref. id: 2017061587), The Capital Region Ethical Committee (H-17018846) and the Danish Data Protection Agency (AHH-2017-093). Our results, regardless of outcome, will be published in relevant scientific journals. In addition, we will seek to disseminate results through presentations at scientific meetings. Publication will take place as soon as scientifically feasible.

DISCUSSION

The profound and swift benefits of SGLT2i on cardiovascular risk in T2D have inspired the discussion of metabolism and its importance for cardiac function in patients with T2D.37 39 50–57 Especially since, SGLT2 inhibitors have metabolic effects that by and large are opposite to those of insulin treatment. Thus, insulin treatment is associated with increased tissue glucose uptake and utilisation, but suppression of lipid mobilisation and oxidation as well as lowering of plasma concentrations of ketone bodies.58 SGLT2 inhibitors increase lipid mobilisation and oxidation, increase plasma ketone body concentrations and reduce tissue glucose uptake.28 29 Both treatments lower PG, but insulin treatment increases, whereas SGLT2i treatment decreases plasma insulin concentrations. Whether such changes in metabolism affect cardiac function, is still unsettled, but forcing cardiac glucose uptake and utilisation through insulin treatment has been suggested by some to benefit and by others to impair cardiac function,45 59–62 while yet others have suggested increased lipid and ketone body oxidation to be important for proper cardiac function in T2D.57 44 62 Studies on SGLT2i treatments and the effects on cardiac function are beginning to emerge. In a recent study, 42 patients with T2D were randomised to 12 weeks of empagliflozin 10 mg or placebo once daily. SGLT2 inhibition was shown to rapidly improve diastolic cardiac function as evaluated with echocardiography.63 In a placebo-controlled crossover design, after 4 weeks of empagliflozin treatment in patients with T2D, myocardial glucose uptake was reduced and fatty acid oxidation unaltered, but this did not significantly change myocardial oxygen consumption or cardiac efficiency, nor any measure of cardiac function.64 In a Swedish study, 6-week dapagliflozin treatment showed unchanged cardiac fatty acid uptake, a trend towards reduced left atrial maximal volume, and reduced LV oxygen consumption and external work compared with placebo,64 and in the only study found, where an active comparator was used, 10 mg empagliflozin once daily for 12 weeks did not change cardiac lipid accumulation (as measured by MRI spectrometry), cardiac function or cardiac metabolism compared with sitagliptin 50 mg daily.65

In conclusion, existing studies in humans have shown divergent results regarding changes in cardiac diastolic function with little changes in cardiac metabolism. However, most studies have compared cardiac effects of SGLT2i to placebo, thus not accounting for the circumstances that characterised the EMPA-REG trial, where antglycaemic treatment was intensified in the placebo group concurrently.25 Thus, the CVD risk benefits of the study may have arisen from unfavourable metabolic consequences of the treatment in the placebo arm. In the one study with an active comparator empagliflozin was compared with sitagliptin, which not only affects the incretin system but also has less specific metabolic effect.66 Therefore, to date, our study is the one to most directly pursue the coupling between metabolism and cardiac function, by choosing insulin as the comparator, and by including the effects of acute lowering of FFA concentrations in plasma on cardiac function.

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Contributors RT conducted the study, collected all the data, performed data analyses and cosubmit the study. EAR contributed to the study design, designed the keton body and biopsy analysis scheme and performed these analyses. JPG designed the pro-ANP and pro-BNP analysis scheme and performed these analyses. MF designed multiple metabolic analysis schemes (triglycerides, insulin, free fatty acids, uric acid, urea nitrogen and performed these analyses. GHV designed the three-tracer measuring analysis schemes we are using in this study and performed these analyses. UD provided the study with analysis tools (Holter monitors and blood pressure monitors). He performed the analyses of these data. JHJ participated in the protocol design, designed multiple metabolic analyses (GLP-1, glucagon and PYY) and performed the analyses of these data. SM is the principal investigator, coplanned the study and contributed with several analysing tools. NW codesigned the cardiac MRI guideline for the study and contributed with analysing tool (MRI scanner). PLM coplanned the study, codesigned the cardiac MRI guideline for the study and analysed the cardiac MRI data. NBJN coplanned the study, performed all data analyses with RT and cosubmit the study. All of the authors contributed with writing, revising and approving this manuscript.

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