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Monitoring SARS-CoV-2 incidence and seroconversion in a university cohort in California, June to August 2020

Journal:	BMJ Open
Manuscript ID	bmjopen-2022-063999
Article Type:	Original research
Date Submitted by the Author:	20-Apr-2022
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Keywords:	COVID-19, EPIDEMIOLOGY, Public health < INFECTIOUS DISEASES, Infection control < INFECTIOUS DISEASES, Epidemiology < INFECTIOUS DISEASES, PUBLIC HEALTH

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Title: Monitoring SARS-CoV-2 incidence and seroconversion in a university cohort in California, June to August 2020 Authors: Lauren A. Hunter, MPH¹; Stacia Wyman, PhD, MS²; Laura Packel, PhD¹; Shelley Facente, PhD^{1,3-4}; Yi Li, BS¹; Anna Harte, MD⁵; Guy Nicolette, MD⁵; the IGI SARS-CoV-2 Testing Consortium²; Clara Di Germanio, PhD⁴; Michael P. Busch, MD PhD^{4,6}; Arthur Reingold, MD¹; Maya Petersen, MD PhD¹ ¹ School of Public Health; University of California, Berkeley; Berkeley, CA ² Innovative Genomics Institute; University of California, Berkeley; Berkeley, CA ³ Facente Consulting; Richmond, CA ⁴ Vitalant Research Institute; San Francisco, CA ⁵ University Health Services; University of California, Berkeley; Berkeley, CA ⁶ Department of Laboratory Medicine; University of California, San Francisco; San Francisco, CA Correspondence to: Ms. Lauren Hunter **Division of Epidemiology and Biostatistics** University of California, Berkeley School of Public Health 2121 Berkeley Way #5302 Berkeley, CA 94720 lahunter@berkeley.edu **Word count:** 3,916 Abstract word count: 290 **Number of tables/figures:** 5 (2 supplementary)

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1 2		
3 4	27	Abstract
5 6	28	Objective: To inform effective SARS-CoV-2 mitigation strategies in university settings, we
7 8	29	piloted an integrated symptom and exposure monitoring and testing system among a cohort of
9 10	30	university students and employees.
11 12	31	Methods: We aimed to identify incident SARS-CoV-2 infections in a longitudinal cohort of 2,180
13 14	32	students and 738 employees of a public university in California from June to August 2020. At
15 16	33	baseline and endline, we tested participants for active SARS-CoV-2 infection via quantitative
17 18	34	polymerase chain reaction (qPCR) test and collected blood for antibody testing. Participants
19 20 21	35	received notifications to complete additional qPCR tests throughout the study if they reported
21 22 23	36	symptoms or exposures in daily surveys or were selected for surveillance testing. Viral whole
24 25	37	genome sequencing was performed on positive qPCR samples, and phylogenetic trees were
26 27	38	constructed with these genomes and external genomes retrieved from GISAID.
28 29	39	Results: Over the study period, 57 students (2.6%) and 3 employees (0.3%) were diagnosed
30 31	40	with SARS-CoV-2 infection via qPCR test. Phylogenetic analyses revealed that a super-
32 33	41	spreader event among undergraduates in congregate housing accounted for at least 48% of
34 35	42	cases but did not spread beyond campus. Test positivity was higher among participants who
36 37	43	self-reported symptoms (incidence rate ratio [IRR]: 12.4; 95% confidence interval [CI]: 7.3, 21.3)
38 39 40	44	or had household exposures (IRR: 12.3; 95% CI: 5.6, 26.9) which triggered notifications to test.
40 41 42	45	Most (91%) participants with newly identified antibodies at endline had been diagnosed with
43 44	46	incident infection via qPCR test during the study.
45 46	47	Conclusions: Our findings suggest that integrated monitoring systems can successfully identify
47 48	48	and link at-risk students to SARS-CoV-2 testing. Building upon such systems may prove key in
49 50	49	the next stage of the pandemic, as universities grapple with highly transmissible variants,
51 52	50	incomplete vaccine coverage and breakthrough infections, and reduced reliance on prevention
53 54	51	strategies such as masking and remote learning.
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57 58		

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2 3	52	Strongthe and limitations of this study
4 5		Strengths and limitations of this study
6	53	 The study is strengthened by rich longitudinal data including more than 117,000 daily
7 8	54	symptom surveys; 17,000 weekly exposure surveys; 7,600 qPCR tests to detect active
9 10	55	SARS-CoV-2 infection; and 4,900 antibody tests to detect previous infection collected
11 12	56	from 2,918 university students and employees over three months.
13 14	57	Using seroconversion data from serial antibody tests and phylogenetic analyses
15 16 17	58	comparing viral genome sequences to a broader database, we were able to evaluate the
18 19	59	extent to which the study system identified incident cases and contained an outbreak
20 21	60	among university students. However, our identification of participants who seroconverted
22 23	61	between baseline and endline may be incomplete due to loss-to-follow up and imperfect
24 25	62	sensitivity of SARS-CoV-2 antibody testing.
26 27	63	 A high proportion of identified cases were traced to one outbreak, limiting the
28 29	64	generalizability of our exploratory assessment of risk factors for incident infection. While
30 31	65	self-referral into the study in the context of the outbreak is likely to induce selection bias,
32 33	66	it also illustrates the utility of implementing non-stigmatizing, incentivized testing
34 35	67	approaches to increase testing uptake among at-risk students.
36 37 38	68	As the study took place before the development of highly transmissible variants and
39 40	69	vaccine rollout, further research is necessary to adapt and evaluate similar systems in
41 42	70	the context of both heightened transmissibility and more prevalent natural and vaccine-
43 44	71	induced immunity.
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3 4	72	Background
5 6	73	Universities have been identified as hotspots for SARS-CoV-2 transmission in the United
7 8	74	States, ¹ where SARS-CoV-2 incidence is highest among young adults. ² Young adults may be
9 10	75	less likely to adhere to social distancing guidelines and more likely to experience workplace
11 12	76	exposure (for example, at food service or retail jobs). ² Their risk may be heightened in university
13 14	77	settings where many live in congregate housing, interact with wide social networks, or attend
15 16 17	78	large gatherings. ³ Although young adults are at low risk of serious acute illness or death from
17 18 19	79	COVID-19 (the disease caused by SARS-CoV-2), ⁴ the higher likelihood of asymptomatic or
20 21	80	mildly symptomatic infection in this age group makes young adults a key population through
22 23	81	which SARS-CoV-2 may be spread to other, more vulnerable groups. ^{2,5} Indeed, there is
24 25	82	evidence that transmission among university students may lead to increased COVID-19-related
26 27	83	mortality in the surrounding counties. ^{6–8} Although widespread vaccination has enabled most
28 29	84	campuses to return to in-person activities, the elimination of SARS-CoV-2 transmission in
30 31	85	campus populations may be stymied by vaccine hesitancy among students and employees and
32 33	86	breakthrough infection and subsequent transmission by vaccinated persons, particularly in the
34 35	87	context of waning immunity and viral variants which reduce vaccine efficacy.9,10 Therefore, rapid
36 37	88	and resource-efficient identification of incident cases in university populations is a critical first
38 39 40	89	step of outbreak investigation and control, followed by isolation, case investigation, and contact
41 42	90	tracing, to minimize transmission within campus and to the broader community.
43 44	91	Universities have adopted a wide range of approaches for testing and outbreak
45 46	92	mitigation. ^{11–13} While a number of well-resourced universities have scaled up testing capacity in
47 48	93	order to frequently test all students and employees accessing campus or living in university-

94 affiliated housing,¹³ many other universities do not have well-defined testing strategies or restrict

95 testing to those with symptoms or known exposure.¹² Beyond investing in testing programs,

96 some universities have sought to reduce on-campus transmission by mandating the completion

97 of self-administered symptom screening tools by students and employees. However, such tools

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have primarily been used to regulate daily access to campus (i.e., deny entry to those who
report COVID-19-like symptoms), rather than to detect emergent outbreaks among university
populations. As universities resume normal operations and discontinue mitigation strategies
such as masking, non-punitive, resource-efficient strategies which can both identify those who
are at highest risk of infection *and* expediently link them to low-barrier testing services may play
a key role in transitioning from a "one-size-fits-all" approach of uniform testing to a sustainable
monitoring paradigm.

In 2020, we piloted an integrated symptom and exposure monitoring and testing system designed to identify incident SARS-CoV-2 infections among a cohort of university students and employees.¹⁴ Here we describe the incidence and seroprevalence of SARS-CoV-2 infection within this cohort to evaluate the extent to which incident infections were successfully detected and contained over the study period, identify sociodemographic factors associated with incident infection, and ascertain which self-reported symptoms and exposures tracked by the monitoring system were predictive of test positivity, with the ultimate objective of informing monitoring and testing strategies in university settings.

5 113

7 114 Methods

115 Study design and setting

The study comprised three prospective cohorts of University of California, Berkeley affiliates followed from June to August 2020: students, essential workers (i.e., employees working on campus in health, facilities, or key student services), and other employees (hereafter, "faculty/staff"). We report the findings according to the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) checklist for cohort studies.¹⁵ Throughout the study period, UC Berkeley did not offer in-person classes, and on-campus work was restricted to essential workers and a small subset of faculty, staff, and student researchers. Although few students were living in on-campus residence halls, many

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students continued to live in congregate living settings off campus, such as fraternities,	MJ Op
sororities, and co-operative housing.	MJ Open: first published as 10.1136/bmjopen-2022-063999 on 6 April 2023. Downloaded from http://bmjopen.bmj.com/ on April 20, 2024 by guest. Protected by copyright.
	publish
Participant recruitment and eligibility	ed as
The study was promoted through targeted messages from university officials to campus	10.11
email listservs and social media platforms from early June to mid-July 2020. To increase reach	36/bm
to students expected to be at higher risk of COVID-19, we also placed flyers in congregate livin	g jopen-
settings and conducted in-person recruitment for student athletes who had resumed training on	2022-
campus. Participants were eligible to enroll in the study if they were at least 18 years of age,	06399
were a current student or employee at UC Berkeley, and planned to live in or near Berkeley	9 on 6
during summer 2020. Specific eligibility criteria and enrollment windows varied by cohort	April
(Supplementary Table 1, Supplementary Figure 1).	<u>2</u> 023. [
Upon enrollment, participants were linked to an online baseline survey that collected	Downle
sociodemographic data and information about their COVID-19-related health history.	baded
Participants were then referred to a baseline testing appointment at University Health Services	from h
(UHS) which included a SARS-CoV-2 quantitative polymerase chain reaction (qPCR) test and	ttp://br
blood collection for antibody testing (procedures described below). To facilitate daily	njoper
temperature monitoring, study staff also provided participants with free oral thermometers upon	ı.bmj.o
request at testing appointments. Participants who completed this appointment or a non-study	om/ o
qPCR test at UHS by July 20th were eligible to remain in the study. We pre-specified a	n April
maximum sample size of 4,000 participants across cohorts but did not reach this limit before the	e 20, 20
final day of baseline data collection.)24 by (
	guest.
Symptom and exposure surveys	Protec
Participants received daily text messages or emails, depending on their preference	cted by
specified in the baseline survey, which linked to short symptom surveys through which they	/ сору
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2 3 4	150	reported their daily body temperature and any symptoms of illness. Once per week, the daily
5 6	151	survey included a longer exposure module, which asked about recent symptoms of illness
7 8	152	among their household member(s), potential exposure(s) to COVID-19, and activities related to
9 10	153	potential COVID-19 risk. All surveys were administered via REDCap. ^{16,17}
11 12	154	
13 14	155	Endline survey and testing
15 16	156	In early August, participants were sent an endline survey which collected updated
17 18	157	information on their COVID-19 history to identify any diagnoses outside of the study.
19 20	158	Participants in the student and essential worker cohorts were also invited to complete endline
21 22	159	testing appointments by August 18th, including a final qPCR test and blood collection.
23 24	160	
25 26 27	161	qPCR testing
27 28 29	162	Midturbinate nasal and oral swabs were collected by UHS clinical staff and tested for
30 31	163	SARS-CoV-2 by qPCR at the Innovative Genomics Institute (IGI).18 qPCR tests were performed
32 33	164	at baseline for all three cohorts and at endline for the student and essential worker cohorts.
34 35	165	Between baseline and endline testing, additional qPCR tests were performed for the following
36 37	166	reasons:
38 39	167	 Symptom- or exposure-based tests triggered based on participants' responses in
40 41	168	daily surveys: Participants who reported COVID-19-like signs or symptoms ¹ (in
42 43	169	themselves or household member(s)) or who reported a suspected or confirmed COVID-
44 45 46	170	19 case in their household were automatically notified to sign up for a qPCR test.
40 47 48		
49 50		
50 51 52		
52		

¹ Signs or symptoms which triggered a testing notification when reported were: temperature of ≥100.4°F, dry cough (without mucus), coughing up mucus, feeling feverish, unusual pain or pressure in the chest, difficulty breathing, shortness of breath, unexplained trouble thinking or concentrating, loss of sense of taste, or loss of sense of smell.

1 2		
3 4	171	• Random surveillance testing: A subset of participants in the student and faculty/staff
5 6	172	cohorts who had not had a qPCR test within a week were randomly selected and
7 8	173	emailed notifications to come in for surveillance testing in July.
9 10	174	Address-based surveillance testing: Participants who lived at the same address as
11 12	175	another participant who tested positive for SARS-CoV-2 were immediately emailed
13 14	176	surveillance testing notifications. Following an outbreak among group-housed students
15 16	177	in early July, surveillance testing notifications were also emailed to all participants who
17 18 19	178	had not been tested within the week and who reported living in fraternities, sororities, or
20 21	179	co-operative housing.
22 23	180	Participant-initiated testing: Participants could self-schedule study testing
24 25	181	appointments on demand, with or without consulting a healthcare provider and
26 27	182	regardless of exposure history.
28 29	183	Participants with positive qPCR test results were informed by phone by UHS clinical staff, who
30 31	184	provided guidance on isolation and performed case investigation to identify potential contacts.
32 33	185	Participants with negative qPCR test results were informed of their results via the UHS online
34 35	186	patient portal.
36 37	187	
38 39 40	188	SARS-CoV-2 sequencing and phylogenetic analyses
40 41 42	189	Viral whole genome sequencing was performed on a set of positive samples at the IGI,
43 44	190	using previously described procedures. ¹⁹ Briefly, SARS-CoV-2 RNA extracted from swabs was
45 46	191	reverse transcribed using SuperScript IV (Invitrogen), and the viral genome was amplified from
47 48	192	the resulting cDNA in four separate qPCR reactions using distinct primer sets tiling the SARS-
49 50	193	CoV-2 genome. The four qPCR reactions were pooled 1:1:1:1 and diluted 1:50 in H_2O . A
51 52	194	second qPCR reaction was set up to add Nextera Unique Dual Indexing (UDI) sequences to
53 54	195	either end of the amplicons. The resulting qPCR reaction was cleaned up using 0.7x AMPureXP
55 56	196	beads (Beckman Coulter) and quantified using a Qubit dsDNA HS Assay Kit (Thermo Fisher).
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59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

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The libraries were then pooled to an equimolar ratio and sequenced with a 10% PhiX spike in using a MiSeq v3 kit at 300bp PE reads.

Fastq sequencing files were processed through a custom pipeline using publicly available software. The reads were preprocessed by guality trimming, removing adaptors, and PhiX cleaning with BBTools,²⁰ and then aligned to the Wuhan reference sequence (NC 045512.2) with minimap2 v2.16-r922. ARTICv3 primers were trimmed, and the consensus sequence was built with iVar v1.3.1, where an 'N' is called if the depth is less than 10 reads at any nucleotide. The genomes were then processed through the Nextstrain Auger pipeline with other genomes from GISAID to construct a maximum likelihood tree.^{21,22} Several phylogenies were constructed for this analysis: a tree of 7,091 genomes subsampled from the worldwide genomes in GISAID at the time (approximately 200,000 genomes as of October 2020) was used to place the IGI genomes in the larger tree; a tree with all IGI genomes sequenced at the time of analysis (356 genomes); and a tree containing 500 genomes (from 1 million genomes as of April 2021) was constructed using UShER.23

Antibody testing

Up to 10 mL of blood was collected by phlebotomists via venipuncture at baseline from participants in all three cohorts and again at endline from participants in the student and essential worker cohorts. Blood was centrifuged and serum was stored at -20°C for 2 to 4 months before being tested at Vitalant Research Institute using the VITROS Immunodiagnostic Products Anti-SARS-CoV-2 Total Reagent Pack, which detects IgA, IgG, and IgM antibodies and has an estimated clinical specificity of 100% and unreported sensitivity.24

Participant compensation

Participants in the student cohort received a \$50 gift card after completing baseline testing and 10 daily surveys; this incentive was conditional on daily survey completion to

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223	encourage early habit formation. ²⁵ Student participants received a second \$50 gift card at their	
224	endline testing appointment. To facilitate travel to and from UHS for testing appointments,	
225	student participants were also offered pre-paid car rides via a ride-sharing app.	
226	Participants in the essential worker cohort received a gift card worth \$1 per daily survey	
227	completed (to a maximum of \$70) after the study ended. Participants in the faculty/staff cohort	
228	were not compensated.	
229		
230	Statistical analyses	
231	To identify sociodemographic factors associated with incident infection, we used Poissor	ı
232	regression to estimate unadjusted incidence rate ratios (IRRs) for SARS-CoV-2 infection by	
233	study cohort and within strata of sociodemographic variables self-reported in the baseline	
234	survey (e.g., age, gender, housing type), setting person-months of enrollment as an offset term	
235	to account for differing lengths of follow-up.	
236	We also calculated IRRs comparing test positivity by recent signs/symptoms, exposures,	,
237	and activities reported in the daily and weekly surveys. We estimated IRRs for several	
238	temperature thresholds (i.e., ≥100.4°F, ≥100.0°F, ≥99.0°F) to compare to symptom-specific	
239	IRRs; however, continuous associations between temperature and positivity have been	
240	previously explored in this cohort. ²⁶ We accounted for clustered observations due to repeated	
241	tests per participant using a generalized estimating equation approach with Huber-White	
242	standard error estimates and an exchangeable working correlation structure. ²⁷	
243	Finally, to assess the extent to which the testing and monitoring system captured	
244	incident infections, we identified participants who seroconverted from having non-reactive (no	
245	antibodies detected) to reactive (antibodies detected) blood samples between baseline and	
246	endline and calculated the proportion of these participants who were also diagnosed with	
247	incident SARS-CoV-2 infection via positive qPCR test during the study period. Analyses were	
248	conducted in R version 4.0.4. ²⁸	
	10)
	225 226 227 228 230 231 232 233 234 235 236 237 238 239 240 241 242 243 244 245 244 245 246	 endline testing appointment. To facilitate travel to and from UHS for testing appointments, student participants were also offered pre-paid car rides via a ride-sharing app. Participants in the essential worker cohort received a gift card worth \$1 per daily survey completed (to a maximum of \$70) after the study ended. Participants in the faculty/staff cohort were not compensated. Statistical analyses To identify sociodemographic factors associated with incident infection, we used Poissor regression to estimate unadjusted incidence rate ratios (IRRs) for SARS-CoV-2 infection by study cohort and within strata of sociodemographic variables self-reported in the baseline survey (e.g., age, gender, housing type), setting person-months of enrollment as an offset term to account for differing lengths of follow-up. We also calculated IRRs comparing test positivity by recent signs/symptoms, exposures and activities reported in the daily and weekly surveys. We estimated IRRs for several temperature thresholds (i.e., ≥100.4°F, ≥100.0°F, ≥99.0°F) to compare to symptom-specific IRRs; however, continuous associations between temperature and positivity have been previously explored in this cohort ²⁰ We accounted for clustered observations due to repeated tests per participant using a generalized estimating equation approach with Huber-White standard error estimates and an exchangeable working correlation structure.²⁷ Finally, to assess the extent to which the testing and monitoring system captured incident infections, we identified participants who sero also diagnosed with incident SARS-CoV-2 infection via positive qPCR test during the study period. Analyses were conducted in R version 4.0.4.²⁸

1 2		
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5 6	250	Ethical approvals
7 8	251	All study activities were approved by the University of California, Berkeley Committee for
9 10	252	the Protection of Human Subjects (#2020-06-13349, #2020-05-13261, #2020-04-13238).
11 12	253	
13 14	254	Patient and public involvement
15 16	255	The study's target population comprised university students and employees. While the
17 18 19	256	study was conducted by faculty, staff, and graduate students from the UC Berkeley School of
20 21	257	Public Health, University Health Services, and the Innovative Genomics Institute, the broader
22 23	258	student body and university workforce were not involved in designing the study or selecting the
24 25	259	research question, outcome measures, or method of disseminating results.
26 27	260	
28 29	261	Results
30 31	262	Participant recruitment and retention
32 33	263	Between June 1 and July 20th, 2020, we enrolled 2,180 students, 268 essential workers,
34 35	264	and 470 faculty/staff who completed at least one qPCR test or antibody test (Table 1,
36 37 38	265	Supplementary Figure 1). The student cohort was split between undergraduate (52%) and
38 39 40	266	graduate (48%) students. Nearly half (44%) of essential workers worked in health services.
41 42	267	While 85% of essential workers were working on campus at the time of enrollment, most (81%)
43 44	268	faculty/staff were working entirely remotely. At the time of enrollment, only 12 (0.4%)
45 46	269	participants reported a previous COVID-19 diagnosis.
47 48	270	Participants provided a total of 5,545 person-months of follow-up from enrollment to the
49 50	271	end of the study (mean person-days per participant: 57, range: 32-78). Participants completed a
51 52	272	mean of 40 daily symptom surveys and 6 weekly exposure surveys over the study period, for a
53 54	273	total of 117,235 symptom and 17,172 exposure surveys. A subset of participants did not
55 56 57	274	complete any daily symptom surveys (1.7%) or weekly exposure surveys (4.2%).
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Table 1. Baseline characteristics of participants in the Berkeley COVID-19 Safe Campus

276 Initiative by study cohort, June-August 2020.

	All	Students	Essential Workers	Faculty/Staff
N (row %)	2,918 (100)	2,180 (74.7)	268 (9.2)	470 (16.1)
Age, mean ± SD	29.4 ± 11.6	24.3 ± 5.4	42.5 ± 12.3	45.2 ± 12.3
Gender, n (column %) Man Woman Non-binary/other	1,177 (40.3) 1,653 (56.6) 51 (1.7)	911 (41.8) 1,187 (54.4) 46 (2.1)	103 (38.4) 164 (61.2) 1 (0.4)	163 (34.7) 302 (64.3) 4 (0.9)
Race/ethnicity, n (column %)* American Indian/Alaska Native Asian/Pacific Islander Black/African American Hispanic/Latine/Spanish origin White Other	39 (1.3) 833 (28.5) 103 (3.5) 420 (14.4) 1,814 (62.2) 280 (9.6)	29 (1.3) 703 (32.2) 83 (3.8) 346 (15.9) 1,261 (57.8) 223 (10.2)	2 (0.7) 66 (24.6) 16 (6.0) 39 (14.6) 160 (59.7) 31 (11.6)	8 (1.7) 64 (13.6) 4 (0.9) 35 (7.4) 393 (83.6) 26 (5.5)
Program level, n (column %) Undergraduate Graduate	-	1,114 (51.7) 1,039 (48.2)	-	-
Living at fraternity/sorority, n (column %)	-	125 (5.7%)	-	-
Education, n (column %) High school diploma/GED Some college or trade school Bachelor's degree Graduate/professional degree	- - -	- - -	6 (2.2) 59 (22.0) 78 (29.1) 121 (45.1)	0 (0) 13 (2.8) 119 (25.3) 337 (71.7)
Department, n (column %) Health services Facilities/building services Student services/other	- -	- -	129 (48.1) 61 (22.8) 77 (28.7)	-
Job title, n (column %) Faculty Staff Postdoctoral scholar/other	- -	-	- - -	110 (23.4) 311 (66.2) 49 (10.4)
Currently working outside the home, n (column %)	748 (25.6)	418 (19.2)	228 (85.1)	102 (21.7)
Pre-enrollment COVID-19 diagnosis, n (column %)	12 (0.4)	8 (0.4)	1 (0.4)	3 (0.6)

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279 SARS-CoV-2 incidence

During the study period, participants underwent 7,638 qPCR tests for active SARS-CoV2 infection, with a mean of 2.6 tests per participant (range: 0-9). Almost all (99.9%) participants
completed at least one qPCR test. Overall, 60 participants (2.0%) tested positive: 57 students, 2
essential workers, and 1 faculty/staff.

Among cohorts, students were at highest risk of incident infection over the study period (IRR students vs. faculty/staff: 5.83; 95% confidence interval [CI]: 1.28, 102.99). Due to the low number of cases outside of the student cohort, we examined additional risk factors for infections among students only (Table 2), finding higher rates of infection among students who were 18-19 years old (IRR vs. students ≥22 years: 8.34; 95% CI: 4.17, 17.48) and undergraduates (IRR vs. graduate students: 4.12; 95% CI: 2.17, 8.66). We also observed a higher incidence among white students (IRR: 2.80 vs. non-white students; 95% CI: 1.53, 5.54). These associations were largely driven by an outbreak among participants living in fraternities or sororities. Nearly one-quarter of participants living in fraternities or sororities were infected with SARS-CoV-2 during the study period (IRR vs. other students: 20.86; 95% CI: 12.27, 35.54), and these participants accounted for 49% of cases observed among student participants.

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Table 2. Bivariate associations between sociodemographic characteristics and SARS-CoV-2
 incidence among student participants in the Safe Campus Initiative, June-August 2020.

		Cases, N (row %)	Non-Cases, N (row %)	IRR (95% CI)
Ô١	verall*	57 (2.6)	2,120 (97.4)	
Ag				
	18-19 years	21 (8.0)	243 (92.0)	8.34 (4.17, 17.48)
	20-21 years	24 (3.8)	607 (96.2)	4.15 (2.11, 8.58
2	≥22 years	12 (0.9)	1,270 (99.1)	Reference
	ender			
	Woman	37 (3.1)	1,147 (96.9)	1.45 (0.85, 2.58
	Van	19 (2.1)	892 (97.9)	Reference
1	Non-binary/other	0 (0)	46 (100)	
Ra	ace/ethnicity**			
	American Indian/Alaska Native	0 (0)	29 (100)	
/	Asian/Pacific Islander	11 (1.6)	691 (98.4)	0.49 (0.24, 0.91
E	Black/African American	1 (1.2)	82 (98.8)	0.45 (0.03, 2.03
ł	Hispanic/Latine/Spanish origin	8 (2.3)	337 (97.7)	0.88 (0.39, 1.76
١	White	45 (3.6)	1,216 (96.4)	2.80 (1.53, 5.54
(Other	4 (1.8)	217 (98.2)	0.65 (0.20, 1.58
Pr	ogram level			
l	Jndergraduate	46 (4.1)	1,067 (95.9)	4.12 (2.17, 8.66
(Graduate	10 (1.0)	1,027 (99.0)	Reference
Liv	ving at fraternity/sorority	28 (22.4)	97 (77.6)	20.86 (12.27, 35.54
		6 (1.4)	410 (98.6)	0.51 (0.20, 1.11

40 302 Phylogenetic analysis
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⁴² 303 We retrieved whole viral genome sequences for 35 of the 60 positive cases from this

study, 29 (83%) of which were found to be part of a campus super-spreader event involving a

47 305 total of 57 campus-affiliated individuals with samples sequenced by IGI (Figure 1A). Most (69%)

 $_{49}^{48}$ 306 study participants within this cluster lived at one of two residences, with likely a single

51 307 participant originating the super-spreader event. The cluster of genomes was defined by three

53 308 mutations (A6360G, C24502A and G110083T), two of which were extremely rare at the time of

55 309 the outbreak. The combination of the three variants was only found in four genomes outside of

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this cluster (two in the UK and two in Florida) by October 2020, making it a strong phylogenetic signature. Phylogenetic analysis demonstrated that the cluster remained confined to campus, as this signature was not observed in any genomes from samples in the surrounding communities or California state in the months following the super-spreader event. When the trio of mutations was searched in a phylogeny constructed from over 1.2 million genomes worldwide using UShER in April 2021,²³ no descendent leaves were found in the tree under the cluster (Figure 1B), indicating that the lineage died out after the super-spreader event. Factors associated with test positivity At least one symptom survey was completed in the 7 days before sample collection for 90% of tests (n=6,864), including 72% of tests (n=5,469) that had symptom data from the day of sample collection. Of the 54 cases who completed at least one survey during the week before their positive sample was collected (mean: 4 surveys), 23 cases (43%) had reported at least one of the nine COVID-19 symptoms that triggered a notification for them to test. Test positivity was 12.4 times higher among participants who had a recent symptom-triggered notification (95% CI: 7.3, 21.3) (Table 3). Notification-triggering symptoms most strongly associated with test positivity included loss of sense of taste or smell and feeling feverish. Weakness, sweats or chills, and swollen glands were the non-triggering symptoms most strongly associated with test positivity.

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3	330	Table 3. Bivariate associations between prospectively monitored symptoms and exposures and
1 -	331	SARS-CoV-2 qPCR test positivity among participants in the Safe Campus Initiative, June-
) -	332	August 2020.

age 17 of 31	BMJ Open					
330 331 332	Table 3. Bivariate associations between prospectively monitored symptoms and exposures and SARS-CoV-2 qPCR test positivity among participants in the Safe Campus Initiative, June-August 2020.					
, })		Test Positivity, % (+ Tests / All Tests)	IRR (95% CI)			
)	Overall*	0.8 (60 / 7,629)	-			
1 2 3 4 5 6 7 8 9 0 1 2 3 4 5 8 9 0 1 2 3 4 5 6 7 8 9 0 1 2 3 4 5 8 9 0 1 2 3 4 5 8 9 0 1 2 3 4 5 8 9 0 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 4 5 8 9 0 1 1 2 3 8 9 0 1 2 3 4 5 8 9 0 1 1 2 3 8 9 0 1 2 8 9 0 1 2 3 8 9 0 1 1 2 8 9 0 1 1 2 8 9 0 1 2 3 8 9 0 1 8 9 0 1 2 3 8 9 1 8 9 1 8 9 1 8 9 1 8 9 1 8 9 1 8 1 8	Signs/symptoms within 7 days of test No Yes (any) - Temperature ≥100.4°F† - Temperature ≥99.0°F - Temperature ≥99.0°F - Feeling feverish† - Dry cough† - Coughing up mucus† - Unusual chest pain or pressure† - Difficulty breathing† - Shortness of breath† - Trouble thinking/concentrating† - Loss of sense of taste† - Loss of sense of smell † - Any notification-triggering symptom† - Loss of appetite - Fatigue - Trouble sleeping - Headache - Runny, blocked, or painful sinuses - Sneezing - Swollen, red, or painful eyes - Sore throat - Diarrhea - Nausea or vomiting - Body aches or muscle pain	$\begin{array}{c} 0.4 \ (21 \ / \ 5, 704) \\ 3.2 \ (31 \ / \ 971) \\ 0.0 \ (0 \ / \ 8) \\ 11.8 \ (2 \ / \ 17) \\ 2.6 \ (9 \ / \ 346) \\ 14.9 \ (11 \ / \ 74) \\ 5.5 \ (7 \ / \ 128) \\ 5.5 \ (5 \ / \ 91) \\ 9.7 \ (6 \ / \ 62) \\ 5.6 \ (1 \ / \ 18) \\ 8.7 \ (4 \ / \ 46) \\ 7.6 \ (5 \ / \ 66) \\ 42.9 \ (3 \ / \ 7) \\ 33.3 \ (4 \ / \ 12) \\ 5.8 \ (23 \ / \ 397) \\ 10.0 \ (6 \ / \ 60) \\ 3.5 \ (13 \ / \ 373) \\ 5.1 \ (7 \ / \ 137) \\ 4.6 \ (14 \ / \ 302) \\ 5.2 \ (14 \ / \ 268) \\ 1.9 \ (2 \ / \ 104) \\ 8.6 \ (5 \ / \ 53) \\ 3.1 \ (8 \ / \ 259) \\ 5.8 \ (5 \ / \ 86) \\ 4.8 \ (4 \ / \ 83) \\ 3.3 \ (3 \ / \ 92) \\ 8.1 \ (12 \ / \ 149) \end{array}$	4.4 (1.4, 13.6) 13.0 (7.0, 24.4)			
<u>2</u>	- Sweats or chills	11.3 (10 / 89)				
	Swollen glandsWeakness	13.2 (10 / 76)	16.6 (7.0, 39.7) 20.5 (10.6, 39.4)			
	Exposures within 14 days before test	10.2 (10770)	_0.0 (10.0, 00.1)			
	No	0.3 (14 / 4,179)	Reference			
	Yes (any) - Suspected or confirmed COVID-19 case in household [†]	3.4 (17 / 499) 6.7 (6 / 89)	10.1 (5.0, 20.4) 14.7 (6.0, 35.9)			
	 Close contact with suspected or confirmed case outside household 	2.9 (4 / 138)	6.3 (2.2, 18.2)			
2	- Household member with new	4.4 (5 / 114)	7.6 (3.0, 19.6)			
3 4 5	 COVID-19-like symptoms [†] Household member with any new symptoms of illness 	2.4 (8 / 336)	4.7 (2.1, 10.4)			
5 7				4.0		

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1 2		
3		- Any notification-triggering exposure [†] 5.2 (9 / 173) 12.3 (5.6, 26.9)
4		Activities within 14 days before test
5 6		No 0.5 (3 / 630) Reference
7		Yes (any) 0.7 (29 / 4,142) 1.5 (0.5, 4.8)
8		- Spent time at another residence 1.1 (26 / 2,330) 4.6 (1.9, 11.1)
9		- Had visitors at own residence 1.0 (22 / 2,203) 2.5 (1.2, 5.4)
10		- Attended gathering >10 people 2.8 (19 / 672) 9.0 (4.4, 18.1)
11		- Worked outside of home 0.5 (10 / 2,132) 0.6 (0.3, 1.2)
12		- Used public restroom 0.7 (12 / 1,830) 1.0 (0.5, 2.0)
13		- Used public transportation 0.6 (5 / 695) 0.8 (0.3, 2.3)
14 15		- Participated in group sports 1.6 (4 / 255) 2.6 (0.9, 7.3)
16	333	qPCR: quantitative polymerase chain reaction, IRR: incidence rate ratio, CI: confidence interval.
17	334	*Excluding resamples and repeated positives; includes N=2,914 participants with at least one qPCR test for SARS-
18	335 336	CoV-2 during the study period. [†] Reporting triggered notification to test.
19	337	Reporting triggered notification to test.
20	007	
21 22	338	Participants completed at least one weekly exposure survey in the 14 days before
22		
24	339	sample collection for 61% of tests (n=4,678). Of the 31 cases who had recently completed an
25	o 4 o	
26	340	exposure survey at the time of sample collection, 9 (29%) reported a potential household
27 28	341	exposure that triggered a notification for them to test (Table 3). Test positivity was 12.3 times
28 29	541	exposure that inggered a notification for them to test (Table 3). Test positivity was 12.3 times
30	342	higher among participants who had a recent exposure-triggered notification (95% CI: 5.6, 26.9).
31	•	
32	343	Test positivity was also significantly higher among participants who reported recent engagement
33 34	~	
35	344	in 'higher risk' social activities, most notably attending a gathering of more than 10 people (IRR:
36	345	9.0; 95% CI: 4.4, 18.1).
37	••••	
38 39	346	
39 40		
41	347	SARS-CoV-2 seroprevalence
42	0.40	
43	348	Only 18 (0.6%) of 2,877 participants who provided blood samples at baseline had
44	349	SARS-CoV-2 antibodies (Table 4), all but one of them students. Most participants with
45 46	040	
47	350	antibodies at baseline either suspected past infection (28%), had been previously diagnosed
48		· · · · · · · · · · · · · · · · · · ·
49	351	(22%), or had a positive qPCR test the day blood was drawn (11%). Most (85%) participants in
50		
51 52	352	the student and essential worker cohorts provided blood samples at both baseline and endline
52 53	353	(mean interval between complex: 48 days). Among 2.076 participants with baseling and andling
54	555	(mean interval between samples: 48 days). Among 2,076 participants with baseline and endline
55	354	blood samples, 33 (1.6%) seroconverted from non-reactive at baseline to reactive at endline, 30
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2 3 4	355	of whom (91%) were also diagnosed via qPCR test during the study. Of the three participants						
5 6	356	who seroconverted without a positive qPCR test, two self-reported suspected past infection (one						
7 8	357	st infection and						
9 10	358	had four negative qPCR tests over 40 days of study participation.						
11 12	359	Of the 60 participants with inciden	t SARS-CoV-2 infection	ction during the stu	dy period, 41			
13 14	360	(68%) provided an endline blood sample at least one week after the date of their first positive						
15 16	361	qPCR test (mean time between positive qPCR test and blood sample: 36 days; range 13-52						
17 18	362	days). Of these, 34 (83%) were reactive (Table 4).						
19 20	363							
21 22 23 24	364 365	Table 4. Seroprevalence of SARS-CoV-2Initiative, June-August 2020.	antibodies among	participants in the	Safe Campus			
25 26		9	Baseline, N (%)	Endline, N (%)	Both, N (%)			
27 28 29 30		Serostatus – Cross-sectional* Reactive Non-reactive	18 (0.6) 2,859 (99.4)	48 (2.3) 2,039 (97.7)	-			
31 32 33 34 35 36		Serostatus – Longitudinal** Non-Reactive → Non-Reactive Non-Reactive → Reactive Reactive → Non-Reactive Reactive → Reactive	-	-	2,029 (97.7) 33 (1.6) 0 (0) 14 (0.7)			
37 38 39		Serostatus – Previous qPCR Positive [†] Reactive Non-reactive	-	34 (82.9) 7 (17.1)	-			
40 41 42 43 44 45 46	366 367 368 369 370 371	qPCR: quantitative polymerase chain reaction. *N=2,888 participants who provided at least one bl **N=2,076 participants who provided blood sample [†] N=41 participants who provided an endline blood positive qPCR test.	es at baseline and endlin		/-2 identified via			
47 48	372	Discussion						
49 50	373	This study provides a model of a voluntary, incentivized system to identify and link at-risk						
51 52 53	374	students to SARS-CoV-2 testing. While the	ne incidence and se	roprevalence of SA	ARS-CoV-2 were			
54 55	375	generally low in this cohort of university s	tudents and employ	ees in the summe	r of 2020, we			
56 57 58 59 60		For peer review only - http://b	omjopen.bmj.com/site/a	about/guidelines.xhtr	18 nl			

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observed the highest incidence among undergraduate students living in congregate settings,with nearly half of cases found to be associated with a super-spreader event.

Within this cohort, we previously demonstrated the acceptability of our low-barrier SARS-CoV-2 mitigation approach and the limitations of temperature monitoring as a tool for case identification.^{14,26} The present analysis builds upon these contributions by triangulating prospective gPCR testing data with phylogenetic analyses of positive samples and serial antibody testing to evaluate whether case identification and containment were achieved. In doing so, we found evidence that the system successfully identified a high proportion of incident SARS-CoV-2 cases among participants and may have mitigated community transmission after an outbreak. Specifically, 91% of participants with newly-identified antibodies for SARS-CoV-2 at the end of the study had also been diagnosed with incident infection via gPCR test during the study period. While a sizeable cluster of cases among participants was traced to a single super-spreader event, the associated cluster lineage was successfully contained without spreading beyond campus. As the outbreak unfolded, the system also allowed for rapid real-time response (i.e., surveillance testing notifications to students living in congregate housing) and offered a readily accessible, incentivized entry point for testing for students concerned about potential exposure.

Although some universities have adopted punitive measures intended to prevent transmission by controlling student behavior (for example, suspending students for hosting gatherings),^{29–31} this approach has been criticized for its potential to reduce students' trust and cooperation.³²⁻³⁴ Instead of punishing or shaming students who fail to adhere to public health guidance, some epidemiologists have called for a harm-reduction approach which supports and engages students as part of the solution.^{32–34} The present study reinforces the potential to integrate voluntary testing and risk monitoring systems to support targeted case identification, as evidenced by the significantly higher positivity rates found among participants whose self-reported symptoms and exposures triggered notifications to test. Our findings also support

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402 increased outreach to groups of students at highest risk, particularly younger students in403 congregate housing.

This study is strengthened by rich longitudinal data, including symptom and exposure tracking, gPCR testing, and seroprevalence data from more than 2,000 participants. The study population comprised of a broad sample of university affiliates, both students and employees, with strong representation of university subpopulations perceived to be at higher risk of infection (e.g., undergraduates, essential healthcare workers). As on-campus activities were severely restricted throughout the study period (all classes were held online, and few students were living in residence halls), this study cannot provide insight into SARS-CoV-2 transmission risks related to on-campus student activities. Nevertheless, as 73% of UC Berkeley undergraduate students lived off campus before the pandemic,³⁵ systems to detect off-campus (i.e., community and household) transmission remain important for SARS-CoV-2 monitoring efforts among students. Additionally, all participants in the essential workers cohort and a subset of participants in the faculty/cohort were working on campus during the study period, further motivating efforts to monitor incidence in this population.

There remain several limitations. We observed relatively few SARS-CoV-2 cases during the study period, which took place before the development of highly transmissible variants, such as Delta and Omicron, and before vaccine rollout. Further research is necessary to adapt and evaluate similar systems in the context of both heightened transmissibility and more prevalent natural and vaccine-induced immunity. Observed associations between symptoms and positivity may also differ among those who have been infected by more recent variants and/or vaccinated. Additionally, a high proportion of identified cases were traced to one outbreak, limiting the generalizability of our exploratory assessment of risk factors for incident infection. There was also anecdotal evidence that the outbreak prompted exposed students to enroll as study participants.¹⁴ While this self-referral into the study is likely to increase selection bias, it also illustrates the utility of implementing non-stigmatizing, incentivized testing approaches to

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428 increase testing uptake among at-risk students. Finally, our identification of participants who
429 seroconverted between baseline and endline may be incomplete due to loss-to-follow up and
430 imperfect sensitivity of SARS-CoV-2 antibody testing.

By integrating symptom and exposure monitoring systems with low-barrier testing, we identified incident SARS-CoV-2 infections to reduce transmission within a university setting. While there have been seismic shifts in the SARS-CoV-2 pandemic since 2020, universities continue to grapple with how best to mitigate on-campus spread in the face of emerging variants, incomplete vaccination coverage, breakthrough infections, and decreased reliance on other mitigation strategies (e.g., masking, remote learning).^{36,37} The lessons learned through this study may inform the design of future adaptive strategies, ideally building beyond symptom/exposure monitoring and gPCR testing to integrate complementary interventions such as rapid antigen self-testing and vaccination promotion.

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47	461
48	462
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0 Keywords: COVID-19, SARS-CoV-2, United States, young adults, students, universities, 1 essential workers, seroprevalence

-3 Acknowledgements

4 We are grateful for the contributions of an exceptional team of graduate student -5 researchers (Mariah De Zuzuarregui, Darren Frank, Sarah Gomez-Aladino, Ariel Muñoz, Ruben 6 Prado, Lawrence Tello, Emily Wang, and Sabrina Williamson) and our collaborators at UC -7 Berkeley's University Health Services (including but not limited to: Judith Sansone, Melody 8 Heller, Holly Stern, Tyler Crooks, Desi Gallardo, Jeff Kreutzen, Rebecca Stephenson, Lisa 9 Polley, and Melissa Hennings), the Innovative Genomics Institute (including but not limited to: 50 Fyodor Urnov, Shana McDevitt, Ariana Hirsch, Alexander Ehrenberg, and the other members of the IGI SARS-CoV-2 testing consortium: M Amen, Kerrie W Barry, John M Boyle, Cara E Brook, 51 52 Seunga Choo, L T Cornmesser, David J Dilworth, Jennifer A Doudna, Indro Fedrigo, Skyler E 53 Friedline, Thomas G W Graham, Ralph Green, Jennifer R Hamilton, Megan L Hochstrasser, 54 Dirk Hockemeyer, Netravathi Krishnappa, Azra Lari, Hangin Li, Enrique Lin-Shiao, Tianlin Lu, 55 Elijah F Lyons, Kevin G Mark, Lisa Argento Martell, A Raguel O Martins, Patrick S Mitchell, 56 Erica A Moehle, Christine Naca, Divya Nandakumar, Elizabeth O'Brien, Derek J Pappas, 57 Kathleen Pestal, Diana L Quach, Benjamin E Rubin, Rohan Sachdeva, Elizabeth C Stahl, 58 Abdullah Muhammad Syed, I-Li Tan, Amy L Tollner, Connor A Tsuchida, C Kimberly Tsui, 59 Timothy K Turkalo, M Bryan Warf, Oscar N Whitney, and Lea B Witkowsky), and Vitalant 60 Research Institute (including but not limited to: Mars Stone, Chloe Thorbrogger, Alice Lee, and 51 Heather Tanner). The author would also like to thank Drs. Sandra McCoy, Stefano Bertozzi, and 62 Lauren Ralph for their feedback on this manuscript.

63 Contributors: LH performed statistical analyses and wrote the first draft of the manuscript. SW performed phylogenomic analyses and prepared associated figures and paragraphs. AR and 64 6 MP designed the study and provided input on the manuscript. LP, SF, AH, GN, the IGI SARS-

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CoV-2 Testing Consortium, CDG, and MB provided feedback on the study design and
manuscript. YL assisted with data analyses. All authors hold final responsibility for the decision
to submit for publication.

469 Declaration of interests: Vitalant Research Institute, of which Dr. Michael Busch is Director,
470 receives research funding and free assay kits from Ortho Clinical Diagnostics. Dr. Busch does
471 not receive salary support or personal compensation from Ortho Clinical Diagnostics. The
472 remaining authors declare no competing interests.

Funding: The study was funded by private donors who had no role in study design, data

474 collection, data analysis, data interpretation, or writing of the report.

475 Data sharing: De-identified data sets used in analyses and accompanying R Markdown script
476 files will be publicly available at the time of publication at the following link:

477 https://github.com/lauren-hunter/bcsci

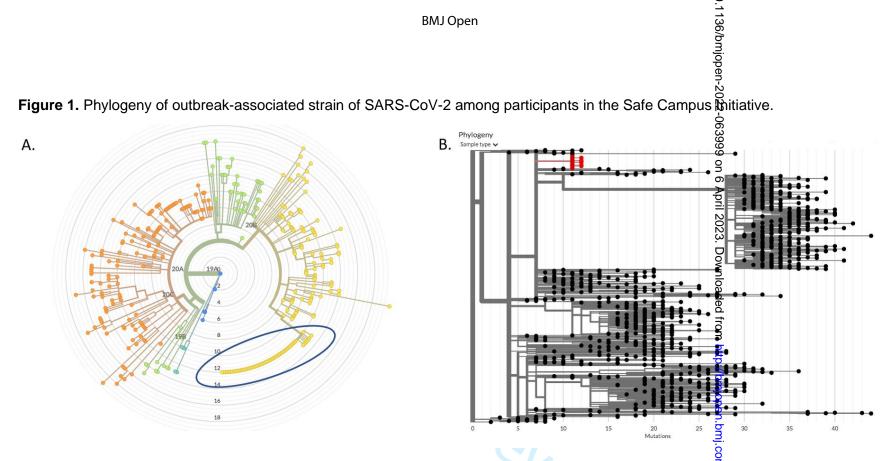
478 Ethical approvals: All study activities were approved by the University of California, Berkeley
 479 Committee for the Protection of Human Subjects (#2020-06-13349, #2020-05-13261, #2020-04 480 13238).

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A. A maximum likelihood phylogeny constructed from 357 genomes sequenced by the Innovative Genomics Institute between May and July 2020 constructed using Nextstrain. Branch lengths represent divergence from Wuhan reference genome at center. Blue circle marks cluster of identical genomes from a campus

super-spreader event. April 2021), showing the most similar genomes to the super-spreader event cluster (in red). There are no descendant branches from the cluster, demonstrating that the outbreak was contained and the lineage died out.

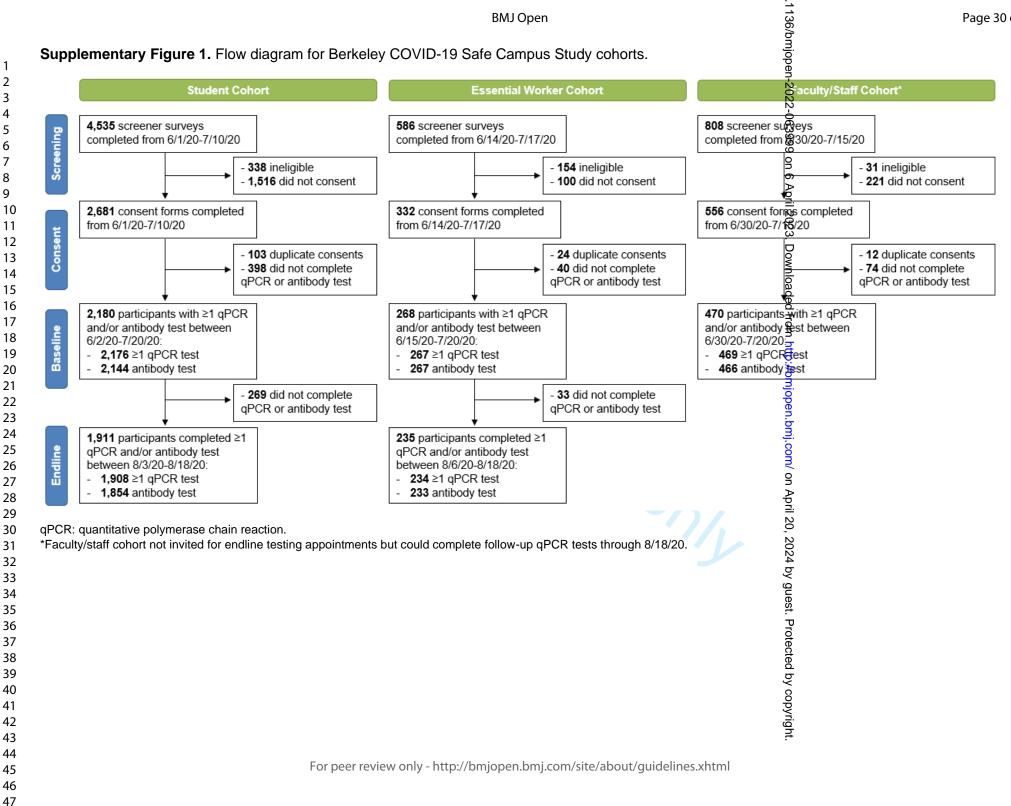
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Supplementary Table 1. Eligibility criteria across Berkeley COVID-19 Safe Campus Study cohorts.

	Student Cohort	Essential Worker Cohort	Faculty/Staff 2 ohort
	- At least 18 years of age	- At least 18 years of age	- At least 18 years of age
Eligibility Criteria	- Currently enrolled as an undergraduate or graduate student at UC Berkeley (i.e., not graduated in Spring 2020 or incoming for Fall 2020)	 Currently employed in one of the following departments at UC Berkeley: health services, police, facility services or other building management, environmental health and safety, laboratory animal care, athletics, dining, childcare, other residential or student services Currently working on campus at UC Berkeley <i>or</i> expected to return to work during June 2020 	- Currently employed a a faculty member, staff member, or postdoctor scholar at UC Berkeley - Not already enrolled ig the essential workers cohort
-	- Primarily residing in Alameda County or Contra Costa Country between 6/1/20-8/31/20	N/A	- Primarily residing in Astronomic Count or Contra Costa Country between 6/1/20-8/31/20
	- Willing to sign release of information for COVID-19-related medical records	- Willing to sign release of information for COVID-19-related medical records	- Willing to sign release of information
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31		BMJ Open	
		STROBE 2007 (v4) Statement—Checklist of items that should be included in reports of cohort studies	
Section/Topic	Item #	Recommendation 999	Reported on page
Title and abstract	1	(<i>a</i>) Indicate the study's design with a commonly used term in the title or the abstract 관	1-2
		(b) Provide in the abstract an informative and balanced summary of what was done and what was figund	2
Introduction		N	
Background/rationale	2	Explain the scientific background and rationale for the investigation being reported	4-5
Objectives	3	State specific objectives, including any prespecified hypotheses	5
Methods			
Study design	4	Present key elements of study design early in the paper	5
Setting	5	Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, foller -up, and data collection	5-9, Supplementa Figure 1
Participants	6	(a) Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up	5-9, Supplementa Table 1
		(b) For matched studies, give matching criteria and number of exposed and unexposed	N/A
Variables	7	Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable	6-10
Data sources/ measurement	8*	For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe	6-10
Bias	9		10
Study size	10	Explain how the study size was arrived at	6
Quantitative variables	11	Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why	10
Statistical methods	12	(a) Describe all statistical methods, including those used to control for confounding	10
		(a) Describe an statistical methods, including those used to control for comounding (b) Describe any methods used to examine subgroups and interactions (c) Explain how missing data were addressed	N/A
			N/A
		(d) If applicable, explain how loss to follow-up was addressed	N/A
		(e) Describe any sensitivity analyses	N/A

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13*	(a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examine for eligibility, confirmed	Supplementary
	eligible, included in the study, completing follow-up, and analysed	Figure 1
	(b) Give reasons for non-participation at each stage ගි දු	Supplementary Figure 1
	(c) Consider use of a flow diagram	Supplementary Figure 1
14*	(a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential	11, Table 1
	(b) Indicate number of participants with missing data for each variable of interest	11-13
	(c) Summarise follow-up time (eg, average and total amount)	11
15*	Report numbers of outcome events or summary measures over time	12
16	(<i>a</i>) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision egg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included	12-14, Tables 2-3
	(b) Report category boundaries when continuous variables were categorized	10, Table 3
	(c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period	N/A
17	Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses	N/A
	ý měl se	
18	Summarise key results with reference to study objectives	14-15
20	Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of a key lyses, results from similar studies, and other relevant evidence	14-17
21	Discuss the generalisability (external validity) of the study results	16
	gu du	
22	Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based	19
	14* 15* 16 17 18 20 21	eligible, included in the study, completing follow-up, and analysed Image: Completion of the study of the study of the study and information on exposures and potential confounders (c) Consider use of a flow diagram Image: Completion of the study participants (eg demographic, clinical, social) and information on exposures and potential confounders (b) Indicate number of participants with missing data for each variable of interest Image: Completion of the study and the present study and, if applicable, for the original study on

Monitoring SARS-CoV-2 incidence and seroconversion among university students and employees: a longitudinal cohort study in California, June to August 2020

Journal:	BMJ Open
Manuscript ID	bmjopen-2022-063999.R1
Article Type:	Original research
Date Submitted by the Author:	17-Feb-2023
Complete List of Authors:	Hunter, Lauren; University of California Berkeley, School of Public Health Wyman, Stacia; University of California Berkeley, Innovative Genomics Institute Packel, Laura; University of California Berkeley, School of Public Health Facente, Shelley; University of California Berkeley, School of Public Health; Facente Consulting, Li, Yi; University of California Berkeley, School of Public Health Harte, Anna; University of California Berkeley, University Health Services Nicolette, Guy; University of California Berkeley, University Health Services the IGI SARS-CoV-2 Testing Consortium, N/A; University of California Berkeley, Innovative Genomics Institute Di Germanio, Clara; Vitalant Research Institute Busch, Michael; Vitalant Research Institute; University of California San Francisco, Department of Laboratory Medicine Reingold, Art; University of California Berkeley, School of Public Health Petersen, Maya L.; University of California Berkeley, School of Public Health
Primary Subject Heading :	Epidemiology
Secondary Subject Heading:	Public health, Infectious diseases
Keywords:	COVID-19, EPIDEMIOLOGY, Public health < INFECTIOUS DISEASES, Infection control < INFECTIOUS DISEASES, Epidemiology < INFECTIOUS DISEASES, PUBLIC HEALTH

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1 2									
2 3 4	1	Title: Monitoring SARS-CoV-2 incidence and seroconversion among university students and							
5 6	2	employees: a longitudinal cohort study in California, June to August 2020							
7 8 9	3 4	Authors: Lauren A. Hunter, MPH ¹ ; Stacia Wyman, PhD, MS ² ; Laura Packel, PhD ¹ ; Shelley							
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 12 13 6 Testing Consortium²; Clara Di Germanio, PhD⁴; Michael P. Busch, MD PhD^{4,6}; Arth 									
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49 50	24	Word count: 4,213							
51 52	25	Abstract word count: 285							
53 54 55	26	Number of tables/figures: 5 (3 supplementary)							
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1 2		
2 3 4	27	Abstract
5 6 7 8	28	Objectives: To identify incident SARS-CoV-2 infections and inform effective mitigation
	29	strategies in university settings, we piloted an integrated symptom and exposure monitoring and
9 10	30	testing system among a cohort of university students and employees.
11 12 13 14	31	Design: Prospective cohort study.
	32	Setting: A public university in California from June to August 2020.
15 16	33	Participants: 2,180 university students and 738 university employees.
17 18 19	34	Primary outcome measures: At baseline and endline, we tested participants for active SARS-
20 21	35	CoV-2 infection via quantitative polymerase chain reaction (qPCR) test and collected blood
22 23	36	samples for antibody testing. Participants received notifications to complete additional qPCR
24 25 26 27	37	tests throughout the study if they reported symptoms or exposures in daily surveys or were
	38	selected for surveillance testing. Viral whole genome sequencing was performed on positive
28 29	39	qPCR samples, and phylogenetic trees were constructed with these genomes and external
30 31 32 33 34 35	40	genomes.
	41	Results: Over the study period, 57 students (2.6%) and 3 employees (0.4%) were diagnosed
	42	with SARS-CoV-2 infection via qPCR test. Phylogenetic analyses revealed that a super-
36 37	43	spreader event among undergraduates in congregate housing accounted for at least 48% of
38 39 40	44	cases but did not spread beyond campus. Test positivity was higher among participants who
40 41 42	45	self-reported symptoms (incidence rate ratio [IRR]: 12.7; 95% confidence interval [CI]: 7.4, 21.8)
43 44	46	or had household exposures (IRR: 10.3; 95% CI: 4.8, 22.0) that triggered notifications to test.
45 46	47	Most (91%) participants with newly identified antibodies at endline had been diagnosed with
47 48	48	incident infection via qPCR test during the study.
49 50	49	Conclusions: Our findings suggest that integrated monitoring systems can successfully identify
51 52	50	and link at-risk students to SARS-CoV-2 testing. As the study took place before the evolution of
53 54	51	highly transmissible variants and widespread availability of vaccines and rapid antigen tests,
55 56	52	further research is necessary to adapt and evaluate similar systems in the present context.
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2		
3 4	53	Strengths and limitations of this study
5 6	54	• The study is strengthened by rich longitudinal data including more than 117,000 daily
7 8	55	symptom surveys; 17,000 weekly exposure surveys; 7,600 qPCR tests to detect active
9 10	56	SARS-CoV-2 infection; and 4,900 antibody tests to detect previous infection collected
11 12	57	from 2,918 university students and employees over three months.
13 14	58	We used seroconversion data from serial antibody tests and phylogenetic analyses
15 16	59	comparing viral genome sequences to a broader database to evaluate the extent to
17 18 19	60	which the study system identified incident cases and contained an outbreak among
20 21	61	university students.
22 23	62	Our identification of participants who seroconverted between baseline and endline may
24 25	63	be incomplete due to loss-to-follow up and imperfect sensitivity of SARS-CoV-2 antibody
26 27	64	testing.
28 29	65	 A high proportion of identified cases were traced to one outbreak, limiting the
30 31	66	generalizability of our exploratory assessment of risk factors for incident infection.
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<u> </u>		
3 1	67	Background
5	68	Universities have been identified as hotspots for SARS-CoV-2 transmission in the United
7 3	69	States,[1] where SARS-CoV-2 incidence is highest among young adults.[2] Young adults may
) 10	70	be less likely to adhere to social distancing guidelines and more likely to experience workplace
1 2	71	exposure (for example, at food service or retail jobs).[2] Their risk may be heightened in
3 4	72	university settings where many live in congregate housing, interact with wide social networks, or
15 16	73	attend large gatherings.[3] Although young adults are at low risk of serious acute illness or

74 death from COVID-19 (the disease caused by SARS-CoV-2),[4] the higher likelihood of

75 asymptomatic or mildly symptomatic infection in this age group makes young adults a key

76 population through which SARS-CoV-2 may spread to other, more vulnerable groups.[2,5]

78 COVID-19-related mortality in the surrounding counties.[6–8] Although widespread vaccination

Indeed, there is evidence that transmission among university students may lead to increased

79 has enabled campuses to return to in-person activities, the elimination of SARS-CoV-2

80 transmission in campus populations may be stymied by vaccine hesitancy among students and

81 employees and breakthrough infection and subsequent transmission by vaccinated persons,

particularly in the context of waning immunity and viral variants which reduce vaccine
efficacy.[9,10] Therefore, rapid and resource-efficient identification of incident cases in
university populations is a critical first step of outbreak investigation and control, followed by
isolation, case investigation, and contact tracing, to minimize transmission within campus and to

86 the broader community.

Universities have adopted a wide range of approaches for testing and outbreak
mitigation.[11–13] While a number of well-resourced universities have scaled up testing capacity
in order to frequently test all students and employees accessing campus or living in universityaffiliated housing,[13] many other universities do not have well-defined testing strategies or
restrict testing to those with symptoms or known exposure.[12] Beyond investing in testing
programs, some universities have sought to reduce on-campus transmission by mandating the

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completion of self-administered symptom screening tools by students and employees. However, such tools have primarily been used to regulate daily access to campus (i.e., deny entry to those who report COVID-19-like symptoms), rather than to detect emergent outbreaks among university populations. As universities resume normal operations and discontinue mitigation strategies such as masking, non-punitive, resource-efficient strategies which can both identify those who are at highest risk of infection and expediently link them to low-barrier testing services may play a key role in transitioning from a "one-size-fits-all" approach of uniform testing to a sustainable monitoring paradigm.

In 2020, we piloted an integrated symptom and exposure monitoring and testing system designed to identify incident SARS-CoV-2 infections among a cohort of university students and employees.[14] Here we describe the incidence and seroprevalence of SARS-CoV-2 infection within this cohort to evaluate the extent to which incident infections were successfully detected and contained over the study period, identify sociodemographic factors associated with incident infection, and ascertain which self-reported symptoms and exposures tracked by the monitoring system were predictive of test positivity, with the ultimate objective of informing monitoring and testing strategies in university settings.

9 110 Methods

111 Study design and setting

112 The study comprised three prospective cohorts of University of California, Berkeley
113 affiliates followed from June to August 2020: students, essential workers (i.e., employees
114 working on campus in health, facilities, or student services), and other employees (hereafter,
115 "faculty/staff"). We report the findings according to the Strengthening the Reporting of
116 Observational Studies in Epidemiology (STROBE) checklist for cohort studies.[15]
117 Throughout the study period, public health orders mandated the use of face coverings in
118 public and upheld many restrictions set forth by earlier shelter-in-place orders, while allowing

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3 4 5 6 7 8 9 10	119	phased reopening of certain businesses and activities.[16] UC Berkeley did not offer in-person	I
	120	classes, and on-campus work was restricted to essential workers and a small subset of faculty	/,
	121	staff, and student researchers. Although few students were living in on-campus residence halls	S,
	122	many students continued to live in congregate living settings off campus, such as fraternities,	
11 12	123	sororities, and co-operative housing. From June to August 2020, daily case counts in Alameda	a
13 14	124	County ranged from approximately 50 to 350 (0 to 17 within the city of Berkeley).[17]	
15 16	125		
17 18	126	Participant recruitment and eligibility	
19 20 21	127	The study was promoted through targeted messages from university officials to campu	S
21 22 23	128	email listservs and social media platforms from early June to mid-July 2020. To increase reach	h
23 24 25	129	to students expected to be at higher risk of COVID-19, we also placed flyers in congregate livit	ng
26 27	130	settings and conducted in-person recruitment for student athletes who had resumed training o	n
28 29 30 31 32 33 34 35 36 37 38 39	131	campus. Participants were eligible to enroll in the study if they were at least 18 years of age,	
	132	were a current student or employee at UC Berkeley, and planned to live in or near Berkeley	
	133	during the summer of 2020. Specific eligibility criteria and enrollment windows varied by cohor	t
	134	(Supplementary Table 1, Supplementary Figure 1).	
	135	Upon enrollment, participants were linked to an online baseline survey that collected	
	136	sociodemographic data and information about their COVID-19-related health history.	
40 41	137	Participants were then referred to a baseline testing appointment at University Health Services	5
42 43 44	138	(UHS) which included a SARS-CoV-2 quantitative polymerase chain reaction (qPCR) test and	
45 46	139	blood collection for antibody testing (procedures described below). To facilitate daily	
47 48	140	temperature monitoring, study staff also provided participants with free oral thermometers upo	n
49 50	141	request at testing appointments. Participants who completed this appointment or a non-study	
51 52	142	qPCR test at UHS by July 20 were eligible to remain in the study. We pre-specified a maximur	n
53 54	143	sample size of 4,000 participants across cohorts but did not reach this limit before the final day	y
55 56	144	of baseline data collection.	
57 58			6

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2 3 4 5 6 7 8 9 10 11 12 13 14	145				
	146	Symptom and exposure surveys			
	147	Participants received daily text messages or emails, depending on their preference			
	148	specified in the baseline survey, which linked to short symptom surveys through which they			
	149	reported their body temperature and any symptoms of illness. Once per week, the daily survey			
	150	included a longer exposure module, which asked about recent symptoms of illness among their			
15 16	151	household member(s), potential exposure(s) to COVID-19, and activities related to potential			
17 18	152	COVID-19 risk. All surveys were administered via REDCap.[18,19]			
19 20	153				
21 22	154	Endline survey and testing			
23 24 25	155	In early August, participants were sent an endline survey which collected updated			
25 26 27	156	information on their COVID-19 history to identify any diagnoses outside of the study.			
27 28 29 30 31	157	Participants in the student and essential worker cohorts were also invited to complete endline			
	158	testing appointments by August 18, including a final qPCR test and blood collection.			
32 33	159				
34 35	160	qPCR testing			
36 37	161	Midturbinate nasal and oral swabs were collected by UHS clinical staff and tested for			
38 39	162	SARS-CoV-2 by qPCR at the Innovative Genomics Institute (IGI).[20] qPCR tests were			
40 41	163	performed at baseline for all three cohorts and at endline for the student and essential worker			
42 43	164	cohorts. Between baseline and endline testing, additional qPCR tests were performed for the			
44 45	165	following reasons:			
46 47 48	166	 Symptom- or exposure-based tests triggered based on participants' responses in 			
48 49 50	167	daily surveys: Participants who reported COVID-19-like signs or symptoms ¹ (in			
50 51 52					
53 54					
55 56		¹ Signs or symptoms which triggered a testing notification when reported were: temperature of ≥100.4°F, dry cough (without mucus), coughing up mucus, feeling feverish, unusual pain or pressure in the chest, difficulty breathing, shortness of breath, unexplained trouble thinking or concentrating, loss of sense of taste, or loss of sense of smell.			
57		הוסינוובים סו שרבמוו, מוופגעומוופט נוסטשופ מוווזגוווץ סו כסווכפותמוווץ, וסכי סו כפווכפ סו נמצופ, סו וסכי סו כפווכפ סו			

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1		
2 3 4	168	themselves or household member(s)) or who reported a suspected or confirmed COVID-
5 6	169	19 case in their household were automatically notified to sign up for a qPCR test.
7 8	170	• Random surveillance testing: A subset of participants in the student and faculty/staff
9 10	171	cohorts who had not had a qPCR test within a week were randomly selected and
11 12 13 14	172	emailed notifications to come in for surveillance testing in July.
	173	Address-based surveillance testing: Participants who lived at the same address as
15 16	174	another participant who tested positive for SARS-CoV-2 were immediately emailed
17 18 19	175	surveillance testing notifications. Following an outbreak among group-housed students
20 21	176	in early July, surveillance testing notifications were also emailed to all participants who
22 23	177	had not been tested within the week and who reported living in fraternities, sororities, or
24 25	178	co-operative housing.
26 27	179	Participant-initiated testing: Participants could self-schedule study testing
28 29 30 31 32 33	180	appointments on demand, with or without consulting a healthcare provider and
	181	regardless of exposure history.
	182	Participants with positive qPCR test results were informed by phone by UHS clinical staff, who
34 35	183	provided guidance on isolation and performed case investigation to identify potential contacts.
36 37 38	184	Participants with negative qPCR test results were informed of their results via the UHS online
39 40	185	patient portal.
41 42	186	
43 44	187	SARS-CoV-2 sequencing and phylogenetic analyses
45 46	188	Viral whole genome sequencing was performed on a set of positive samples at the IGI,
47 48	189	using previously described procedures.[21] Briefly, SARS-CoV-2 RNA extracted from swabs
49 50	190	was reverse transcribed using SuperScript IV (Invitrogen), and the viral genome was amplified
51 52	191	from the resulting cDNA in four separate qPCR reactions using distinct primer sets tiling the
53 54	192	SARS-CoV-2 genome. The four qPCR reactions were pooled 1:1:1:1 and diluted 1:50 in H_2O . A
55 56 57	193	second qPCR reaction was set up to add Nextera Unique Dual Indexing (UDI) sequences to
57 58 59		8
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

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3 4	194	either end of the amplicons. The resulting qPCR reaction was cleaned up using 0.7x AMPure>	KΡ
5 6	195	beads (Beckman Coulter) and quantified using a Qubit dsDNA HS Assay Kit (Thermo Fisher).	
7 8	196	The libraries were then pooled to an equimolar ratio and sequenced with a 10% PhiX spike in	
9 10	197	using a MiSeq v3 kit at 300bp PE reads.	
11 12	198	Fastq sequencing files were processed through a custom pipeline using publicly	
13 14	199	available software. The reads were preprocessed by quality trimming, removing adaptors, and	ł
15 16	200	PhiX cleaning with BBTools,[22] and then aligned to the Wuhan reference sequence	
17 18 19	201	(NC_045512.2) with minimap2 v2.16-r922. ARTICv3 primers were trimmed, and the consensu	JS
20 21	202	sequence was built with iVar v1.3.1, where an 'N' is called if the depth is less than 10 reads at	i
22 23	203	any nucleotide. The genomes were then processed through the Nextstrain Auger pipeline with	ı
24 25	204	other genomes from GISAID to construct a maximum likelihood tree.[23,24] Several	
26 27	205	phylogenies were constructed for this analysis: a tree of 7,091 genomes subsampled from the	;
28 29	206	worldwide genomes in GISAID at the time (approximately 200,000 genomes as of October	
30 31	207	2020) was used to place the IGI genomes in the larger tree; a tree with all IGI genomes	
32 33	208	sequenced at the time of analysis (356 genomes); and a tree containing 500 genomes (from 1	l
34 35	209	million genomes as of April 2021) was constructed using UShER.[25]	
36 37	210		
38 39 40	211	Antibody testing	
40 41 42	212	Up to 10 mL of blood was collected by phlebotomists via venipuncture at baseline from	ı
43 44	213	participants in all three cohorts and again at endline from participants in the student and	
45 46	214	essential worker cohorts. Blood was centrifuged and serum was stored at -20°C for 2 to 4	
47 48	215	months before being tested at Vitalant Research Institute using the VITROS Immunodiagnosti	iC
49 50	216	Products Anti-SARS-CoV-2 Total Reagent Pack, which detects IgA, IgG, and IgM antibodies	
51 52	217	against the SARS-CoV-2 spike protein S1 antigen and has an estimated clinical specificity of	
53 54	218	100% and unreported sensitivity.[26]	
55 56	219		
57 58 59			9
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2		
3 4	220	Participant compensation
5 6	221	Participants in the student cohort received a \$50 gift card after completing baseline
7 8	222	testing and 10 daily surveys; this incentive was conditional on daily survey completion to
9 10	223	encourage early habit formation.[27] Student participants received a second \$50 gift card at
11 12	224	their endline testing appointment. To facilitate travel to and from UHS for testing appointments,
13 14	225	student participants were also offered pre-paid car rides via a ride-sharing app.
15 16	226	Participants in the essential worker cohort received a gift card worth \$1 per daily survey
17 18 19	227	completed (to a maximum of \$70) after the study ended. Participants in the faculty/staff cohort
20 21	228	were not compensated.
22 23	229	
24 25	230	Statistical analyses
26 27	231	To identify sociodemographic factors associated with incident infection, we used Poisson
28 29	232	regression to estimate unadjusted incidence rate ratios (IRRs) for SARS-CoV-2 infection by
30 31	233	study cohort and within strata of sociodemographic variables self-reported in the baseline
32 33	234	survey (e.g., age, gender, housing type), setting person-months of enrollment as an offset term
34 35	235	to account for differing lengths of follow-up.
36 37	236	We also calculated IRRs comparing test positivity by recent signs/symptoms, exposures,
38 39 40	237	and activities reported in the daily and weekly surveys. We estimated IRRs for several
40 41 42	238	temperature thresholds (i.e., ≥100.4°F, ≥100.0°F, ≥99.0°F) to compare to symptom-specific
43 44	239	IRRs; however, continuous associations between temperature and positivity have been
45 46	240	previously explored in this cohort, finding that temperature screening has low sensitivity to
47 48	241	SARS-CoV-2 infection and, thus, limited efficacy as a primary means of detection.[28] While it
49 50	242	was not possible to isolate participants' specific reason(s) for testing over the study period (e.g.,
51 52	243	participants could receive symptom- and/or exposure-triggered testing notifications over the
53 54	244	same time window in which they completed baseline or endline testing), we linked qPCR test
55 56	245	results to recently-completed symptom and exposure surveys to identify testing appointments
57 58		10
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2					
2 3 4	246	that took place in the days or weeks following symptom- and exposure-triggered testing			
5 6 7 8	247	notifications (Supplementary Figure 2). We accounted for clustered observations due to			
	248	repeated tests per participant using a generalized estimating equation approach with Huber-			
9 10	249	White standard error estimates and an exchangeable working correlation structure.[29]			
11 12	250	Finally, to assess the extent to which the testing and monitoring system captured			
13 14	251	incident infections, we identified participants who seroconverted from having non-reactive (no			
15 16	252	antibodies detected) to reactive (antibodies detected) blood samples between baseline and			
17 18	253	endline and calculated the proportion of these participants who were also diagnosed with			
19 20 21	254	incident SARS-CoV-2 infection via positive qPCR test during the study period. Analyses were			
21 22 23	255	conducted in R version 4.2.1.[30]			
24 25	256				
26 27	257	Ethical approvals			
28 29	258	All study activities were approved by the University of California, Berkeley Committee for			
30 31	259	the Protection of Human Subjects (#2020-06-13349, #2020-05-13261, #2020-04-13238).			
32 33	260				
34 35	261	Patient and public involvement			
36 37	262	The study's target population comprised university students and employees. While the			
38 39 263 stud 40		study was conducted by faculty, staff, and graduate students from the UC Berkeley School of			
41 42	264	Public Health, University Health Services, and the Innovative Genomics Institute, the broader			
43 44	265	student body and university workforce were not involved in designing the study or selecting the			
45 46	266	research question, outcome measures, or method of disseminating results.			
47 48	267				
49 50	268	Results			
51 52	269	Participant recruitment and retention			
53 54	270	Between June 1 and July 20, 2020, we enrolled 2,180 students, 268 essential workers,			
55 56 57	271	and 470 faculty/staff who completed at least one qPCR test or antibody test (Table 1,			
57 58 59		11			
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1 2								
2 3 4	272	Supplementary Figure 1). The student cohort was split between undergraduate (52%) and						
5 6	273	graduate (48%) students. Nearly ha	lf (44%) of esse	ential workers v	vorked in healt	h services.		
7 8	274	While 85% of essential workers wer	e working on ca	ampus at the tir	me of enrollme	ent, most (81%)		
9 10	275	faculty/staff were working entirely re	motely. At the t	time of enrollm	ent, only 12 (0.	.4%)		
11 12	276	participants reported a previous COVID-19 diagnosis.						
13 14	277	Participants provided a total of 5,545 person-months of follow-up from enrollment to the						
15 16	278	end of the study (mean person-days per participant: 57, range: 32-78). Participants completed a						
17 18	279	mean of 40 daily symptom surveys and 6 weekly exposure surveys over the study period, for a						
19 20 21	280	total of 117,239 symptom and 17,16	2 exposure sur	veys. A subset	t of participants	s did not		
21 22 23	281	complete any daily symptom survey	rs (1.7%) or wee	ekly exposure s	surveys (4.2%)).		
24 25 26	282 283	Table 1. Baseline characteristics of Initiative by study cohort, June-Augu	the Berkeley C	OVID-19 Safe	Campus			
27 28 29			All	Students	Essential Workers	Faculty/Staff		
30		N (row %)	2,918 (100)	2,180 (74.7)	268 (9.2)	470 (16.1)		
31		Age, mean ± SD	29.4 ± 11.6	24.3 ± 5.4	42.5 ± 12.3	45.2 ± 12.3		
32 33 34 35 36		Gender, n (column %) Man Woman Non-binary/other	1,177 (40.3) 1,653 (56.6) 51 (1.7)	911 (41.8) 1,187 (54.4) 46 (2.1)	103 (38.4) 164 (61.2) 1 (0.4)	163 (34.7) 302 (64.3) 4 (0.9)		
37 38 39 40 41 42 43 44		Race/ethnicity, n (column %)* American Indian/Alaska Native Asian/Pacific Islander Black/African American Hispanic/Latine/Spanish origin White Other	39 (1.3) 833 (28.5) 103 (3.5) 420 (14.4) 1,814 (62.2) 280 (9.6)	Ć		8 (1.7) 64 (13.6) 4 (0.9) 35 (7.4) 393 (83.6) 26 (5.5)		
45 46 47		Program level, n (column %) Undergraduate Graduate	-	1,114 (51.7) 1,039 (48.2)	-	-		
48 49 50		Living at fraternity/sorority, n (column %)	-	125 (5.7%)	-	-		
51 52 53 54 55 56 57 58		Education, n (column %) High school diploma/GED Some college or trade school Bachelor's degree Graduate/professional degree	- - -	- - -	6 (2.2) 59 (22.0) 78 (29.1) 121 (45.1)	0 (0) 13 (2.8) 119 (25.3) 337 (71.7)		
59						. 12		

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	- - -	- -	129 (48.1) 61 (22.8) 77 (28.7)	- - -			
	- - -	- -	- -	110 (23.4) 311 (66.2) 49 (10.4)			
е	748 (25.6)	418 (19.2)	228 (85.1)	102 (21.7)			
	12 (0.4)	8 (0.4)	1 (0.4)	3 (0.6)			
d, part	icipants underwe	ent 7,638 qPC	CR tests for ac	tive SARS-CoV-			
6 tests	per participant (range: 0-9). A	lmost all (99.9	%) participants			
test. C	Overall, 60 partic	ipants (2.0%)	tested positive	e: 57 students, 2			
lty/staf	f.						
nts were at highest risk of incident infection over the study period							
: 5.8; 95% confidence interval [CI]: 1.3, 103.0). Due to the low							
e stud	ent cohort, we e	xamined addi	tional risk facte	ors for infections			
2), findi	ng higher rates o	of infection an	nong students	who were 18-19			
22 yea	rs: 8.3; 95% CI:	4.2, 17.5) and	l undergradua	tes (IRR vs.			
CI: 2.2,	8.7). We also o	bserved a hig	her incidence	among white			
ite stu	dents; 95% CI: 1	.5, 5.5). Thes	e associations	s were largely			
partici	pants living in fra	aternities or so	ororities. Near	ly one-quarter of			
es or so	prorities were inf	ected with SA	.RS-CoV-2 du	ring the study			
s: 20.9;	; 95% CI: 12.3, 3	35.5), and the	se participants	accounted for			
ng stud	g student participants.						
	-						
				13			

	Health services Facilities/building services Student services/other	- -	
	Job title, n (column %) Faculty Staff Postdoctoral scholar/other	- -	
	Currently working outside the home, n (column %)	748 (25.6)	418 (′
	Pre-enrollment COVID-19 diagnosis, n (column %)	12 (0.4)	8
284 285	*Categories not mutually exclusive.		
286	SARS-CoV-2 incidence		
287	During the study period, part	icipants underw	ent 7,63
288	2 infection, with a mean of 2.6 tests	per participant (range: 0
289	completed at least one qPCR test. C	overall, 60 partic	ipants (2
290	essential workers, and 1 faculty/staf		
291	Among cohorts, students we	re at highest risl	k of incid
292	(IRR students vs. faculty/staff: 5.8; 9	5% confidence	interval

Department, n (column %)

ent infection over the study period CI]: 1.3, 103.0). Due to the low number of cases outside of the additional risk factors for infections among students only (Table 2) on among students who were 18-19 years old (IRR vs. students ≥2 and undergraduates (IRR vs. graduate students: 4.1; 95% C a higher incidence among white students (IRR: 2.8 vs. non-whi These associations were largely driven by an outbreak among or sororities. Nearly one-quarter of participants living in fraternities th SARS-CoV-2 during the study period (IRR vs. other students d these participants accounted for 49% of cases observed among

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Table 2. Bivariate associations between sociodemographic characteristics and SARS-CoV-2
 incidence among student participants in the Safe Campus Initiative, June-August 2020.

	Cases, N (row %)	Non-Cases, N (row %)	IRR (95% CI)
Overall*	57 (2.6)	2,120 (97.4)	
Age 18-19 years 20-21 years ≥22 years	21 (8.0) 24 (3.8) 12 (0.9)	243 (92.0) 607 (96.2) 1,270 (99.1)	8.3 (4.2, 17.5 4.2 (2.1, 8.6 Reference
Gender Woman Man Non-binary/other	37 (3.1) 19 (2.1) 0 (0)	1,147 (96.9) 892 (97.9) 46 (100)	1.5 (0.9, 2.6 Reference
Race/ethnicity** American Indian/Alaska Native Asian/Pacific Islander Black/African American Hispanic/Latine/Spanish origin White Other	0 (0) 11 (1.6) 1 (1.2) 8 (2.3) 45 (3.6) 4 (1.8)		0.5 (0.2, 0.9 0.5 (0.03, 2.0 0.9 (0.4, 1.8 2.8 (1.5, 5.5 0.7 (0.2, 1.6
Program level Undergraduate Graduate	46 (4.1) 10 (1.0)	1,027 (99.0)	4.1 (2.2, 8.7 Reference
Living at fraternity/sorority Currently working outside the home	28 (22.4) 6 (1.4)	97 (77.6) 410 (98.6)	20.9 (12.3, 35.5 0.5 (0.2, 1.1

305 *N=2,177 students with at least one qPCR test for SARS-CoV-2 during the study period.

306 **Not mutually exclusive; all participants not included in specified racial/ethnic category served as reference for each comparison.

⁴⁰ 309 Phylogenetic analysis

⁴² 310 We retrieved whole viral genome sequences for 35 of the 60 positive cases from this

⁴⁴ 311 study, 29 (83%) of which were found to be part of a campus super-spreader event involving a

⁴⁶ 312 total of 57 campus-affiliated individuals with samples sequenced by IGI (Figure 1A). Most (69%)

 $_{49}^{48}$ 313 study participants within this cluster lived at one of two residences, with likely a single

51 314 participant originating the super-spreader event. The cluster of genomes was defined by three

53 315 mutations (A6360G, C24502A and G110083T), two of which were extremely rare at the time of

55 316 the outbreak. The combination of the three variants was only found in four genomes outside of

this cluster (two in the UK and two in Florida) by October 2020, making it a strong phylogenetic signature. Phylogenetic analysis demonstrated that the cluster remained confined to campus, as this signature was not observed in any genomes from samples in the surrounding communities or California state in the months following the super-spreader event. When the trio of mutations was searched in a phylogeny constructed from over 1.2 million genomes worldwide using UShER in April 2021, [25] no descendent leaves were found in the tree under the cluster (Figure 1B), indicating that the lineage died out after the super-spreader event. Factors associated with test positivity At least one symptom survey was completed in the 7 days before sample collection for 88% of tests (n=6,668), including 72% of tests (n=5,465) that had symptom data from the day of sample collection. Of the 52 cases who completed at least one survey during the week before their positive sample was collected (mean: 4 surveys), 23 cases (44%) had reported at least one of the nine COVID-19 symptoms that triggered a notification for them to test. Test positivity was 12.7 times higher among participants who had a recent symptom-triggered notification (95% CI: 7.4, 21.8) (Table 3). Notification-triggering symptoms most strongly associated with test positivity included loss of sense of taste or smell and feeling feverish. Weakness, sweats or chills, and swollen glands were the non-triggering symptoms most strongly associated with test positivity.

-		
3	337	Table 3. Bivariate associations between prospectively monitored symptoms and exposures and
1 -	338	SARS-CoV-2 qPCR test positivity among participants in the Safe Campus Initiative, June-

August 2020.

0			
7 8		Test Positivity, % (+ Tests / All Tests)	IRR (95% CI)
9 10	Overall*	0.8 (60 / 7,615)	
11		0.0 (0077,010)	-
12	Signs/symptoms within 7 days of test		
13	No	0.3 (18 / 5,489)	Reference**
14	Yes (any)	2.9 (34 / 1,179)	8.8 (5.0, 15.5)
15	- Temperature ≥100.4°F [†]	0.0 (0 / 10)	0.0 (0.0, 0.0)
16	 Temperature ≥100.0°F 	10.5 (2 / 19)	13.2 (3.4, 50.9)
17	 Temperature ≥99.0°F 	2.9 (12 / 417)	4.3 (2.2, 8.2)
18	 Feeling feverish † 	15.3 (11 / 72)	24.6 (13.2, 45.8)
19	 Dry cough † 	5.6 (7 / 126)	8.1 (3.7, 17.7)
20	 Coughing up mucus [†] 	5.5 (5 / 91)	7.7 (3.1, 19.0)
21	 Unusual chest pain or pressure [†] 	9.7 (6 / 62)	13.9 (6.1, 31.6)
22	 Difficulty breathing † 	5.6 (1 / 18)	7.2 (1.0, 49.8)
23	 Shortness of breath † 	8.9 (4 / 45)	12.3 (4.6, 32.9)
24	 Trouble thinking/concentrating [†] 	7.6 (5 / 66)	10.7 (4.4, 26.1)
25	- Loss of sense of taste †	42.9 (3 / 7)	58.3 (23.7, 143)
26	 Loss of sense of smell [†] 	33.3 (4 / 12)	46.2 (19.2, 111)
27	 Any notification-triggering symptom [†] 	5.9 (23 / 393)	12.7 (7.4, 21.8)
28	- Loss of appetite	10.3 (6 / 58)	14.8 (6.5, 33.9)
29	- Fatigue	3.5 (13 / 371)	5.4 (3.0, 10.6)
30	- Trouble sleeping	5.1 (7 / 136)	7.5 (3.4, 16.4)
31	- Headache	4.7 (14 / 300)	7.8 (4.3, 14.3)
32	- Runny, blocked, or painful sinuses	5.2 (14 / 267)	8.8 (4.8, 16.2)
33	- Sneezing	1.9 (2 / 106)	2.5 (0.6, 10.1)
34	- Swollen, red, or painful eyes	8.6 (5 / 58)	12.1 (4.9, 30.0)
35	- Sore throat	3.1 (8 / 258)	4.5 (2.1, 9.5)
36	- Stomach pain	5.8 (5 / 86)	8.1 (3.3, 20.1)
37	- Diarrhea	4.9 (4 / 82)	6.7 (2.5, 18.2)
38		3.3 (3 / 90)	4.5 (1.4, 14.2)
39	- Nausea or vomiting		
40	- Body aches or muscle pain	8.2 (12 / 146)	13.4 (7.2, 25.2)
41 42	- Sweats or chills	11.5 (10 / 87)	18.0 (9.2, 35.2)
42	- Swollen glands	12.2 (5 / 41)	17.3 (7.2, 41.2)
43	- Weakness	13.5 (10 / 74)	21.2 (11.0, 40.9)
44 45	Exposures within 14 days before test		
45 46	No	0.3 (15 / 4,319)	Reference**
40 47	Yes (any)	3.4 (17 / 506)	9.6 (4.8, 19.2)
47	- Suspected or confirmed COVID-19	7.4 (7 / 95)	13.9 (6.1, 31.8)
49	case in household ⁺		
50	 Close contact with suspected or 	3.5 (5 / 144)	6.0 (2.3, 15.4)
51	confirmed case outside household		
52	- Household member with new	4.4 (5 / 114)	7.6 (3.0, 19.6)
53	COVID-19-like symptoms †		(0.0, 0.0)
54	- Household member with any new	2.6 (9 / 347)	5.0 (2.3, 10.8)
55	symptoms of illness	2.0 (0, 011)	0.0 (2.0, 10.0)
56			

1 2							
3		 Any notification-triggering exposure [†] 	5.1 (9 / 177)	10.3 (4.8, 22.09)			
4 5		Activities within 14 days before test					
6		No	0.4 (3 / 678)	Reference**			
7		Yes (any)	0.7 (29 / 4,145)	1.6 (0.5, 5.1)			
8		- Spent time at another residence	1.1 (26 / 2,327)	4.6 (1.9, 11.3)			
9		- Had visitors at own residence	1.0 (22 / 2,205)	2.6 (1.2, 5.5)			
10 11		 Attended gathering >10 people Worked outside of home 	2.8 (19 / 672) 0.5 (10 / 2,152)	9.0 (4.5, 18.1) 0.6 (0.3, 1.2)			
12		- Used public restroom	0.7 (12 / 1,821)	1.0 (0.5, 2.0)			
13		- Used public transportation	0.6 (4 / 699)	0.8 (0.3, 2.4)			
14		 Participated in group sports 	1.6 (4 / 257)	2.5 (0.9, 7.2)			
15	340	qPCR: quantitative polymerase chain reaction, IRR: incidence ra	te ratio. CI: confidence inter	val.			
16 17	341	*Excluding resamples, same-day re-tests, and repeated positives					
18	342 343	qPCR test for SARS-CoV-2 during the study period.	· · · · · · · · · · · · · · · · · · ·				
19	343 344	**Reference group for "Yes (any)" comparisons; reference group those who did not report that symptom/exposure/activity.	s for specific symptoms/exp	osures/activities were			
20	345	[†] Reporting triggered notification to test.					
21 22	346						
22							
24	347	Participants completed at least one weekly ex	posure survey in the 1	4 days before			
25	348	sample collection for 63% of tests ($n=4,825$). Of the 3	32 cases who had recei	ntly completed an			
26 27	010						
27 28	349	exposure survey at the time of sample collection, 9 (2	29%) reported a potenti	al household			
29							
30 31 32 33 34	350	exposure that triggered a notification for them to test	(Table 3). Test positivit	y was 10.3 times			
	351	higher among participants who had a recent exposur	e-triggered notification	(95% CI: 4.8, 22.0).			
	352	Test positivity was also significantly higher among pa	urticipants who reported	recent engagement			
35		in 'higher risk' social activities, most notably attending a gathering of more than 10 people (IRR:					
36 37	353	in 'higher risk' social activities, most notably attending		nan 10 people (IRR:			
38 39	354	9.0; 95% CI: 4.5, 18.1).					
40	355						
41 42							
43	356	SARS-CoV-2 seroprevalence					
44 45	357	Only 18 (0.6%) of 2,877 participants who prov	vided blood samples at	baseline had			
46 47	358	SARS-CoV-2 antibodies (Table 4), all but one of them students. Most participants with					
48	050						
49 50	359	antibodies at baseline either suspected past infection	i (28%), had been prev	lously diagnosed			
51 52	360	(22%), or had a positive qPCR test the day blood was drawn (11%). Most (85%) participants in					
53	361	the student and essential worker cohorts provided blood samples at both baseline and endline					
54 55	362	(mean interval between samples: 48 days). Among 2,076 participants with baseline and endline					
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2 3	363	blood samples, 33 (1.6%) seroconverted from non-reactive at baseline to reactive at endline, 30							
4 5 6	364	of whom (91%) were also diagnosed via qPCR test during the study. Of the three participants							
7 8	365	who seroconverted without a positive qPCR test, two self-reported suspected past infection (one							
9 10	366	before baseline, one during the study per	riod), while the third	did not suspect pa	st infection and				
11 12	367	had four negative qPCR tests over 40 days of study participation.							
13 14	368	Of the 60 participants with incident SARS-CoV-2 infection during the study period, 41							
15 16	369	(68%) provided an endline blood sample	at least one week a	fter the date of the	ir first positive				
17 18	370	qPCR test (mean time between positive of	qPCR test and blood	d sample: 36 days;	range 13-52				
19 20	371	days). Of these, 34 (83%) were reactive ((Table 4).						
21 22 23	372								
24 25 26	373 374	Table 4. Seroprevalence of SARS-CoV-2Initiative, June-August 2020.	2 antibodies among	participants in the	Safe Campus				
27 28			Baseline, N (%)	Endline, N (%)	Both, N (%)				
29 30 31 32		Serostatus – Cross-sectional* Reactive Non-reactive	18 (0.6) 2,859 (99.4)	48 (2.3) 2,039 (97.7)	-				
33 34 35 36 37		Serostatus – Longitudinal** Non-Reactive → Non-Reactive Non-Reactive → Reactive Reactive → Non-Reactive Reactive → Reactive	-,	,, (c, , 	2,029 (97.7) 33 (1.6) 0 (0) 14 (0.7)				
38 39 40 41		Serostatus – Previous qPCR Positive [†] Reactive Non-reactive	-	34 (82.9) 7 (17.1)	-				
42 43 44 45 46 47 48	375 376 377 378 379 380	qPCR: quantitative polymerase chain reaction. *N=2,888 participants who provided at least one b **N=2,076 participants who provided blood sample [†] N=41 participants who provided an endline blood positive qPCR test.	es at baseline and endlir		/-2 identified via				
49 50 51	381	Discussion							
52 53	382	This study provides a model of a	voluntary, incentiviz	ed system to identi	fy and link at-risk				
54 55 56	383	students to SARS-CoV-2 testing. While the	he incidence and se	roprevalence of SA	ARS-CoV-2 were				
57 58					18				
59 60		For peer review only - http://b	omjopen.bmj.com/site/	about/guidelines.xhtr	nl				

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generally low in this cohort of university students and employees in the summer of 2020, we observed the highest incidence among undergraduate students living in congregate settings, with nearly half of cases found to be associated with a super-spreader event.

At the time of the study, many infection control strategies centered on symptomatic testing, reducing the likelihood of identifying asymptomatic, mildly symptomatic, and pre-symptomatic infections. Our approach sought to integrate symptom-based monitoring with exposure monitoring, random surveillance testing, and targeted surveillance testing in the context of an outbreak. Within this cohort, we previously demonstrated the acceptability of our low-barrier SARS-CoV-2 mitigation approach and the limitations of temperature monitoring as a tool for case identification.[14,28] The present analysis builds upon these contributions by triangulating prospective gPCR testing data with phylogenetic analyses of positive samples and serial antibody testing to evaluate whether case identification and containment were achieved. In doing so, we found evidence that the system successfully identified a high proportion of incident SARS-CoV-2 cases among participants and may have mitigated community transmission after an outbreak. Specifically, 91% of participants with newly-identified antibodies for SARS-CoV-2 at the end of the study had also been diagnosed with incident infection via qPCR test during the study period. While a sizeable cluster of cases among participants was traced to a single super-spreader event, the associated cluster lineage was successfully contained without spreading beyond campus. As the outbreak unfolded, the system also allowed for rapid real-time response (i.e., surveillance testing notifications to students living in congregate housing) and offered a readily accessible, incentivized entry point for testing for students concerned about potential exposure.

Although some universities have adopted punitive measures intended to prevent transmission by controlling student behavior (for example, suspending students for hosting gatherings),[31–33] this approach has been criticized for its potential to reduce students' trust and cooperation.[34-36] Instead of punishing or shaming students who fail to adhere to public

health guidance, some epidemiologists have called for a harm-reduction approach which
supports and engages students as part of the solution.[34–36] The present study reinforces the
potential to integrate voluntary testing and risk monitoring systems to support targeted case
identification, as evidenced by the significantly higher positivity rates found among participants
whose self-reported symptoms and exposures triggered notifications to test. Our findings also
support increased outreach to groups of students at highest risk, particularly younger students
in congregate housing.

This study is strengthened by rich longitudinal data, including symptom and exposure tracking, qPCR testing, and seroprevalence data from more than 2,000 participants. The study population comprised of a broad sample of university affiliates, both students and employees, with strong representation of university subpopulations perceived to be at higher risk of infection (e.g., undergraduates, essential healthcare workers). As on-campus activities were severely restricted throughout the study period (all classes were held online, and few students were living in residence halls), this study cannot provide insight into SARS-CoV-2 transmission risks related to on-campus student activities. Nevertheless, as 73% of UC Berkeley undergraduate students lived off campus before the pandemic, [37] systems to detect off-campus (i.e., community and household) transmission remain important for SARS-CoV-2 monitoring efforts among students. Additionally, all participants in the essential workers cohort and a subset of participants in the faculty/cohort were working on campus during the study period, further motivating efforts to monitor incidence in this population.

430 There remain several limitations. We observed relatively few SARS-CoV-2 cases during
 431 the study period. Accordingly, although many associations are statistically significant, our
 432 estimates are imprecise (i.e., have wide confidence intervals) and must be interpreted with
 433 caution. This study took place before the development of highly transmissible variants, such as
 434 Delta and Omicron, and before vaccine rollout. Observed associations between symptoms and
 435 positivity may also differ among those who have been infected by more recent variants and/or

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vaccinated. Further research is necessary to adapt and evaluate similar systems in the context of both heightened transmissibility and more prevalent natural and vaccine-induced immunity. Additionally, a high proportion of identified cases were traced to one outbreak, limiting the generalizability of our exploratory assessment of risk factors for incident infection. There was also anecdotal evidence that the outbreak prompted exposed students to enroll as study participants.[14] While this self-referral into the study is likely to increase selection bias, it also illustrates the utility of implementing non-stigmatizing, incentivized testing approaches to increase testing uptake among at-risk students. Finally, our identification of participants who seroconverted between baseline and endline may be incomplete due to loss-to-follow up and imperfect sensitivity of SARS-CoV-2 antibody testing.

By integrating symptom and exposure monitoring systems with low-barrier testing, we identified incident SARS-CoV-2 infections to reduce transmission within a university setting. Our study contributes to a growing body of literature on novel, integrated SARS-CoV-2 surveillance strategies in university settings.[38-44] While there have been seismic shifts in the SARS-CoV-2 pandemic since 2020, universities continue to grapple with how best to mitigate on-campus spread in the face of emerging variants, incomplete vaccination coverage, breakthrough infections, and decreased reliance on other mitigation strategies (e.g., masking, remote learning).[45,46] In light of universities' resource constraints and persistently high case counts, incentivized approaches may not be feasible or sustainable in many settings. Thus, further research is needed to identify and test non-monetary incentives and other behavioral nudge strategies that encourage students and other campus community members to actively participate in public health efforts to combat the pandemic. The lessons learned through this study may inform the design of future adaptive strategies, ideally building beyond symptom/exposure monitoring and qPCR testing to integrate complementary interventions such as rapid antigen self-testing and vaccination promotion.

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461 Keywords: COVID-19, SARS-CoV-2, United States, young adults, students, universities,
462 essential workers, seroprevalence

464 Acknowledgements

We are grateful for the contributions of an exceptional team of graduate student researchers (Mariah De Zuzuarregui, Darren Frank, Sarah Gomez-Aladino, Ariel Muñoz, Ruben Prado, Lawrence Tello, Emily Wang, and Sabrina Williamson) and our collaborators at UC Berkeley's University Health Services (including but not limited to: Judith Sansone, Melody Heller, Holly Stern, Tyler Crooks, Desi Gallardo, Jeff Kreutzen, Rebecca Stephenson, Lisa Polley, and Melissa Hennings), the Innovative Genomics Institute (including but not limited to: Fyodor Urnov, Shana McDevitt, Ariana Hirsch, Alexander Ehrenberg, and the other members of the IGI SARS-CoV-2 testing consortium: M Amen, Kerrie W Barry, John M Boyle, Cara E Brook, Seunga Choo, L T Cornmesser, David J Dilworth, Jennifer A Doudna, Indro Fedrigo, Skyler E Friedline, Thomas G W Graham, Ralph Green, Jennifer R Hamilton, Megan L Hochstrasser, Dirk Hockemeyer, Netravathi Krishnappa, Azra Lari, Hangin Li, Enrique Lin-Shiao, Tianlin Lu, Elijah F Lyons, Kevin G Mark, Lisa Argento Martell, A Raquel O Martins, Patrick S Mitchell, Erica A Moehle, Christine Naca, Divya Nandakumar, Elizabeth O'Brien, Derek J Pappas, Kathleen Pestal, Diana L Quach, Benjamin E Rubin, Rohan Sachdeva, Elizabeth C Stahl, Abdullah Muhammad Syed, I-Li Tan, Amy L Tollner, Connor A Tsuchida, C Kimberly Tsui, Timothy K Turkalo, M Bryan Warf, Oscar N Whitney, and Lea B Witkowsky), and Vitalant Research Institute (including but not limited to: Mars Stone, Chloe Thorbrogger, Alice Lee, and Heather Tanner). The author would also like to thank Drs. Sandra McCoy, Stefano Bertozzi, and Lauren Ralph for their feedback on this manuscript.

484 Contributors: LH performed statistical analyses and wrote the first draft of the manuscript. SW
485 performed phylogenomic analyses and prepared associated figures and paragraphs. AR and
486 MP designed the study and provided input on the manuscript. LP, SF, AH, GN, the IGI SARS-

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487 CoV-2 Testing Consortium, CDG, and MB provided feedback on the study design and
488 manuscript. YL assisted with data analyses. All authors hold final responsibility for the decision
489 to submit for publication.

490 Declaration of interests: Vitalant Research Institute, of which Dr. Michael Busch is Director,
491 receives research funding and free assay kits from Ortho Clinical Diagnostics. Dr. Busch does
492 not receive salary support or personal compensation from Ortho Clinical Diagnostics. The
493 remaining authors declare no competing interests.

494 Funding: The study was funded by private donors who had no role in study design, data
495 collection, data analysis, data interpretation, or writing of the report.

496 Data sharing: De-identified data sets used in analyses and accompanying R Markdown script497 files will be publicly available at the time of publication at the following link:

498 https://github.com/lauren-hunter/bcsci

499 Ethical approvals: All study activities were approved by the University of California, Berkeley
 500 Committee for the Protection of Human Subjects (#2020-06-13349, #2020-05-13261, #2020-04 501 13238).

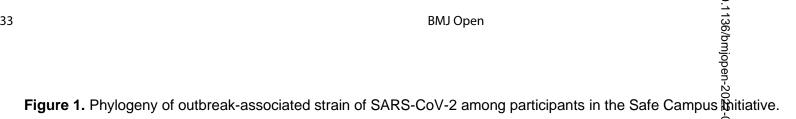
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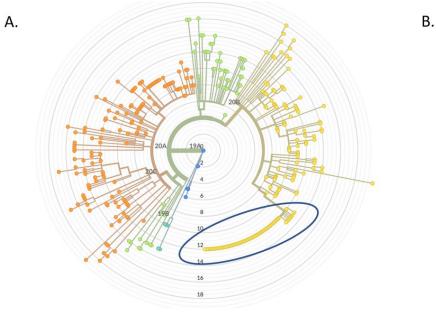
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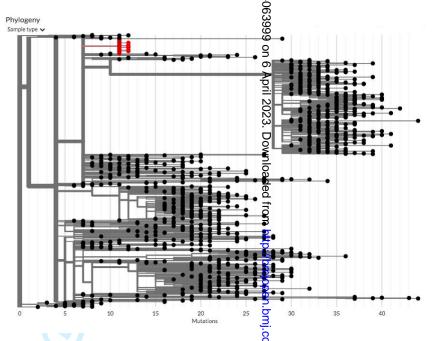
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A. A maximum likelihood phylogeny constructed from 357 genomes sequenced by the Innovative Genomics Institute between May and July 2020 constructed using Nextstrain. Branch lengths represent divergence from Wuhan reference genome at center. Blue circle marks cluster of identical genomes from a campus

super-spreader event. April 2021), showing the most similar genomes to the super-spreader event cluster (in red). There are no descendant branches from the cluster, demonstrating that the outbreak was contained and the lineage died out.

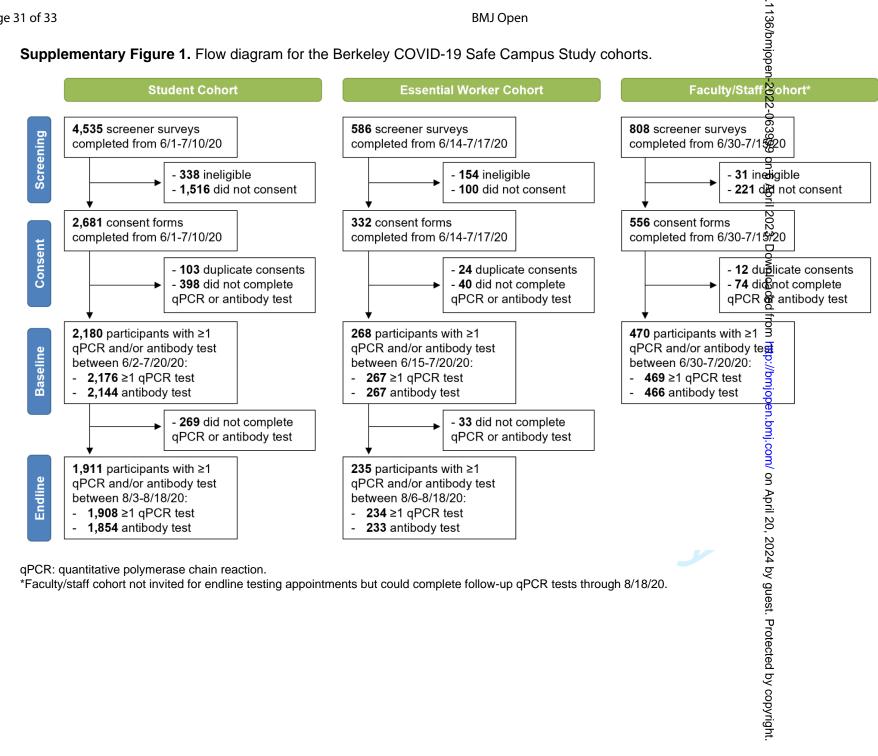
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Supplementary Table 1. Eligibility criteria across the Berkeley COVID-19 Safe Campus Study cohorts.

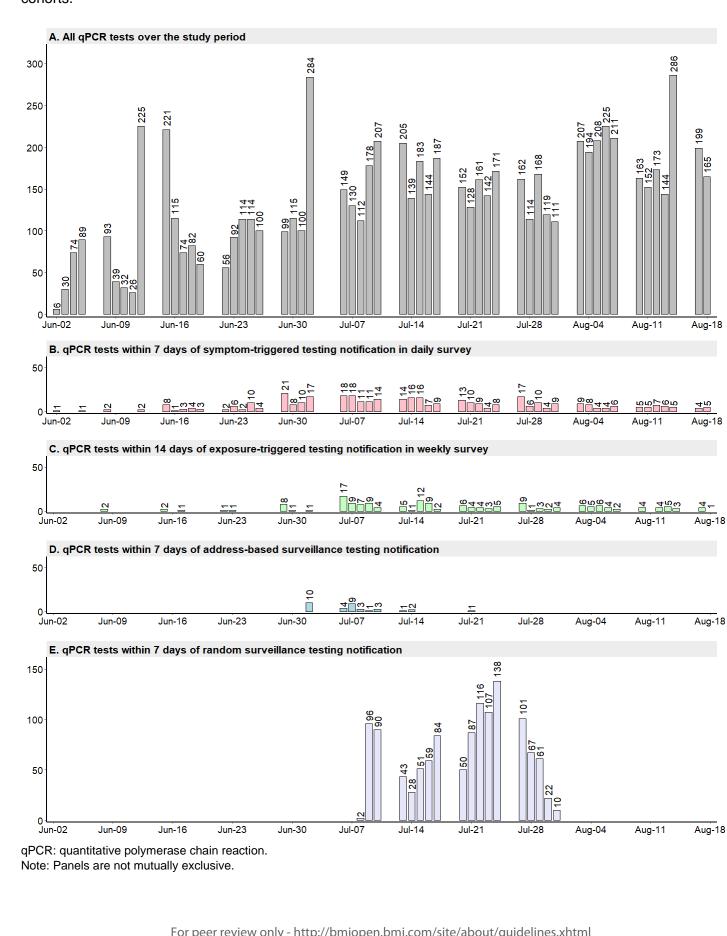
	Student Cohort	Essential Worker Cohort	Faculty/Staff & Ohort	
	- At least 18 years of age	- At least 18 years of age	- At least 18 years of age	
Eligibility Criteria	- Currently enrolled as an undergraduate or graduate student at UC Berkeley (i.e., not graduated in Spring 2020 or incoming for Fall 2020)	 Currently employed in one of the following departments at UC Berkeley: health services, police, facility services or other building management, environmental health and safety, laboratory animal care, athletics, dining, childcare, other residential or student services Currently working on campus at UC Berkeley <i>or</i> expected to return to work during June 2020 	- Currently employed a ga faculty member, staff member, or postdoctora scholar at UC Berkeley - Not already enrolled in the essential workers cohort	
	- Primarily residing in Alameda County or Contra Costa Country between 6/1/20-8/31/20	N/A	- Primarily residing in Asterneda County or Contra Costa Country between 6/1/20-8/31/20	
	- Willing to sign release of information for COVID-19-related medical records	- Willing to sign release of information for COVID-19-related medical records	- Willing to sign release of information for COVID-19-related medical records	
			bmj.com/ on April 20, 2024 by guest. Protected by copyright	

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Supplementary Figure 1. Flow diagram for the Berkeley COVID-19 Safe Campus Study cohorts.



Supplementary Figure 2. qPCR testing over time across the Berkeley COVID-19 Safe Campus Study cohorts.



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	STROBE 2007 (v4) Statement—Checklist of items that should be included in reports of conort studies					
Section/Topic	ltem #	Recommendation 9	Reported on page			
Title and abstract	1	(<i>a</i>) Indicate the study's design with a commonly used term in the title or the abstract	1-2			
		(b) Provide in the abstract an informative and balanced summary of what was done and what was figund	2			
Introduction	1					
Background/rationale	2	Explain the scientific background and rationale for the investigation being reported	4-5			
Objectives	3	State specific objectives, including any prespecified hypotheses	5			
Methods	1					
Study design	4	Present key elements of study design early in the paper	5			
Setting	5	Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection	5-9, Supplementa Figure 1			
Participants	6	(a) Give the eligibility criteria, and the sources and methods of selection of participants. Describe m_{eff}^{\exists} thods of follow-up	5-9, Supplementa Table 1			
		(b) For matched studies, give matching criteria and number of exposed and unexposed	N/A			
Variables	7	Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable	6-10			
Data sources/ measurement	8*	For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe	6-10			
Bias	9		10			
Study size	10	Explain how the study size was arrived at	6			
Quantitative variables	11	Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why	10			
Statistical methods	12	(a) Describe all statistical methods, including these used to control for confounding	10			
		(a) Describe an statistical methods, including those used to control for combanding (b) Describe any methods used to examine subgroups and interactions (c) Explain how missing data were addressed	N/A			
			N/A			
		(d) If applicable, explain how loss to follow-up was addressed	N/A			
		(e) Describe any sensitivity analyses	N/A			

Results		BMJ Open	
Participants	13*	(a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examine for eligibility, confirmed	Supplementary
		eligible, included in the study, completing follow-up, and analysed	Figure 1
		(b) Give reasons for non-participation at each stage	Supplementary Figure 1
		(c) Consider use of a flow diagram	Supplementary Figure 1
Descriptive data	14*	(a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders	11, Table 1
		(b) Indicate number of participants with missing data for each variable of interest	11-13
		(c) Summarise follow-up time (eg, average and total amount)	11
Outcome data	15*	Report numbers of outcome events or summary measures over time	12
Main results	16	(a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision geg, 95% confidence	12-14, Tables 2-3
		interval). Make clear which confounders were adjusted for and why they were included	
		(b) Report category boundaries when continuous variables were categorized	10, Table 3
		(c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period	N/A
Other analyses	17	Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses	N/A
Discussion		ă și cara cara cara cara cara cara cara car	
Key results	18	Summarise key results with reference to study objectives	14-15
Limitations			
Interpretation	20	Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of a lyses, results from similar studies, and other relevant evidence	14-17
Generalisability	21	Discuss the generalisability (external validity) of the study results	16
Other information			
Funding	22	Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based	19
*Give information sepa	arately fo	which the present article is based r cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and, if applicable, for exposed and unexposed groups in controls in case-control studies and it is the control stu	tudies.