Liquid biopsies and patient-reported outcome measures for integrative monitoring of patients with early-stage breast cancer: a study protocol for the longitudinal observational Prospective Breast Cancer Biobanking (PBCB) study

Håvard Søiland,1,2 Emiel A M Janssen,3,4 Thomas Helland,2,5 Finn Magnus Eliassen,1 Magnus Hagland,2,5 Oddmund Nordgård,6,7 Siri Lunde,1 Tone Hoel Lende,1 Jørn Vegard Sagen,2,5 Kjersti Tjensvoll,8 Bjørnar Gilje,8 Kristin Jonsdottir,9 Einar Gudlaugsson,3 Kirsten Lode,9,10 Kari Britt Hagen,11 Birgitta Haga Gripsrud,10 Ragna Lind,12,13 Anette Heie,11 Turid Aas,11 Marie Austdal,3,9 Nina Gran Egeland,1 Tomm Bernklev,14,15 Timothy L Lash,16 Linn Skartveit,5 Ann Cathrine Kroksveen,17 Satu Oltedal,6 Jan Terje Kvaløy,9,18 Tone Hoel Lende,1 Jørn Vegard Sagen,2,5 Kjersti Tjensvoll,6 Bjørnar Gilje,8 Ernst A Lien,2 Linda Sleire,5 Gunnar Mellgren,2,17 The PBCB-study group


ABSTRACT

Introduction Breast cancer is still the most common malignancy among women worldwide. The Prospective Breast Cancer Biobank (PBCB) collects blood and urine from patients with breast cancer every 6 or 12 months for 11 years from 2011 to 2030 at two university hospitals in Western Norway. The project aims to identify new biomarkers that enable detection of systemic recurrences at the molecular level. As blood represents the biological interface between the primary tumour, the microenvironment and distant metastases, liquid biopsies represent the ideal medium to monitor the patient’s cancer biology for identification of patients at high risk of relapse and for early detection systemic relapse. Including patient-reported outcome measures (PROMs) allows for a vast number of possibilities to compare PROM data with biological information, enabling the study of fatigue and Quality of Life in patients with breast cancer.

Methods and analysis A total of 1455 patients with early-stage breast cancer are enrolled in the PBCB study, which has a one-armed prospective observational design. Participants consent to contribute liquid biopsies (i.e., peripheral blood and urine samples) every 6 or 12 months for 11 years. The liquid biopsies are the basis for detection of circulating tumour cells, circulating tumour DNA (ctDNA), exosomal micro-RNA (miRNA), miRNA in Tumour Educated Platelet and metabolomic profiles. In addition, participants respond to 10 PROM questionnaires collected annually. Moreover, a control group comprising 200 women without cancer aged 25–70 years will provide the same data.

Ethics and dissemination The general research biobank PBCB was approved by the Ministry of Health and Care Services in 2007, by the Regional Ethics Committee (REK) approved by the Ministry of Health and Care Services in 2007, by the Regional Ethics Committee (REK) and the National Richters Committee (Rikshospitalet). The study is registered at ClinicalTrials.gov (NCT04488614). All participants provided informed consent for the use of their samples for research and treatment-related use. The study group aims to identify new biomarkers that enable detection of systemic recurrences at the molecular level. As blood represents the biological interface between the primary tumour, the microenvironment and distant metastases, liquid biopsies represent the ideal medium to monitor the patient’s cancer biology for identification of patients at high risk of relapse and for early detection systemic relapse. Including patient-reported outcome measures (PROMs) allows for a vast number of possibilities to compare PROM data with biological information, enabling the study of fatigue and Quality of Life in patients with breast cancer.

Strengths and limitations of this study

- Prospective Breast Cancer Biobank (PBCB) is a population-based observational study of more than 1000 breast cancer patients with a follow-up of 11 years.
- PBCB collects information from ten patient-related outcome measures yearly, including fatigue and health-related quality of life scores.
- PBCB provides liquid biopsies every 6 months, allowing for analysis of an extensive repertoire of liquid biomarkers.
- Some patients will retract from the project due to study attrition, which may introduce a bias.
- Not all variables have undergone statistical power analysis.

INTRODUCTION

Breast cancer remains the world’s most frequent female malignancy. The last 50 years, the incidence has been more than doubled leading to 2083499 new cases registered worldwide.1 Likewise, among the 5.3 million population in Norway 3726 women were diagnosed with breast...
cancer in 2019. Furthermore, due to both early and late relapses (>5 years after diagnosis), 626,799 global deaths and 650 deaths in Norway occurred in 2019. Moreover, in 2019, there was a prevalence of 51,192 of breast cancer survivors in Norway.

Hence, the classical prognostic and predictive factors such as tumour (T), lymph nodes (N) and metastasis (M) stage, oestrogen receptor (ER) and progesterone receptor (PR) status, histological grade, the proliferation marker Ki67 and human epidermal growth factor receptor 2 (HER-2) amplification seem to be insufficient for optimal treatment decisions to cure systemic disease. This is due to primary tumour heterogeneity or evolutionary changes in residual tumour cells during therapy. Clearly, new biomarkers for detection of systemic relapse are urgently needed for more accurate diagnostics to guide treatment and to detect patients with systemic spread at the earliest possible time point.

As blood represents the biological interface between the primary tumour, the microenvironment and the distant metastases it forms an ideal medium as liquid biopsies to monitor the patient’s cancer biology. Until a clinical overt metastasis is detectable, blood is the only real intermediate medium that can be investigated. It is therefore a paradox that current practice does not include searches for early systemic relapse in blood in early-stage / operable breast cancer patients. In light of this, we have designed this study protocol with the aim to establish methods and biomarkers to detect relapses on the molecular level. Such approaches are circulating tumour cells (CTC), circulating tumour DNA (ctDNA), micro-RNA (miRNA) and metabolomics, which are further described below.

Circulating tumour cell
CTCs are tumour cells that have detached from the primary tumour either by passive tumour cell shedding, or by an active mechanism involving the epithelial-to-mesenchymal transition. CTC count provides independent prognostic information both in operable and metastatic breast cancer.

ctDNA
ctDNA is cell-free tumour-derived DNA present in the plasma. It may originate from apoptotic and necrotic tumour cells in the primary tumour, metastases or circulation, as well as being directly released from viable cells. Substantial evidence supports the idea that ctDNA detection may provide therapeutically relevant, predictive and prognostic information in breast cancer. Recent evidence suggests that residual tumour cell material can be found in peripheral blood samples and function as possible biomarkers. When comparing genetic aberration from primary tumours with metastases from the same patient reports show that many of the acquired driver mutations detected in the metastases cannot be found in the primary tumour.

miRNA, extracellular vesicles and platelets
miRNAs are small, noncoding RNA molecules that control gene expression. They are remarkable stable in plasma and serum. Recently, miRNAs have been found in exosomes, which are small extracellular vesicles (sEV) with a diameter of 40–150 nm. Data also suggest that sEVs may act as messengers of metastasis and assist in the formation of metastasis by ‘priming’ the metastatic site to create a microenvironment that is conducive for cancer cells (The premetastatic niche hypothesis). Analysis of miRNAs in patients with blood of breast cancer can provide an novel way for long-term monitoring of tumour related miRNAs. Recently the use of exosomal miRNAs from urine for the detection of breast cancer has been described as well.

More recently, platelets have emerged as central players in the systemic and local responses to tumour growth. Confrontation of platelets with tumour cells via transfer of tumour-associated biomolecules (‘education’) is an emerging concept and results in the term tumour educated platelets (TEPs). It has become apparent that TEPs absorb protein and RNA molecules from tumours, possibly playing a role in tumour growth and metastasis.

Metabolomics
Metabolomics is the study of small molecules comprising substrates, intermediates and end-products of cellular metabolism, such as amino acids, sugars and small organic acids. The metabolic state of cancer cells is substantially altered compared with normal cells, a fact that can be used for diagnostic purposes. Specific metabolic signatures from tumour tissue provide additional information for determination of breast cancer subtypes and prediction of outcome.

So, there is proof of principle showing the use of preoperative blood and urine samples for metabolic profiling in patients with breast cancer.

Concept of detection of molecular relapse in breast cancer
Based on the described molecular biomarkers, we propose to establish a method for early detection of systemic relapse in patients with early-stage breast cancer. We recommend calling this type of recurrence a ‘molecular relapse’ as it is only detectable by molecular biological methods. The model in figure 1 presents our main hypothesis in the Prospective Breast Cancer Biobank (PBCB). We propose that it is possible to find a molecular relapse in the blood before the recurrence is clinically or radiologically detectable.

Patients-reported outcome measures
Healthcare has shifted towards an increasingly value-based framework for quality-of-care improvement during recent
Decades. As a consequence, more attention has been paid to patient-reported outcomes. The information gained from patient-reported outcome measures (PROMs) comes directly from the patients without intermediate processing. Although PROMs are increasingly collected in breast cancer care, studies reporting on the implementation of systematic PROM administration, including methods and challenges, are less than frequent. PROMs consist of questionnaires that may be single item, unidirectional or multidirectional. They may be collected by several methods, locations and at repeated time points. Systematic PROM collection in breast cancer care is feasible and has a promising result in multiple outcome areas (eg, adherence, satisfaction and clinical decision making) on multiple levels (patients, care providers and care processes). Thus, PROMs may add valuable information in monitoring of psychosocial disease burden in breast cancer survivors.

**METHODS AND ANALYSIS**

**Aims**

The primary aim of the PBCB is to develop a diagnostic tool to detect systemic relapses in patients with early-stage breast cancer on the earliest possible level (molecular level). The secondary aim is to map the long-term psychosocial disease burden in breast cancer survivors.

**Design and setting**

The PBCB project is a one-armed prospective long-term observational study of patients with early-stage breast cancer in Western Norway. The follow-up time is 11 years. The present paper is a protocol paper outlining the plan of the PBCB study and follows the guidelines for preparation outlined by the BMC. We have performed some analyses, and the results are pending for publication.

**Characteristics of patients and normal controls**

The PBCB project is a regional collaboration between Haukeland (HUH) and Stavanger University Hospitals (SUH), in Western Norway. Approximately 550 patients with breast cancer are diagnosed annually at HUH and SUH, approximately 310 and 220, respectively. Women diagnosed with ductal carcinoma in situ (DCIS) or early-stage breast cancer (pT1-pT2/pN0pN1/M0) at HUH and SUH were consecutively invited to participate in the PBCB-project. The clinic-pathological features of our study population are shown in Table 1. Norwegian citizens have unique personal social security numbers allowing coupling of information to various nationwide registries including the Norwegian Cancer Registry, the Cause of Death Registry, Norwegian Prescription Database, the Norwegian Labour and Welfare Administration Database. Therefore, a complete follow-up of the patients is facilitated, creating a robust population-based prospective biobank of women with early-stage breast cancer.

The enrolment of the control cohort of women without breast cancer commenced at 1 September 2011 at SUH and is estimated to last until 1 September 2032. From these women, blood will be sampled once, and they will fill in the questionnaires once (see Section 3.2).
Table 1  Clinical and pathological features of the PBCB population

<table>
<thead>
<tr>
<th>Clinical pathological Feature</th>
<th>No HUH</th>
<th>HUH (%)</th>
<th>No SUH</th>
<th>SUH (%)</th>
<th>No overall</th>
<th>Overall (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Enrolled</td>
<td>1104</td>
<td>75.9</td>
<td>351</td>
<td>24.1</td>
<td>1455</td>
<td>100</td>
</tr>
<tr>
<td>Age (mean)</td>
<td>57.9</td>
<td>58.6</td>
<td>59.0</td>
<td>58.3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (median)</td>
<td>58.0</td>
<td>59.0</td>
<td>58.5</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age: range</td>
<td>23–86</td>
<td>24–90</td>
<td>23–90</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;35 years</td>
<td>11</td>
<td>1.0</td>
<td>3</td>
<td>0.9</td>
<td>14</td>
<td>0.96</td>
</tr>
<tr>
<td>36–55 years</td>
<td>417</td>
<td>37.8</td>
<td>123</td>
<td>35.0</td>
<td>540</td>
<td>37.1</td>
</tr>
<tr>
<td>55–69 years</td>
<td>537</td>
<td>48.6</td>
<td>181</td>
<td>51.6</td>
<td>717</td>
<td>49.3</td>
</tr>
<tr>
<td>≥70 years</td>
<td>139</td>
<td>12.6</td>
<td>44</td>
<td>12.5</td>
<td>183</td>
<td>12.6</td>
</tr>
<tr>
<td>Missing</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>DCIS</td>
<td>20</td>
<td>1.9</td>
<td>30</td>
<td>8.5</td>
<td>50</td>
<td>3.4</td>
</tr>
<tr>
<td>pT1mic</td>
<td>2</td>
<td>0.2</td>
<td>3</td>
<td>0.9</td>
<td>5</td>
<td>0.3</td>
</tr>
<tr>
<td>pT1a</td>
<td>43</td>
<td>3.9</td>
<td>8</td>
<td>2.3</td>
<td>51</td>
<td>3.5</td>
</tr>
<tr>
<td>pT1b</td>
<td>147</td>
<td>13.3</td>
<td>52</td>
<td>14.8</td>
<td>199</td>
<td>13.6</td>
</tr>
<tr>
<td>pT1c</td>
<td>430</td>
<td>38.9</td>
<td>141</td>
<td>40.2</td>
<td>571</td>
<td>39.2</td>
</tr>
<tr>
<td>pT2</td>
<td>304</td>
<td>27.5</td>
<td>110</td>
<td>31.3</td>
<td>414</td>
<td>28.5</td>
</tr>
<tr>
<td>pT3</td>
<td>71</td>
<td>6.4</td>
<td>2</td>
<td>0.6</td>
<td>79</td>
<td>5.4</td>
</tr>
<tr>
<td>PT4a</td>
<td>10</td>
<td>0.9</td>
<td>0</td>
<td>0</td>
<td>10</td>
<td>0.7</td>
</tr>
<tr>
<td>PT4b</td>
<td>2</td>
<td>0.18</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>0.1</td>
</tr>
<tr>
<td>Missing*</td>
<td>75</td>
<td>6.8</td>
<td>5</td>
<td>1.4</td>
<td>80</td>
<td>5.5</td>
</tr>
<tr>
<td>pN0</td>
<td>766</td>
<td>69.4</td>
<td>238</td>
<td>67.8</td>
<td>1004</td>
<td>69.0</td>
</tr>
<tr>
<td>pN1mic</td>
<td>2</td>
<td>0.2</td>
<td>6</td>
<td>1.7</td>
<td>8</td>
<td>0.5</td>
</tr>
<tr>
<td>pN1</td>
<td>213</td>
<td>19.2</td>
<td>60</td>
<td>17.1</td>
<td>273</td>
<td>18.8</td>
</tr>
<tr>
<td>pN2</td>
<td>29</td>
<td>2.6</td>
<td>17</td>
<td>4.8</td>
<td>46</td>
<td>3.2</td>
</tr>
<tr>
<td>pN3</td>
<td>13</td>
<td>1.2</td>
<td>4</td>
<td>1.1</td>
<td>17</td>
<td>1.2</td>
</tr>
<tr>
<td>pN positive</td>
<td>257</td>
<td>23.2</td>
<td>87</td>
<td>24.8</td>
<td>344</td>
<td>25.9</td>
</tr>
<tr>
<td>Missing*</td>
<td>81</td>
<td>7.3</td>
<td>26</td>
<td>7.4</td>
<td>107</td>
<td>7.4</td>
</tr>
<tr>
<td>ER pos</td>
<td>916</td>
<td>83.0</td>
<td>271</td>
<td>77.2</td>
<td>1187</td>
<td>81.6</td>
</tr>
<tr>
<td>ER neg</td>
<td>124</td>
<td>11.2</td>
<td>14</td>
<td>4.0</td>
<td>138</td>
<td>9.5</td>
</tr>
<tr>
<td>Missing*</td>
<td>64</td>
<td>5.8</td>
<td>66</td>
<td>18.8</td>
<td>130</td>
<td>8.9</td>
</tr>
<tr>
<td>PR pos</td>
<td>794</td>
<td>71.9</td>
<td>229</td>
<td>65.2</td>
<td>1020</td>
<td>70.1</td>
</tr>
<tr>
<td>PR neg</td>
<td>229</td>
<td>20.7</td>
<td>84</td>
<td>23.9</td>
<td>313</td>
<td>21.5</td>
</tr>
<tr>
<td>Missing</td>
<td>81</td>
<td>7.3</td>
<td>38</td>
<td>10.8</td>
<td>119</td>
<td>12.2</td>
</tr>
<tr>
<td>HER2 neg</td>
<td>917</td>
<td>83.1</td>
<td>285</td>
<td>81.2</td>
<td>1202</td>
<td>82.6</td>
</tr>
<tr>
<td>HER2 pos</td>
<td>121</td>
<td>11.0</td>
<td>28</td>
<td>8.0</td>
<td>149</td>
<td>10.2</td>
</tr>
<tr>
<td>Missing*</td>
<td>66</td>
<td>6.0</td>
<td>38</td>
<td>10.8</td>
<td>104</td>
<td>7.2</td>
</tr>
<tr>
<td>Ki67 &lt;15%</td>
<td>171</td>
<td>15.5</td>
<td>72</td>
<td>20.5</td>
<td>243</td>
<td>16.7</td>
</tr>
<tr>
<td>Ki67 15%–29%</td>
<td>280</td>
<td>25.4</td>
<td>102</td>
<td>29.1</td>
<td>382</td>
<td>26.3</td>
</tr>
<tr>
<td>Ki67 ≥30%</td>
<td>263</td>
<td>23.8</td>
<td>156</td>
<td>44.4</td>
<td>419</td>
<td>28.8</td>
</tr>
<tr>
<td>Missing*</td>
<td>390†</td>
<td>35.3</td>
<td>21</td>
<td>6.0</td>
<td>411</td>
<td>28.2</td>
</tr>
<tr>
<td>MAI&lt;10‡</td>
<td>–</td>
<td>–</td>
<td>210</td>
<td>59.8</td>
<td>210</td>
<td>59.8</td>
</tr>
<tr>
<td>MAI≥10</td>
<td>–</td>
<td>–</td>
<td>91</td>
<td>25.9</td>
<td>91</td>
<td>25.9</td>
</tr>
<tr>
<td>Missing*</td>
<td>–</td>
<td>–</td>
<td>50</td>
<td>14.2</td>
<td>50</td>
<td>14.2</td>
</tr>
<tr>
<td>Grade 1</td>
<td>251</td>
<td>22.3</td>
<td>69</td>
<td>19.7</td>
<td>320</td>
<td>22.3</td>
</tr>
<tr>
<td>Grade 2</td>
<td>420</td>
<td>38.0</td>
<td>117</td>
<td>33.3</td>
<td>537</td>
<td>37.3</td>
</tr>
</tbody>
</table>

Continued
**Table 1** Continued

<table>
<thead>
<tr>
<th>Clinical pathological Feature</th>
<th>No HUH (%)</th>
<th>HUH (%)</th>
<th>No SUH (%)</th>
<th>SUH (%)</th>
<th>Overall (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade 3</td>
<td>254</td>
<td>23.0</td>
<td>124</td>
<td>35.3</td>
<td>378</td>
</tr>
<tr>
<td>Missing</td>
<td>179</td>
<td>16.2</td>
<td>41</td>
<td>6.8</td>
<td>203</td>
</tr>
<tr>
<td>BCT</td>
<td>430</td>
<td>38.9</td>
<td>237</td>
<td>67.5</td>
<td>667</td>
</tr>
<tr>
<td>Mastectomy</td>
<td>648</td>
<td>58.7</td>
<td>112</td>
<td>31.9</td>
<td>760</td>
</tr>
<tr>
<td>Missing</td>
<td>26</td>
<td>2.4</td>
<td>9</td>
<td>2.6</td>
<td>35</td>
</tr>
</tbody>
</table>

*The high number of missing data is due to inclusion of DCIS cases.
†Ki-67 was not implemented in the pathology reports until 2013.
‡MAI is not used at HUH.

BCT, breast conservative treatment; DCIS, Ductal Carcinoma in situ; ER, oestrogen receptor; Grade, Histological Grade; HER2, human epidermal growth factor receptor-2; HUH, Haukeland University Hospital; MAI, Mitotic Activity Index; PBCB, Prospective Breast Cancer Biobank; pN, pathological node status; PR, progesterone receptor; pT, pathological tumour status; SUH, Stavanger University Hospital.

...distribution supports a population-based patient accrual in the PBCB study.

**Characteristics of the control group**

In the PBCB project, we will accrue up to 200 healthy women (now 87) without breast cancer from the Norwegian mammography screening programme who will serve as a control group. The enrolled women will be between 25 and 70 years of age. In this cross-sectional study, these women will also provide the same package of liquid biopsies as the patients with breast cancer participating in PBCB. Moreover, they will fill out an adapted questionnaire package with the same PROM-questions as the patients with breast cancer provide. Thus, these healthy women will serve as important reference points.

**Patients and public involvement**

When the study started, the use of patient in planning studies was not very well established in Norway. Later, however, the patients were involved in study planning including patient priorities. Furthermore, the PBCB group at SUH maintains an active collaboration with the regional breast cancer society, which represents all patients with breast cancer in our region. Two society representatives are included in regular PBCB meeting as active partners of the user involvement programme and have provided valuable input into several aspects of the PBCB project including recruitment and conduct of the study.

Dissemination of the results from this project towards the scientific and medical communities will include publication of scientific papers in international peer-reviewed scientific journals. Project results will also be presented at international conferences, as well as communication to the public through national and regional newspapers and websites like www.forskning.no and our home page (www.sus.no). Results will also be presented at scenes of patient enquiry, such as the Norwegian Cancer Society and the National Network of Breast Cancer Research, to strengthen the breast cancer research, inspire user...
### Figure 2

Overview of the study design (one armed observational study) and sampling protocol in the PBCB project. Number of patients at each study site lost to follow-up at each time point. ✔=Completed; ◼=Ongoing; FU, follow-up; HUH, Haukeland University Hospital; PBCB, Prospective Breast Cancer Biobank; PROM, patient-reported outcome measure; SUH, Stavanger University Hospital; TBC, to be conducted.
involvement and provide research dissemination at an approachable level for the public.

**Study period and follow-up**

At HUH, patients with early-stage breast cancer were included from 25 January 2011 to 31 January 2017. Thus, the proportion of included patients in this period was 1104/1860 = 59%. At SUH, 351 patients were included into PBCB from 7 November 2013 to 25 August 2016 giving a proportion of 53% inclusion of total eligible patients. Average for both study sites is 54% inclusion. Follow-up at both sites will last until 31 January 2030 regarding questionnaires and blood samples. Clinical follow-up (ie, relapses) will last until 2050. Recruitment of healthy women included in the control group will be from 2 September 2020 to 2 September 2022 (n=200).

**PROTOCOLS OF ANALYSES**

**Fresh frozen and paraffin-embedded tissue samples**

At SUH, fresh frozen tumour tissue is obtained from the surgical specimens from tumours larger than 10 mm in diameter. Otherwise, paraffin embedded tumour tissue is available from the diagnostic biobanks at both HUH and SUH for all patients. DNA from primary tumours and available metastatic lesions will be isolated using the Allprep FFPE kit (Qiagen). The resulting DNA profile will be analysed by next-generation sequencing using the Oncomine Comprehensive Assay V.3 (ThermoFisher). This is an amplicon-based approach, which detects commonly occurring single nucleotide variants, insertions and deletions, and copy number variations in the tumour genome, especially in ER, p53 and other known oncopgenes. These analyses will provide us with a biological understanding and background knowledge for each of the individual tumours.

**Liquid biopsies**

The sampling protocol of liquid biopsies is detailed in table 2. In brief, 60 mL blood and 20 mL urine samples are collected from each patient. The blood samples are divided and preserved into 31 aliquots while the urine samples are divided into 6 aliquots. All sample aliquots are stored at −80°C. Importantly, plasma is sampled at +4°C for assessment of cytokines and Cell Partitian Tube (CPT) tubes makes DNA isolation from blood lymphocytes possible. Isolation of CTCs and ctDNA is performed from blood and plasma samples collected and processed at room temperature.

**Circulating tumour cells**

CTCs will be enriched from peripheral blood samples by density centrifugation and subsequent immunomagnetic depletion of leucocytes using a new EpCAM-independent

<table>
<thead>
<tr>
<th>Sample</th>
<th>Supplier</th>
<th>Volume</th>
<th>No of tubes</th>
<th>Centrifuge</th>
<th>Aliquots for storage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum</td>
<td>BD Vacutainer CAT</td>
<td>10 mL</td>
<td>2</td>
<td>1500g×25 min at RT</td>
<td>6×0.5 mL and 4×1.5 mL, stored at −80°C</td>
</tr>
<tr>
<td>EDTA-Whole Blood</td>
<td>Vacutette greiner bio-one</td>
<td>3 mL</td>
<td>1</td>
<td>−</td>
<td>2×1.5 mL, stored at −80°C</td>
</tr>
<tr>
<td>EDTA-plasma (cold) (kept on ice)</td>
<td>Vacutette greiner bio-one</td>
<td>9 mL</td>
<td>1</td>
<td>2500g×10 min at 4°C</td>
<td>6×0.5 mL, 2×1.0 mL, 1×0.5 mL and buffy coat, stored at −80°C</td>
</tr>
<tr>
<td>EDTA-plasma (RT)</td>
<td>Vacutette greiner bio-one</td>
<td>9 mL</td>
<td>2</td>
<td>2200g×10 min at room temperature</td>
<td>6×0.5 mL and 2×1.5 mL, stored at −80°C</td>
</tr>
<tr>
<td>EDTA-platelets (kept on ice)</td>
<td>Vacutette greiner bio-one</td>
<td>9 mL</td>
<td>1</td>
<td>1. ii) 200g×20 min /10 min at 4°C with no brake applied. 2. 1000g×10 min at 4°C</td>
<td>1×0.5 mL, stored at −80°C</td>
</tr>
<tr>
<td>EDTA-CTC</td>
<td>Vacutette greiner bio-one</td>
<td>9 mL</td>
<td>1</td>
<td>CTCs enrichment by MINDEC negative depletion according to Lapin et al 2016.27 one tube with lysed CTCs in 350 µL RLT buffer, stored at −80°C</td>
<td></td>
</tr>
<tr>
<td>Vacutainer CPT</td>
<td>BD Vacutainer CPT Mononuclear Cell Preparation Tube</td>
<td>8 mL</td>
<td>1</td>
<td>i. 1500g×25 min at room temperature ii. 1000g×10 min at 4°C</td>
<td>1×0.5 mL, stored at −80°C</td>
</tr>
<tr>
<td>PAX-gene blood RNA tube</td>
<td>PAXgene PreAnalytiX</td>
<td>2.5 mL</td>
<td>1</td>
<td>−</td>
<td>Room temperature at 2–72 hours; −20°C for 24 hours; stored at −80°C</td>
</tr>
<tr>
<td>Urine</td>
<td></td>
<td>20 mL</td>
<td>1</td>
<td>−</td>
<td>6×1.5 mL, stored at −80°C</td>
</tr>
</tbody>
</table>

CAT, Clot Activator Tube; CPT, Cell Partition Tube; PBCB, Prospective Breast Cancer Biobank; RLT, RNeasy Lysis cells and Tissue; RT, room temperature.
**Tumour educated platelets**

miRNA in blood will be isolated from blood platelets, which have been in contact with tumour cells. This contact has resulted in uptake of miRNAs from the tumour into the TEPs. Several studies have also shown that miRNAs from TEPs can be transferred and mediate biological effects in recipient cells. An efficient protocol for miRNA isolation from platelets by using the the miREASY kit (Qiagen) has been established. Preliminary data show that we are able to detect 10 tested miRNAs.

**Circulating tumour DNA**

In the PBCB project, we isolate cell-free DNA (cfDNA), will be isolated from 1 to 2 mL room temperate plasma samples by QIAamp circulating nucleic acid kit (Qiagen). The cfDNA samples are stored at stored at −80°C for subsequent sequencing of ctDNA.

**Determination of miRNA in sEVs**

Total RNA will be separated from sEVs in plasma samples using the exoNeasy Serum/Plasma kit (Qiagen), which isolates sEVs and sEV-contained RNA in a two-step process. In a first phase, miRNA profiling will be performed on the isolated RNA with the Ion total RNA-seq kit V2 (Thermo Fisher) on our Ion Proton deep sequencing instrument. Preliminary results show that miRNAs are present in sEVs at measurable levels.

Although there are few studies that have analysed circulating exosomal miRNAs and TEPs in blood from patients with breast cancer, a recent study reported on 11 exosomal miRNAs that were differentially expressed in patients with breast cancer with and without recurrence.

**Thermolabile biomarkers**

Certain biomarkers decay rapidly at room temperature and must be preserved by immediately cooling of the drawn blood sample on water ice, cooled centrifugation at 4°C, followed by aliquotation and storage in −80°C freezer without any delay.

**Metabolomics**

Serum will be used for the analysis of small molecules, hormones (eg, estradiol), drugs (eg, tamoxifen, aromatase inhibitors) and vitamins (eg, vitamin D) by using ELISA and liquid chromatography-tandem mass spectrometry methodology. Moreover, metabolites will be measured by MR spectroscopy at the MR Core Facility, Norwegian University of Science and Technology. Approximately 30 metabolites and 105 lipoprotein subfractions can be quantified in the serum. The MR Core Facility collaborates with the vendor (Bruker BioSpin GmbH, Germany) to ensure efficient and validated acquisition protocols, making the method feasible for high throughput analysis and implementation into the clinic. Modelling biological pathways will be conducted by integrating multivariate statistics with biological databases (Human Metabolome Database, KEGG), using available online tools (Metaboanalyst).

**Urine samples**

Urine samples are aliquoted into 1.5mL tubes and preserved in −80°C freezer for later analyses of various analytes excreted into the urine.

**Biobanking**

All samples are registered in a regional tracking system for biobank samples, Lab vantage. Several different parameters related to sampling are logged, for example, time of sampling, centrifugation and freezing, discrepancies, number of aliquots, etc. These data are available at all times. All biomarkers will be analysed in batches to avoid systematic errors and an unpredictable day-to-day variation. Long term storage of all analytes exerts a challenge of stability at −80°C. For example, some miRNAs show a decay of 12% after 5 years of storage, while others have shown only minor changes in micro-RNA at −80°C storage levels. Moreover, metabolomics analytes like amino acids, sugars carnitine show a decay of 15% and increment of 14% during long term storage at −80°C during 5-year storage. In this regard, thermolabile biomarkers like cytokines will be analysed first to avoid too much decay of the analytes. In contrast, ctDNA is more stable to long-term storage.

**Preanalytical conditions and centralisation of analysis**

To minimise differences in preanalytical condition between the two study sites the PBCB steering committee meet several times a year to monitor the collection protocol of the various liquid biopsies. Due to the heterogeneity in the PBCB-material between SUH and HUH, all analysis of the various biomarkers will be centralised to Stavanger. This will enhance control of the data quality.
The Mishel Uncertainty in Illness Scale; (8) Diet questionnaire and (9) ROMA III questionnaire for irritable bowel complaints. Some of the PROMs require trained staff to guide the patient in their completion. At HUH, PROM data are obtained annually from a cohort of 160 patients. At SUH, 351 patients have completed the PROM data at baseline. Figure 2 shows how many patients that are lost during PROM data follow-up. PROM questionnaires are further described in table 3.

Scores of the questionnaires
We applied mean sum scores from SHC-Inventory and its five subscales, and the satisfaction with the quality of patient information given by the healthcare professionals. Sum scores of FACT-Endocrine Subscale and FACT-F were used following the FACIT Organisation’s scoring guidelines. We performed the reversals as indicated in the guidelines and summed individual items to obtain a score. The subscale scores were produced by multiplying the sum of each item score by the number of items in the subscale, and then divided by the numbers of items answered. In the FACTIT scales, a high score means better social well-being, or fewer symptoms of fatigue or endocrine symptoms. As an example of the utilisation of the PROM-data we have shown that fatigue and depression overrule side effects of oncological treatment in predicting self-reported health complaints in breast cancer survivors.51

STATISTICAL AND POWER ANALYSIS
We will use univariate (Kaplan Meier) and multivariable linear regression, and logistic regression analysis.

THE PBCB STUDY GROUP ACRONYM
Research members of the PBCB study group are represented collectively under the acronym ‘The PBCB Study Group’, which can be used as a co-author in research papers where material from the PBCB biobank or PROM data are used. These researchers are:

Prof. Håvard Søiland MD, PhD, Prof. Gunnar Mellgren MD, PhD, Tone Hoel Lende MD, PhD, Anette Heie MD, PhD, Jørn V. Sagen MD, PhD, Prof. Emiel AM Janssen PhD, Kristin Jonsdottir PhD, Ann Cathrine Kroksveen MSc, Linda Sleire MSc, Thomas Helland PhD, Einar Guillaume, MD, PhD, Prof. Oddmund Nordgård PhD, Kjersti Tjensvoll PhD, Satu Oltedal PhD, Bjornar Gilje MD, PhD, Prof. Jan Terje Kvaløy PhD, Assoc. Prof. Kirsten Lode RN PhD, Prof. Birgitta Haga Gripsrud, PhD, Kari Britt Hagen RN, PhD, Marie Austdal, PhD, Siri Lunde MSc, Nina Gran Egeland PhD, Prof. & Chair Timothy L Lash, DSc, MPH, Finn Magnus Eliassen MD and Prof. emeritus Ernst A Lien MD, PhD

DISCUSSION
The overall aim of the PBCB is to better capture patients with early-stage breast cancer at high risk of systemic relapse and monitor the patients in order to identify the relapse at the molecular level. With our planned follow-up time of 11 years, we expect to identify patients experiencing both early and late relapse. Previous studies have followed patients with breast cancer with PROM data for 2–3 years at most.53 It is reasonable to anticipate that detection of molecular relapses in liquid biopsies will give a novel opportunity to initiate secondary adjuvant treatment at an earlier time point than with the clinical approach of today. Others have shown that ctDNA may be detected up to 10 months prior to the clinical relapse in patients with early breast cancer irrespective of molecular subtypes.54 We will investigate CTC, ctDNA, TEPs, miRNA and metabolomics in both serum/plasma and urine. Thus, we combine five biomarker modalities to detect molecular relapses, which has not been done before. We anticipate that we will have a good chance of detecting molecular relapses in the earliest possible way. Considering that most relapses appear after 2–3 years, the best approach may be to sample blood every 4 months for 10 years. This would enable detection of molecular relapses treatable by secondary adjuvant treatment, including immune therapy.6 56 However, there are still several issues such as standardisation of protocols, which needs to be resolved before translation into clinical practice.57 We aim to increase our knowledge regarding liquid biopsies in patients with early breast cancer.

Furthermore, PBCB is ideal for therapeutic drug monitoring. Recently, we have shown a reduced breast cancer specific and overall survival in patients with low levels of active tamoxifen metabolites.58 59 The PBCB protocol will allow for measurement of drug metabolites every 6 months, allowing for monitoring of adherence and drug discontinuation.

The focus on disease burden in breast cancer survivors during an 11-year follow-up is to our knowledge the longest prospective observational study of patients with breast cancer using PROM data. The long follow-up creates a unique opportunity to join circulating biomarker data and PROM-data to the extensive Norwegian public patient’s registries.37 38 60 Although this long follow-up may produce bias by attrition,60 we do not anticipate that attrition will become a problem in our study due to close follow-up by our dedicated study nurses.

In summary, the PBCB-project represents an interdisciplinary platform for future research projects on...
<table>
<thead>
<tr>
<th>Questionnaire (abbrev)</th>
<th>Full name</th>
<th>Items</th>
<th>Scoring/scale</th>
<th>Remarks</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>EORTC-QLQ-C30</td>
<td>European Organisation for Research and Treatment of Cancer- Quality of Life Questionnaire-Cancer 30</td>
<td>30</td>
<td>All items address health issues during the last week</td>
<td>Four point rating scale&lt;br&gt;0=not at all&lt;br&gt;1=a little&lt;br&gt;2=some&lt;br&gt;3=very much&lt;br&gt;Total score range: 0–90</td>
<td>Cronbach’s alpha-coefficient for multi-item scales ranged from 0.56 to 0.85, with emotional functioning having the highest Cronbach’s alpha-coefficient. General health/QoL subscale was correlated significantly with all other subscales.</td>
</tr>
<tr>
<td>EORTC-QLQ BR23</td>
<td>European Organisation for Research and Treatment of Cancer-Quality of Life Questionnaire-Breast 23</td>
<td>23</td>
<td>All items address health issues during the last week</td>
<td>Four point rating scale&lt;br&gt;0=not at all&lt;br&gt;1=a little&lt;br&gt;2=some&lt;br&gt;3=very much&lt;br&gt;Total score range: 0–69</td>
<td>Cronbach’s alpha ranged from 0.78 to 0.83 for the EORTC-BR23 questionnaire.</td>
</tr>
<tr>
<td>FACT-B</td>
<td>Functional assessment of Cancer Therapy - Breast</td>
<td>A 37-item instrument designed to measure five domains of HRQOL in patients with breast cancer: Physical, social, emotional, functional well-being as well as a breast-cancer subscale</td>
<td>Five point rating scale&lt;br&gt;0=not at all&lt;br&gt;1=a little&lt;br&gt;2=some&lt;br&gt;3=a lot&lt;br&gt;4=very much&lt;br&gt;Range: 0–148&lt;br&gt;According to the guidelines, all the questionnaires are drafted in a way that higher scores represent improved quality of life</td>
<td>Cronbach’s alpha for the various constructs is behind 0.66 and 0.85</td>
<td>65</td>
</tr>
<tr>
<td>FACT-ES</td>
<td>Functional Assessment of Cancer Therapy-Endocrine Subscale</td>
<td>Contains 19 statements examining self-reported menopausal and sexual symptoms related to breast cancer endocrine therapy during the last week</td>
<td>Five point rating scale&lt;br&gt;0=not at all&lt;br&gt;1=a little&lt;br&gt;2=some&lt;br&gt;3=a lot&lt;br&gt;4=very much&lt;br&gt;High scores=low symptom complaints</td>
<td>Cronbach’s alpha for the Norwegian version of the 19 items endocrine subscale was 0.86 in patients and 0.83 in healthy controls</td>
<td>66–70</td>
</tr>
<tr>
<td>FACT-F version 4</td>
<td>Functional Assessment of Chronic Illness Therapy- Fatigue</td>
<td>Consists of 13 items measuring physical and mental fatigue affecting daily life during the last week</td>
<td>Five point rating scale&lt;br&gt;0=not at all&lt;br&gt;1=a little&lt;br&gt;2=some&lt;br&gt;3=a lot&lt;br&gt;4=very much&lt;br&gt;High scores=low symptom complaints</td>
<td>Cronbach’s alpha for the questionnaire was 0.84 in patients and 0.81 in healthy controls</td>
<td>51, 66, 71</td>
</tr>
<tr>
<td>SHC</td>
<td>Subjective Health Complaints</td>
<td>The instrument measures 29 subjective, somatic and psychological complaints experienced during the last month&lt;br&gt;1. ‘Musculoskeletal pain’ (back pain, low back pain, arm pain, shoulder pain, migraine and leg pain, range 0–24),&lt;br&gt;2. ‘Pseudoneurology’ (tiredness, sleep problems, anxiety, sadness/depression, extra heartbeats, heat flushes and dizziness, range 0–21),&lt;br&gt;3. ‘Gastrointestinal problems’ (gas discomfort, stomach discomfort, diarrhoea, constipation, gastritis/ulcer, heartburn and stomach pain, range 0–21),&lt;br&gt;4. ‘Allergy’ (allergies, breathing difficulties, eczema, and asthma, range 0–15) and&lt;br&gt;5. ‘Influenza’ (colds/influenza and coughing, range 0–6)</td>
<td>Four point rating scale&lt;br&gt;0=no complaints&lt;br&gt;1=a little complaint&lt;br&gt;2=some complaints&lt;br&gt;3=severe complaints&lt;br&gt;Total score range: 0–87</td>
<td>The instrument has a satisfactory validity and reliability in both the general Norwegian population, and Norwegian patients with irritable bowel syndrome&lt;br&gt;The Cronbach’s alpha for Subjective Health Complaints Inventory in patients with breast cancer and healthy controls were 0.89 and 0.87 respectively</td>
<td>62–66</td>
</tr>
</tbody>
</table>

Continued
### Questionnaire (abbrev) Full name Items Scoring/scale Remarks References

<table>
<thead>
<tr>
<th>Questionnaire (abbrev)</th>
<th>Full name</th>
<th>Items</th>
<th>Scoring/scale</th>
<th>Remarks</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>HADS</td>
<td>Hospital Anxiety and Depression Scale</td>
<td>Two subscales: Anxiety (A) and Depression (D)</td>
<td>four point rating scale</td>
<td>The Cronbach alpha for HADS-A ranged from 0.68 to 0.93 (below adequate to excellent) with a mean of 0.83 (very good). Cronbach alpha for HADS-D ranged from 0.67 to 0.90 (below adequate to excellent) with a mean of 0.82 (very good).</td>
<td>77-79</td>
</tr>
<tr>
<td>FSS</td>
<td>Fatigue Severity Scale</td>
<td>9 items</td>
<td>The total score is divided by 9</td>
<td>Total score range from 1.00 to 7.00.</td>
<td>72-75</td>
</tr>
<tr>
<td>fVAS</td>
<td>Fatigue – Visual Analogue Scale</td>
<td>Single item</td>
<td>The scale is a 100 mm line where the far left is 0 mm = 'no problems with fatigue and exhaustion', and the far right is 100 mm = 'so much fatigue and exhaustion that is possible to experience', during the last week</td>
<td>Intra Class Correlation values for the fVAS is between 0.846 and 0.888</td>
<td>77 76 77</td>
</tr>
<tr>
<td>Muscle – Joint Questionnaire</td>
<td>Muscle and joint pains</td>
<td>three items</td>
<td>100 mm VAS scale</td>
<td></td>
<td></td>
</tr>
<tr>
<td>SF-MUIS</td>
<td>Short-form Mishel Uncertainty in Illness Scale</td>
<td>The SF-MUIS covers five statements from the modified 33 questions MUIS, examining the uncertainty in illness in hospitalised adults asking the participants to reflect and report how they agreed to the following five statements; 'I have a lot of questions without answers', 'I understand everything explained to me', 'The doctors say things to me that can have many meanings', 'There are so many different types of staff; it's unclear who is responsible for what', and 'The purpose of each treatment is clear to me'.</td>
<td>five point rating scale: 1 'strongly disagree', 2 'disagree', 3 'uncertain', 4 'agree', 5 'strongly agree'</td>
<td>The MUIS-BT measures four constructs: ambiguity/inconsistency, unpredictability of disease prognosis, unpredictability of symptoms and other triggers, and complexity. The ordinal coefficient alpha for the SF-MUIS was 0.70 in Norwegian patients with breast cancer.</td>
<td>78 79</td>
</tr>
<tr>
<td>Side effects endocrine therapy</td>
<td>Aromatase Inhibitors (AI)</td>
<td>9 organ systems</td>
<td>yes/no/don't know</td>
<td>This questionnaire is based on knowledge on side effects of AIs. Not validated.</td>
<td></td>
</tr>
<tr>
<td>Side effects endocrine therapy</td>
<td>Tamoxifen</td>
<td>6 organ systems</td>
<td>yes/no/don't know</td>
<td>This questionnaire is based on knowledge on side effects of tamoxifen. Not validated.</td>
<td></td>
</tr>
<tr>
<td>Adherence to endocrine treatment</td>
<td>Self-reported adherence</td>
<td>One question: the patients were asked if they take Tamoxifen or Aromatase inhibitor every day?</td>
<td>yes or no</td>
<td></td>
<td>–</td>
</tr>
<tr>
<td>Diet questionnaire</td>
<td>Nordic Nutrition Recommendations 2012</td>
<td>The patients were asked whether they had made any changes in their diet 2 years after they were diagnosed with breast cancer. If the answer was 'yes', they were asked about the major changes they had made in relation to food items excluded or included in their diet. Avoidance of 36 different food items over six clusters: Cereals, fish/shellfish, fruit and vegetables, dairy products, processed food and others. Finally, the participants were asked to report their daily and weekly use of milk and other dairy products, and whether they consumed alcohol or not</td>
<td>The response options: 'yes', 'no' or 'partially'. Further, the respondents who replied positively to consuming alcohol, reported their consumption (daily, every second day, once or twice per week, once per month, or more rarely), and the number of glasses during 1 week.</td>
<td>Cronbach's alphas=0.83 (0.71–0.97)</td>
<td>80 81</td>
</tr>
</tbody>
</table>

HRQoL, health-related quality of life; PBCB, Prospective Breast Cancer Biobank; SF-MUIS, Short Form of the Mishel Uncertainty in Illness Scale.
early-stage breast cancer. Providing a method for molecular detection of systemic relapse will be one small step for the development of biomarkers but one giant leap for the treatment of patients with early-stage breast cancer.

ETHICS
The general research biobank PBCB was approved by the Ministry of Health and Care Services in 2007, by the Regional Ethics Committee (REK) of Northern Norway in 2010 (#2010/1957). The PROM (#2011/2161) and the biomarker study PerMoBreCan (#2015/2010) were approved by REK in 2011 and 2015, respectively. All patients have been informed by study nurse or a research coordinator both certified in Good Clinical Practice, prior to consenting to take part in the study. This took place in the time window between diagnosis and the operation. The patients have given written consent to the general research biobank PBCB and HR-QoL studies (ie, two written consents) from 2010 onward. Results from the PBCB-studies will be published in international conventions, in international peer-reviewed scientific journals, in mass media and in social media. At the end of the study, deidentified data will be curated in-house according to European guidelines, and will be available for researchers on request. Requests should describe the nature of the proposed research and scope of the requirements, and will be reviewed by the PBCB board.

Author affiliations
1Department of Breast and Endocrine Surgery, Stavanger University Hospital, Stavanger, Norway
2Department of Clinical Science, University of Bergen, Bergen, Norway
3Department of Pathology, Stavanger University Hospital, Stavanger, Norway
4Department of Chemistry, Bioscience and Environmental Engineering, University of Stavanger, Stavanger, Norway
5Department of Medical Biochemistry and Pharmacology, Haukeland University Hospital, Bergen, Norway
6Department of Hematology and Oncology, Stavanger University Hospital, Stavanger, Norway
7Department of Chemistry, Bioscience, University of Stavanger, Stavanger, Norway
8Department of Oncology, Stavanger University Hospital, Stavanger, Norway
9Department of Research, Stavanger University Hospital, Stavanger, Norway
10Faculty of Health Sciences Department of Caring and Ethics, University of Stavanger, Stavanger, Norway
11Department of Breast and Endocrine Surgery, Haukeland University Hospital, Bergen, Norway
12Department of Pathology, Stavanger University Hospital, Stavanger, Norway
13Department of Medical Biochemistry and Pharmacology, Haukeland University Hospital, Bergen, Norway
14Department of Medicine, Haukeland University Hospital, Bergen, Norway
15Department of Clinical Medicine, University of Bergen, Bergen, Norway
16Central Hospital in Vestfold, Tønsberg, Norway
17Faculty of Medicine, Institute of Clinical Medicine, University of Oslo, Oslo, Norway
18Department of Epidemiology, Emory University, Atlanta, Georgia, USA
19Laboratory Medicine and Pathology, Haukeland University Hospital, Bergen, Norway
20Mathematics and Physics, Department of Mathematics and Natural Science, University of Stavanger, Stavanger, Norway

Twitter Birgitta Haga Gipsrud @BHagGipsrud

Acknowledgements
Very much appreciated are the contributions from the patients’ representatives in giving advice during the planning and conducting of the study. Indeed, our gratitude extends to all the breast cancer patients and breast cancer survivors that without hesitation and with great enthusiasm have joined the project. Importantly, they diligently continue take part in the long-term follow up in the PBCB project.

Contributors
HS: study idea, design, enrolment of patients, building database, writing of paper; EAMJ: study design, analysis of micro RNA, primary tumour and TEPs, writing of paper; TH: enrolment of patients, writing of paper; FME: analysis of drug concentrations in serum; MH: building database; ON: analysis of cdDNA, CTGs, next-generation sequencing, writing of paper; SL: enrolment of patients; handling of blood samples, biobanking, building database; THL: study idea, study design, enrolment of patients, writing of paper; JVS: building database; KT, analysis of cdDNA, CTGs and next-generation sequencing, writing of paper; BG: enrolment of patients and building av database, writing of paper; KJ: biobanking, analysis of microRNA and TEPs, writing of paper; EG: microscopic analysis of tissue section of the patients, KL: PROM data, writing of paper, KBGH, PROM data, writing of paper; BGH: PROM data, writing of paper, RL, PROM data, writing of paper; AH: enrolment of patients, TA: enrolment of patients, MA: next-generation sequencing of primary tumour and metastases, microRNA, TEPs and writing of paper, NGE, enrolment of patients, analysis of micro-RNA, writing of paper, TB: establishing database for PROM data, TLL: providing instrumental ideas to planning of the PBCB protocol, LS: biobanking; ACK, Biobanking; SD: handling of blood samples, biobanking; JTK: statistics; EAL: Study idea, design, LS: biobanking and GM, study idea, study design, writing of paper.

Funding
This work was supported by Western Norway Health Authorities grant number 911898. Very much appreciated is the funding from Folke Hermansen Cancer Foundation, Stavanger Norway. Also, invaluable support has been received from Inge Deanslands Stiftelse, Stavanger, Norway. Many thanks to Sparebankstiftelsen ‘Banka og Hå’, Time, Norway. Most appreciated is the very important financial support from ‘Sparebankstiftelsen SR-Bank’, Stavanger, Norway.

Competing interests
None declared.

Patient and public involvement
Patients and/or the public were involved in the design, or conduct, or reporting, or dissemination plans of this research. Refer to the Methods section for further details.

Patient consent for publication
Not applicable.

Provenance and peer review
Not commissioned; externally peer reviewed.

Open access
This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/.

ORCID iD
Hilvård Søiland http://orcid.org/0000-0002-9285-2774

REFERENCES


