



BMJ Open is committed to open peer review. As part of this commitment we make the peer review history of every article we publish publicly available.

When an article is published we post the peer reviewers' comments and the authors' responses online. We also post the versions of the paper that were used during peer review. These are the versions that the peer review comments apply to.

The versions of the paper that follow are the versions that were submitted during the peer review process. They are not the versions of record or the final published versions. They should not be cited or distributed as the published version of this manuscript.

BMJ Open is an open access journal and the full, final, typeset and author-corrected version of record of the manuscript is available on our site with no access controls, subscription charges or pay-per-view fees (<http://bmjopen.bmj.com>).

If you have any questions on BMJ Open's open peer review process please email [info.bmjopen@bmj.com](mailto:info.bmjopen@bmj.com)

# BMJ Open

## **BioFACTS - Biomarkers of rhabdomyolysis in the diagnosis of acute compartment syndrome: protocol for a prospective multinational, multi-centre study involving patients with tibial fractures**

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2021-059918
Article Type:	Protocol
Date Submitted by the Author:	05-Dec-2021
Complete List of Authors:	<p>Nilsson, Abraham; Linköping University Hospital, Department of Orthopaedics and Department of Biomedical and Clinical Sciences, Faculty of Health Science</p> <p>Ibounig, Thomas; University of Helsinki, Department of Orthopaedics and Traumatology</p> <p>Lyth, Johan; Linköping University Department of Health Medicine and Caring Sciences,</p> <p>Alkner, Björn; Linköping University, Department of Orthopaedics, Eksjö, Region Jönköping County and Department of Biomedical Clinical Science von Walden, Ferdinand; Karolinska Institutet,</p> <p>Fornander, Lotta; Linköping University Faculty of Medicine, Department of Orthopaedics, Vrinnevi Hospital</p> <p>Rämö, Lasse; University of Helsinki, Orthopaedics and traumatology</p> <p>Schmidt, Andrew; Hennepin Healthcare, Department of orthopaedics</p> <p>Schilcher, Jörg; Linköping University Hospital, Department of Orthopaedics and Department of Biomedical and Clinical Sciences, Faculty of Health Science</p>
Keywords:	Musculoskeletal disorders < ORTHOPAEDIC & TRAUMA SURGERY, ORTHOPAEDIC & TRAUMA SURGERY, Adult orthopaedics < ORTHOPAEDIC & TRAUMA SURGERY

SCHOLARONE™  
Manuscripts

1 BioFACTS - Biomarkers of rhabdomyolysis in the diagnosis  
2 of acute compartment syndrome: protocol for a prospective  
3 multinational, multi-centre study involving patients with tibial  
4 fractures

5  
6 Abraham Nilsson<sup>1</sup>, Thomas Ibounig<sup>2</sup>, Johan Lyth<sup>3</sup>, Björn Alkner<sup>4</sup>, Ferdinand von Walden<sup>5</sup>,  
7 Lotta Fornander<sup>6</sup>, Lasse Rämö<sup>2</sup>, Andrew Schmidt<sup>7</sup> and Jörg Schilcher<sup>1, 8</sup>

8  
9 <sup>1</sup> Department of Orthopaedic Surgery and Department of Biomedical and Clinical Sciences, University  
10 Hospital Linköping, Linköping University, Linköping, Sweden

11 <sup>2</sup> Department of Orthopaedics and Traumatology, Helsinki University Hospital and University of  
12 Helsinki, Finland

13 <sup>3</sup> Department of Health, Medicine and Caring Sciences, Linköping University, Linköping, Sweden

14 <sup>4</sup> Department of Orthopaedics, Eksjö, Region Jönköping County and Department of Biomedical and  
15 Clinical Sciences, Linköping University, Linköping, Sweden

16 <sup>5</sup> Department of Women's and Children's Health, Karolinska Institutet, Stockholm, Sweden

17 <sup>6</sup> Department of Orthopaedic Surgery in Norrköping and Department of Biomedical and Clinical  
18 Sciences, Linköping University, Norrköping, Sweden

19 <sup>7</sup> Department of Orthopaedics, Hennepin Healthcare, Minneapolis, MN, USA

20 <sup>8</sup> Wallenberg Centre for Molecular Medicine, Linköping University, Linköping, Sweden

21  
22 Abraham Nilsson, (abraham.nilsson@regionostergotland.se)  
23 Thomas Ibounig, (thomas.ibounig@hus.fi)  
24 Johan Lyth, (johan.lyth@liu.se)

25 Björn Alkner, (bjorn.alkner@liu.se)  
26 Ferdinand von Walden, (ferdinand.von.walden@ki.se)  
27 Lotta Fornander, (lotta.fornander@liu.se)  
28 Lasse Rämö, (lasse.ramo@hus.fi)  
29 Andrew Schmidt, (andrew.schmidt@hcmmed.org)  
30 Jörg Schilcher, (jorg.schilcher@liu.se)\*

31 **\*Corresponding author**

32

33 **Word count: 2712**

34

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**ABSTRACT**

**Introduction**

The ischemic pain of acute compartment syndrome (ACS) can be difficult to discriminate from the pain linked to an associated fracture. Lacking objective measures, the decision to perform fasciotomy is based on clinical findings and performed at a low level of suspicion. Biomarkers of muscle cell damage may help to identify and monitor patients at risk, similar to current routines for patients with acute myocardial infarction. This study will test the hypothesis that biomarkers of muscle cell damage can predict ACS in patients with tibial fractures.

**Methods and analysis**

Patients aged 15–65 years who have suffered a tibial fracture will be included. Plasma (P)-myoglobin and P-creatine phosphokinase (P-CK) will be analysed at 6-hourly intervals after admission to the hospital and – if applicable – after surgical fixation or fasciotomy. In addition, if ACS is suspected, blood samples will be collected at 6-hourly intervals. An independent expert panel will assess retrospectively the study data and will classify those patients who had undergone fasciotomy into those with ACS and those without ACS. The area under the receiver operator characteristics curves will be used to identify the success of the biomarkers in discriminating between fracture patients who develop ACS and those who do not. Logistic regression analyses will be used to assess the discriminative abilities of the biomarkers to predict ACS corrected for pre-specified covariates. In a separate, exploratory analysis we collect blood samples from patients with ACS but without tibial fractures, to allow comparison with fracture patients who developed ACS.

## 60 **Ethics and dissemination**

61 The study has been approved by the Regional Ethical Review Boards in Linköping  
62 (2017/514-31) and Helsinki/Uusimaa (HUS/2500/2000) and has been registered at  
63 *Clinicaltrials.gov* (Identifier: NCT04674592). The BioFACTS study will be reported in  
64 accordance with the Strengthening the Reporting of Observational Studies in Epidemiology  
65 recommendations.

## 67 **Keywords**

68 Musculoskeletal disorders, orthopaedic & trauma surgery, adult orthopaedics

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**STRENGTHS AND LIMITATIONS OF THIS STUDY**

- The first prospective study to investigate plasma levels of P-myoglobin and P-CK in patients with tibial fractures with and without ACS.
- Multiple participating centres, including different hospital categories from two Nordic countries.
- Expert panel assessment of ACS.

**INTRODUCTION**

Acute compartment syndrome (ACS) is a serious complication in trauma. As the condition is difficult to diagnose and the consequences of a missed compartment syndrome are detrimental, decompressive fasciotomy is performed when there is a low level of suspicion, possibly often leading to unnecessary surgeries.[1]

ACS is caused by either a volume expansion of the muscles, e.g., muscular swelling, or a decrease in the compartment space, e.g., burn injuries, leading to increased intra-compartmental pressure. The increased pressure, in turn, leads to decreased perfusion and, if left untreated, tissue necrosis.[2] ACS can emerge in a wide variety of locations, although most commonly it affects the lower leg due to its tight inelastic fasciae.[3] Tibial fracture is the most common injury associated with ACS, and ACS develops in 2%–8% of the cases.[2, 4] The cardinal symptom of ACS is pain, which can be very difficult to differentiate from the pain caused by a fracture. The treatment for ACS is urgent surgical decompression of the affected muscle compartments through fasciotomy. However, fasciotomy involves a large incision of the skin and compromises the soft tissue envelope around the fractured bone. Therefore, fasciotomy has a profound negative impact on the possibilities for orthopaedic treatment of the fracture, increases the risk for complications, prolongs hospital stays, and

drives up costs.[5-7] Therefore, correct and timely diagnosis is of the outmost importance for these patients.

Currently, ACS is diagnosed using a combination of physical findings and intra-compartmental pressure measurements. Each of these measures has inherent drawbacks in terms of making the correct diagnosis.[1, 8, 9] Specific pressure thresholds at which fasciotomy should be performed have been proposed.[9-11] However, studies have shown that up to 84% of patients with a tibial fracture exceed this pressure threshold without developing ACS.[12] A recent study has shown that perfusion pressure has low specificity,[13] and, furthermore, the pressure varies with distance to the fracture, making the interpretation even more difficult.[9] Inappropriate use of the method may, therefore, lead to unnecessary fasciotomies. Other promising diagnostic modalities include near-infrared spectroscopy (NIRS). Decreased tissue oxygenation levels correlate with increased intramuscular pressure.[14] NIRS can detect a sudden decrease in tissue oxygenation in patients with ACS,[15] although the reliability of NIRS in an injured leg remains uncertain and its role in the diagnosis of ACS has not been defined.[16] Biomarkers, including measurements of pH and intramuscular glucose might allow the identification of patients with impaired muscle metabolism due to ACS. Also, circulating microRNAs might be a potential tool for the future.[17] However, none of these techniques is used in clinical routine today. In the absence of good objective measures and a clear definition of when ACS is present, decision-making regarding fasciotomy relies on the judgment of the individual doctor, which leads to significant variability (2%–24%) in the percentage of fasciotomies performed for ACS per surgeon.[18]

Similar to the diagnosis of acute myocardial infarction, whereby heart muscle-specific Troponin T is measured, markers of muscle cell damage (e.g., P-myoglobin) may be used to diagnose objectively or to monitor ACS. The use of such markers has previously been deemed



1  
2  
3 118 unfeasible on the basis that pathological levels of these markers could be attributed to sources  
4  
5 119 other than the actual muscle compartment, e.g., traumatic muscle damage and heart  
6  
7 120 contusion.[19] Nonetheless, some studies have shown the potential of biomarkers to improve  
8  
9 121 the diagnosis of ACS,[20, 21] albeit never in the presence of a fracture.  
10  
11  
12 122 We have recently shown that high intramuscular pressure coincides with high P-myoglobin  
13  
14 123 levels and that myoglobin may be a relevant, yet unexplored diagnostic tool in ACS  
15  
16  
17 124 associated with trauma.[22]  
18  
19 125  
20

21 126 **Hypothesis**

22  
23  
24 127 P-myoglobin and P-CK can be used to predict ACS in patients with traumatic tibial fractures.  
25  
26 128  
27

28 129 **Aims**

29  
30  
31 130 *Primary aim*

32  
33 131 To describe the diagnostic performances of P-myoglobin and P-CK to predict ACS in patients  
34  
35 132 with traumatic tibial fractures.  
36  
37 133  
38

39  
40 134 *Secondary aim*

41  
42 135 To compare the pathological changes in circulating microRNA (miRNA) and muscle biopsies  
43  
44 136 with the levels of P-myoglobin and P-CK in fracture patients.  
45  
46  
47 137 To compare changes in P-myoglobin and P-CK between fracture patients with ACS and non-  
48  
49 138 fracture patients with ACS.  
50

51 139

52  
53 140 **METHODS AND ANALYSIS**

54  
55  
56 141 The study is a prospective, multi-national, multi-centre study. The study is currently running  
57  
58 142 in Sweden at Linköping University Hospital, Vrinnevi Hospital in Norrköping,  
59  
60

1  
2  
3 143 Högländssjukhuset in Eksjö and Kalmar Hospital, as well as in Finland at Töölö Hospital in  
4  
5 144 Helsinki. Additional hospitals may be included in the future. To enable exploratory analyses  
6  
7 145 of biomarkers from fracture patients with ACS compared with patients suffering ACS but  
8  
9 146 without fractures, we will include a No-fracture group with ACS of the lower leg.  
10  
11  
12 147

## 148 **Study Population**

### 149 *Fracture group*

150 Patients in the age range of 15–65 years who have suffered traumatic fractures of the tibia will  
151 be included. All primary comparisons will be performed between fracture patients with and  
152 without ACS.  
153

### 154 *No-fracture group*

155 Patients in the age range of 15–65 with suspected ACS of the lower leg but *without* associated  
156 fractures will be included to enable exploratory comparisons with fracture patients with ACS.  
157

158 Exclusion criteria for the groups are listed in Table 1.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

Table 1. Inclusion and exclusion criteria.

Fracture group	No-fracture group
<i>Inclusion criteria</i>	
Traumatic tibial fracture* 15–65 years old	Suspected acute compartment syndrome 15–65 years old
<i>Exclusion criteria</i>	
Malignancy	
Acute myocardial infarction	
Kidney failure (GFR $\leq$ 35 ml/min)	
Muscle disease	
Paraplegia/tetraplegia	
	Any other associated fracture
	Acute vascular event

\*Any anatomical location of the tibia, excluding solitary ankle fractures.

**Outcome measures**

*Primary*

Myoglobin is an important myocyte-produced compound that is released into the bloodstream as the first enzyme to show increased levels following any type of muscle injury. Myoglobin becomes measurable when the protein binding capacity is exceeded after 1–3 hours. It peaks after 8-12 hours, and it is cleared from the plasma within 24 hours.[23]

The levels of CK in plasma increase within 12 hours of muscle injury, peak after 24–36 hours, and decrease at a rate of 30%–40% per day.[23] Both P-myoglobin and P-CK are routinely measured in the diagnosis and monitoring of rhabdomyolysis.[23] Therefore, facilities for the

171 analysis of these enzymes are available in most clinical settings. The analyses are relatively  
172 cheap (roughly €30 per sample) and can be performed in less than 1 hour.  
173 For the present study, we chose a combination of these enzymes and a 6-hourly interval over a  
174 24-hour or 48-hour period so as to be able to detect the post-injury peak of at least one of  
175 these enzymes. Considering the urgency of the diagnosis, we will primarily focus on an  
176 increase in the level of myoglobin as an early sign of muscle injury.

177

### 178 *Secondary*

179 We will collect blood samples for later analysis of muscle-specific miRNA at the same time  
180 intervals as for the primary outcome. Due to the rapidly evolving field of miRNA research,  
181 we abstain to prespecify specific miRNA's that will be used as objective measures of muscle  
182 damage.[24] At the time of surgery (internal fixation or fasciotomy), biopsies are taken at  
183 some centres for further histological analyses. Two biopsies are taken from the tibialis  
184 anterior muscle in the fractured leg, one close to the injury and one at a distance. One biopsy  
185 is taken from the same muscle in the uninjured leg (control). Biopsies are frozen within 30  
186 minutes using liquid nitrogen-cooled isopentane.[25] For storage, the samples are kept at -  
187 80°C. Since P-myoglobin and P-CK can be affected by renal function,[26] we will analyse the  
188 level of serum creatine and the glomerular filtration rate (GFR). As sex, body weight and age  
189 can affect muscle volume, these parameters will also be registered. We will record the  
190 mechanism of injury (high or low energy), type of fracture (proximal, mid-shaft or distal and  
191 AO/OTA classification), trauma severity (solitary tibial fracture (with or without fibula  
192 fracture), tibial fracture with concomitant long bone fracture, tibial fracture in combination  
193 with multi-trauma), peri-operative findings of muscle viability (colour, consistency,  
194 contractility, capacity to bleed), and whether the muscle bulges at the point of incision.  
195 Stratified analysis for these subgroups will be performed if feasible.

1  
2  
3 196 *Timing of blood samples*

5 197 Plasma for assaying the levels of myoglobin and CK will be collected at those time-points at  
7  
8 198 which the patient is at high risk to develop ACS (Figure 1):

- 10 199 1) After the trauma, at admission to hospital: at 6-hourly intervals for a maximum of 48  
12 200 hours or until definitive surgical fixation.  
14 201 2) After definitive surgical fracture treatment: at 6-hourly intervals for 24 hours.  
16 202 3) If there is suspicion of ACS, blood samples will be collected at 6-hourly intervals until  
18 203 fasciotomy is performed or the suspicion is dismissed, although no longer than 48 hours.  
20 204 4) After fasciotomy, blood sampling will be continued at 6-hourly intervals for 24 hours.  
22  
23 205 Fasciotomy will be performed according to clinical routine praxis, as deemed feasible by the  
24  
25 206 responsible surgeon.  
26  
27  
28 207

30 208 **Blood sample analysis**

32 209 The levels of P-myoglobin, P-CK and S-creatinine in patients at Linköping, Norrköping and  
34 210 Helsinki will be analysed at the respective hospital. Blood samples from Eksjö and Kalmar  
36 211 will be centrifuged, stored refrigerated, and sent to Linköping for analysis. EDTA-tubes (for  
38 212 analysis of miRNA) will be centrifuged at 2,500 rpm at room temperature for 10 minutes at  
40 213 each hospital. In the Swedish centres, the supernatant will be immediately and carefully  
42 214 removed, frozen in cryo-tubes, and sent to and stored at -80°C until further processing. In  
44 215 Helsinki, the samples will be stored at the premises of the Helsinki Biobank prior to further  
46 216 analysis.  
48  
49  
50 217

52 218 **Expert panel assessment of ACS in patients with fasciotomy**

54 219 Once collected, the data will be anonymised, uniformly compiled and reviewed by an  
56  
57 220 independent expert panel of senior orthopaedic surgeons. The panel will retrospectively assess  
58  
59  
60

the clinical data (injury characteristics, radiographs, pain medication administered, details as to the surgical procedures). Thereafter, the panel will:

- 1) Specify whether or not the patient had an ACS;
- 2) Indicate the two most important factors contributing to that decision; and
- 3) Determine if there are any missing data that might have influenced their decision.

The numbers of patients with ACS adjudged by the expert panel will be compared with the numbers of patients undergoing fasciotomy for ACS.

### **Follow-up**

Patients will be followed clinically with individualised intervals according to routine care and at 1 year. The 1-year follow-up will include completion of the Lower Extremity Functional Score (LEFS) questionnaire and a clinical examination. The findings will be used for descriptive purposes, in particular to detect patients who present with functional deficits that might be related to an undetected ACS. In all fasciotomized patients and those with suspected ACS-related functional impairment found during the clinical examination, we aim to perform bilateral MRI examinations of the lower leg to explore the possibility of MRI as an objective measure of ACS induced tissue damage.

### **Sample size calculation and statistical analysis plan**

We have performed two separate sample size calculations. The first calculation is based on pilot studies on patients with tibial fractures and patients with tibial fractures complicated by fasciotomy due to suspected ACS. The mean pre-operative values for P-myoglobin were 289 and 1449 µg/L, respectively, with standard deviations of 249 and 1044 µg/L, respectively. This corresponds to an effect size of 1.1 if we are conservative and use the largest standard deviation of 1044 µg/L from both groups. Using this effect size of 1.1 with a 2-sided test and

alpha of 0.05, we need a sample size of 14 patients undergoing fasciotomy due to suspected ACS to achieve 80% power. If we expect a fasciotomy prevalence of 5% of patients with tibial fractures, the expected number of non-fasciotomised patients is 266. In the second calculation, we assumed an improvement of the area under the curve (AUC) value from 0.5 (as good as chance) to 0.7 (acceptable diagnostic accuracy[27]), with an alpha of 0.05, a power of 80% and ACS prevalence of 5% in the population. Under these assumptions, we require 16 patients with fasciotomy due to suspected ACS and 311 non-fasciotomised patients. With 3–5 patients with fasciotomies due to suspected ACS presenting within the current study network every year, we estimate an inclusion period of 3 years. Inclusion will continue until 16 patients with tibial fractures and ACS have been recruited, irrespective of the total number of recruited patients.

The difference in P-myoglobin levels between patients with fractures and suspected ACS, and non-fasciotomised fracture patients will be calculated with an independent samples Welch's t-test assuming different standard deviations. The AUC will be used to identify the success of these biomarkers to discriminate fracture patients with ACS from those without.

Logistic regression analyses will be used to assess the discriminative ability of a combination of the two biomarkers to predict ACS, using correction for the following covariates: GFR; sex; body weight; age; trauma mechanism (high or low energy); fracture type (proximal, mid-shaft or distal and AO/OTA classification); and trauma severity (solitary tibial fracture (with or without fibula fracture), tibial fracture with concomitant long bone fracture, tibial fracture in combination with multi-trauma). Due to the limited sample size, we decide to use Leave-one-out cross-validation (LOOCV) for internal validation, which corresponds to analysis type 1b in the TRIPOD checklist.[28] A professional statistician (blinded to the clinical

parameters) will perform the statistical analyses. All data will be anonymised and stored in secure servers within Region Östergötland, Sweden.

Separate statistical analyses will be performed for those patients who have undergone fasciotomies due to suspected ACS and those deemed as true ACS cases by the expert panel. When feasible, stratified subgroup analyses based on the abovementioned covariates will be performed.

### **Trial status**

Patient recruitment started at Linköping University Hospital on April 1<sup>st</sup>, 2018. September 1<sup>st</sup>, 2019 a final study protocol was implemented at Linköping University Hospital and in subsequent months at Höglandssjukhuset district hospital Eksjö, Vrinnevi Hospital in Norrköping, and Kalmar Hospital. In January 2021, Helsinki University Hospital, Finland started to recruit patients. Currently, approximately 150 patients have been included. Of these, two patients in the Fracture group underwent fasciotomy due to suspected ACS and six patients were fasciotomised in the No-fracture comparison group.

### *Study time schedule*

2021: Preliminary analysis of blood samples, muscle biopsies and miRNA to assess the quality levels of these samples.

2021: 150 patients included.

2022: 280 patients included.

2023: Data analysis and manuscript writing.

### **Patient and public involvement**



1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

296 Neither patients nor members of the public are involved in the design, conduct, reporting, or  
297 dissemination plans of this study.

For peer review only

## ETHICS AND DISSEMINATION

The study was approved by the Regional Ethical Review board in Linköping (Dnr. 2017/514-31) and the Ethical Review Board of the Hospital District of Helsinki and Uusimaa (HUS/2500/2000) and has been registered at *Clinicaltrials.gov* (Identifier: NCT04674592).

Oral and written explanations will be provided to all eligible patients, and written consent will be obtained as soon as possible but no later than during clinical rounds the morning after enrolment. Patients might feel compelled to participate in the study as the treating surgeon usually is the same person that seeks the patient's consent. This is a problem that cannot be avoided in the context of recruitment performed in emergency situations.

As all the samples will be analysed in a blinded fashion, there is no risk that these values will disrupt the clinical decision-making process in the emergency situation. The risk for complications associated with study participation is low, and there is a potential diagnostic benefit. The study will increase the level of awareness and knowledge of medical staff. Therefore, regardless of the results, the implementation of the study will increase patient safety and, thereby, balance out any risks that study participation may entail.

If we will be able to define threshold values of P-myoglobin and P-CK for the detection of ACS with good diagnostic accuracy, these values could be implemented in clinical practice without delay. Specifically, these threshold values would allow the individual surgeon to decide to abstain from fasciotomy and instead observe the patient and follow the biomarker dynamics.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**Acknowledgements**

To patients and personnel at all study sites for their invaluable contributions. Stefan Wetterstad for assistance in study organisation and patient recruitment at Kalmar Hospital, Sweden.

**Author contributions**

JS conceptualised the study idea.  
AN and JS planned and implemented the study at all the study sites.  
TI and LR planned and implemented the study at the Helsinki University Hospital.  
JL, AN and JS planned the statistical analysis.  
BA provided advice on biomarker handling and analysis and planned and implemented the study at Eksjö Hospital.  
FW provided advice on biomarker handling and analysis.  
LF planned and implemented the study at Norrköping hospital.  
AS contributed to the overall design of the study.  
All the authors contributed to the design of the study and revised and approved the final manuscript.

**Funding**

This work was supported by Linköping University, Region Östergötland, the Medical Research Council of Southeast Sweden (grant no. FORSS-852501 and RALF-0600911), the Wallenberg Foundation, Futurum – the Academy for Health and Care, Region Jönköping

347 County, Sweden (grant no. FUTURUM-937508 and 870471) and Finnish state funding for  
348 university-level health research.  
349

For peer review only

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**Availability of data and material**

The datasets generated and/or analysed during the current study may be obtained from the corresponding author on reasonable request.

**Competing interests**

The authors declare that they have no competing interests.

**FULL REFERENCES**

1. Schmidt AH. Continuous compartment pressure monitoring-better than clinical assessment? *J Bone Joint Surg Am.* 2013;95(8):e52.

2. Schmidt AH. Acute compartment syndrome. *Injury.* 2017;48 Suppl 1:S22-S5.

3. Matsen FA, 3rd, Krugmire RB, Jr. Compartmental syndromes. *Surg Gynecol Obstet.* 1978;147(6):943-9.

4. Park S, Ahn J, Gee AO, Kuntz AF, Esterhai JL. Compartment syndrome in tibial fractures. *J Orthop Trauma.* 2009;23(7):514-8.

5. Dover M, Memon AR, Marafi H, Kelly G, Quinlan JF. Factors associated with persistent sequelae after fasciotomy for acute compartment syndrome. *J Orthop Surg (Hong Kong).* 2012;20(3):312-5.

6. Schmidt AH. The impact of compartment syndrome on hospital length of stay and charges among adult patients admitted with a fracture of the tibia. *J Orthop Trauma.* 2011;25(6):355-7.

7. Lollo L, Grabinsky A. Clinical and functional outcomes of acute lower extremity compartment syndrome at a Major Trauma Hospital. *Int J Crit Illn Inj Sci.* 2016;6(3):133-42.

8. Shuler FD, Dietz MJ. Physicians' ability to manually detect isolated elevations in leg intracompartmental pressure. *J Bone Joint Surg Am.* 2010;92(2):361-7.

9. Heckman MM, Whitesides TE, Jr., Grewe SR, Rooks MD. Compartment pressure in association with closed tibial fractures. The relationship between tissue pressure, compartment, and the distance from the site of the fracture. *J Bone Joint Surg Am.* 1994;76(9):1285-92.

10. Whitesides TE, Haney TC, Morimoto K, Harada H. Tissue pressure measurements as a determinant for the need of fasciotomy. *Clinical orthopaedics and related research.* 1975(113):43-51.

11. McQueen MM, Court-Brown CM. Compartment monitoring in tibial fractures. The pressure threshold for decompression. *J Bone Joint Surg Br.* 1996;78(1):99-104.

12. Prayson MJ, Chen JL, Hampers D, Vogt M, Fenwick J, Meredick R. Baseline compartment pressure measurements in isolated lower extremity fractures without clinical compartment syndrome. *J Trauma.* 2006;60(5):1037-40.

13. Schmidt AH, Di J, Zipunnikov V, Frey KP, Scharfstein DO, O'Toole RV, et al. Perfusion Pressure Lacks Diagnostic Specificity for the Diagnosis of Acute Compartment Syndrome. *J Orthop Trauma.* 2020;34(6):287-93.

14. Shuler MS, Reisman WM, Kinsey TL, Whitesides TE, Jr., Hammerberg EM, Davila MG, et al. Correlation between muscle oxygenation and compartment pressures in acute compartment syndrome of the leg. *J Bone Joint Surg Am*. 2010;92(4):863-70.
15. Shuler MS, Reisman WM, Cole AL, Whitesides TE, Jr., Moore TJ. Near-infrared spectroscopy in acute compartment syndrome: Case report. *Injury*. 2011;42(12):1506-8.
16. Schmidt AH, Bosse MJ, Obrebsky WT, O'Toole RV, Carroll EA, Stinner DJ, et al. Continuous Near-Infrared Spectroscopy Demonstrates Limitations in Monitoring the Development of Acute Compartment Syndrome in Patients with Leg Injuries. *J Bone Joint Surg Am*. 2018;100(19):1645-52.
17. Etheridge A, Lee I, Hood L, Galas D, Wang K. Extracellular microRNA: a new source of biomarkers. *Mutat Res*. 2011;717(1-2):85-90.
18. O'Toole RV, Whitney A, Merchant N, Hui E, Higgins J, Kim TT, et al. Variation in diagnosis of compartment syndrome by surgeons treating tibial shaft fractures. *J Trauma*. 2009;67(4):735-41.
19. Shadgan B, Menon M, O'Brien PJ, Reid WD. Diagnostic techniques in acute compartment syndrome of the leg. *J Orthop Trauma*. 2008;22(8):581-7.
20. Valdez C, Schroeder E, Amdur R, Pascual J, Sarani B. Serum creatine kinase levels are associated with extremity compartment syndrome. *J Trauma Acute Care Surg*. 2013;74(2):441-5; discussion 5-7.
21. Hefler F. Compartment syndrome after gynecologic laparoscopy: systematic review of the literature and establishment of normal values for postoperative serum creatine kinase and myoglobin levels. *Arch Gynecol Obstet*. 2017;296.2:285-93.
22. Nilsson A, Alkner B, Wetterlov P, Wetterstad S, Palm L, Schilcher J. Low compartment pressure and myoglobin levels in tibial fractures with suspected acute compartment syndrome. *BMC Musculoskelet Disord*. 2019;20(1):15.
23. Giannoglou GD, Chatzizisis YS, Misirli G. The syndrome of rhabdomyolysis: Pathophysiology and diagnosis. *Eur J Intern Med*. 2007;18(2):90-100.
24. Siracusa J, Koulmann N, Bourdon S, Goriot ME, Banzet S. Circulating miRNAs as Biomarkers of Acute Muscle Damage in Rats. *Am J Pathol*. 2016;186(5):1313-27.
25. Dubowitz V, A. Sewry C, Oldfors A. Muscle biopsy: a practical approach: expert consult; online and print.: Elsevier Health Sciences; 2013. 8-10 p.
26. Hällgren R, Karlsson FA, Roxin LE, Venge P. Myoglobin turnover--influence of renal and extrarenal factors. *Journal of Laboratory and Clinical Medicine*. 1978;91(2).
27. Mandrekar JN. Receiver Operating Characteristic Curve in Diagnostic Test Assessment. *Journal of Thoracic Oncology*. 2010;5(9):1315-6.
28. Collins GS, Reitsma JB, Altman DG, Moons KG. Transparent reporting of a multivariable prediction model for Individual Prognosis or Diagnosis (TRIPOD): the TRIPOD statement. *J Clin Epidemiol*. 2015;68(2):134-43.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**Legends**

Figure 1. Flowchart of patient recruitment.

For peer review only

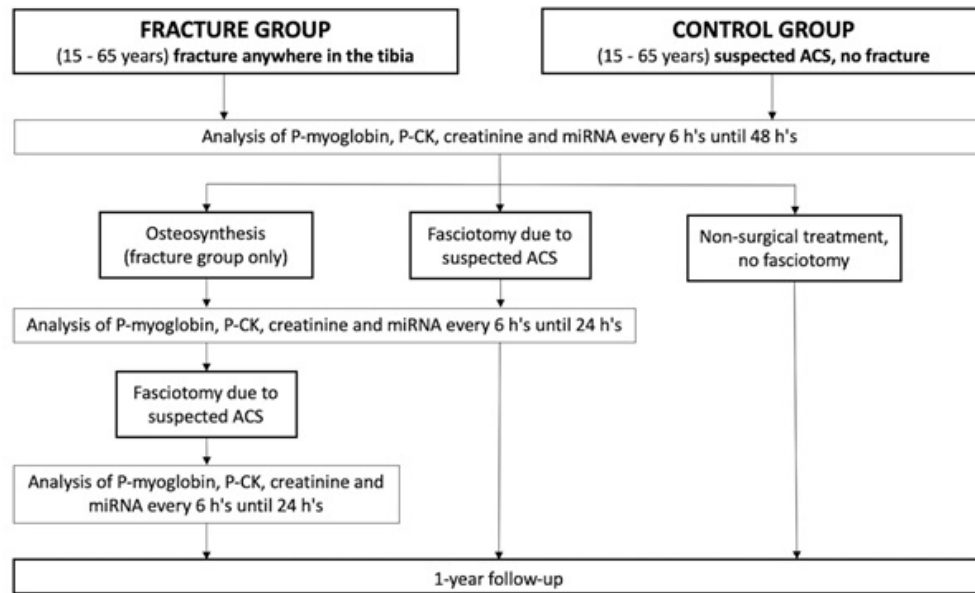


Figure 1. Flowchart of patient recruitment.

160x95mm (96 x 96 DPI)



# BMJ Open

## BioFACTS - Biomarkers of rhabdomyolysis in the diagnosis of acute compartment syndrome: protocol for a prospective multinational, multi-centre study involving patients with tibial fractures

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2021-059918.R1
Article Type:	Protocol
Date Submitted by the Author:	25-Mar-2022
Complete List of Authors:	<p>Nilsson, Abraham; Linköping University Hospital, Department of Orthopaedics and Department of Biomedical and Clinical Sciences, Faculty of Health Science</p> <p>Ibounig, Thomas; University of Helsinki, Department of Orthopaedics and Traumatology</p> <p>Lyth, Johan; Linköping University Department of Health Medicine and Caring Sciences,</p> <p>Alkner, Björn; Linköping University, Department of Orthopaedics, Eksjö, Region Jönköping County and Department of Biomedical Clinical Science von Walden, Ferdinand; Karolinska Institutet,</p> <p>Fornander, Lotta; Linköping University Faculty of Medicine, Department of Orthopaedics, Vrinnevi Hospital</p> <p>Rämö, Lasse; University of Helsinki, Orthopaedics and traumatology</p> <p>Schmidt, Andrew; Hennepin Healthcare, Department of orthopaedics</p> <p>Schilcher, Jörg; Linköping University Hospital, Department of Orthopaedics and Department of Biomedical and Clinical Sciences, Faculty of Health Science</p>
<b>Primary Subject Heading</b>:	Surgery
Secondary Subject Heading:	Emergency medicine
Keywords:	Musculoskeletal disorders < ORTHOPAEDIC & TRAUMA SURGERY, ORTHOPAEDIC & TRAUMA SURGERY, Adult orthopaedics < ORTHOPAEDIC & TRAUMA SURGERY

SCHOLARONE™  
Manuscripts

1 BioFACTS - Biomarkers of rhabdomyolysis in the diagnosis  
2 of acute compartment syndrome: protocol for a prospective  
3 multinational, multi-centre study involving patients with tibial  
4 fractures

5  
6 Abraham Nilsson<sup>1</sup>, Thomas Ibounig<sup>2</sup>, Johan Lyth<sup>3</sup>, Björn Alkner<sup>4</sup>, Ferdinand von Walden<sup>5</sup>,  
7 Lotta Fornander<sup>6</sup>, Lasse Rämö<sup>2</sup>, Andrew Schmidt<sup>7</sup> and Jörg Schilcher<sup>1, 8</sup>

8  
9 <sup>1</sup> Department of Orthopaedic Surgery and Department of Biomedical and Clinical Sciences, University  
10 Hospital Linköping, Linköping University, Linköping, Sweden

11 <sup>2</sup> Department of Orthopaedics and Traumatology, Helsinki University Hospital and University of  
12 Helsinki, Finland

13 <sup>3</sup> Department of Health, Medicine and Caring Sciences, Linköping University, Linköping, Sweden

14 <sup>4</sup> Department of Orthopaedics, Eksjö, Region Jönköping County and Department of Biomedical and  
15 Clinical Sciences, Linköping University, Linköping, Sweden

16 <sup>5</sup> Department of Women's and Children's Health, Karolinska Institutet, Stockholm, Sweden

17 <sup>6</sup> Department of Orthopaedic Surgery in Norrköping and Department of Biomedical and Clinical  
18 Sciences, Linköping University, Norrköping, Sweden

19 <sup>7</sup> Department of Orthopaedics, Hennepin Healthcare, Minneapolis, MN, USA

20 <sup>8</sup> Wallenberg Centre for Molecular Medicine, Linköping University, Linköping, Sweden

21  
22 Abraham Nilsson, (abraham.nilsson@regionostergotland.se)  
23 Thomas Ibounig, (thomas.ibounig@hus.fi)  
24 Johan Lyth, (johan.lyth@liu.se)

25 Björn Alkner, (bjorn.alkner@liu.se)  
26 Ferdinand von Walden, (ferdinand.von.walden@ki.se)  
27 Lotta Fornander, (lotta.fornander@liu.se)  
28 Lasse Rämö, (lasse.ramo@hus.fi)  
29 Andrew Schmidt, (andrew.schmidt@hcmcd.org)  
30 Jörg Schilcher, (jorg.schilcher@liu.se)\*

31 **\*Corresponding author**

32

33 **Word count: 2686**

34

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**ABSTRACT**

**Introduction**

The ischemic pain of acute compartment syndrome (ACS) can be difficult to discriminate from the pain linked to an associated fracture. Lacking objective measures, the decision to perform fasciotomy is based on clinical findings and performed at a low level of suspicion. Biomarkers of muscle cell damage may help to identify and monitor patients at risk, similar to current routines for patients with acute myocardial infarction. This study will test the hypothesis that biomarkers of muscle cell damage can predict ACS in patients with tibial fractures.

**Methods and analysis**

Patients aged 15–65 years who have suffered a tibial fracture will be included. Plasma (P)-myoglobin and P-creatine phosphokinase (P-CK) will be analysed at 6-hourly intervals after admission to the hospital and – if applicable – after surgical fixation or fasciotomy. In addition, if ACS is suspected, blood samples will be collected at 6-hourly intervals. An independent expert panel will assess retrospectively the study data and will classify those patients who had undergone fasciotomy into those with ACS and those without ACS. All primary comparisons will be performed between fracture patients with and without ACS. The area under the receiver operator characteristics curves will be used to identify the success of the biomarkers in discriminating between fracture patients who develop ACS and those who do not. Logistic regression analyses will be used to assess the discriminative abilities of the biomarkers to predict ACS corrected for pre-specified covariates.

**Ethics and dissemination**

59 The study has been approved by the Regional Ethical Review Boards in Linköping  
60 (2017/514-31) and Helsinki/Uusimaa (HUS/2500/2000) and has been registered at  
61 *Clinicaltrials.gov* (Identifier: NCT04674592). The BioFACTS study will be reported in  
62 accordance with the Strengthening the Reporting of Observational Studies in Epidemiology  
63 recommendations.

64

#### 65 **Registration**

66 The study has been registered at *Clinicaltrials.gov* (Identifier: NCT04674592).

67

#### 68 **Keywords**

69 Musculoskeletal disorders, orthopaedic & trauma surgery, adult orthopaedics

**STRENGTHS AND LIMITATIONS OF THIS STUDY**

- The first prospective study to investigate plasma levels of P-myoglobin and P-CK in patients with tibial fractures with and without ACS.
- Multiple participating centres, including different hospital categories from two Nordic countries.
- Expert panel assessment of ACS.

**INTRODUCTION**

Acute compartment syndrome (ACS) is a serious complication in trauma. As the condition is difficult to diagnose and the consequences of a missed compartment syndrome are detrimental, decompressive fasciotomy is performed when there is a low level of suspicion, possibly often leading to unnecessary surgeries.[1]

ACS is caused by either a volume expansion of the muscles, e.g., muscular swelling, or a decrease in the compartment space, e.g., burn injuries, leading to increased intra-compartmental pressure. The increased pressure, in turn, leads to decreased perfusion and, if left untreated, tissue necrosis.[2] ACS can emerge in a wide variety of locations, although most commonly it affects the lower leg due to its tight inelastic fasciae.[3] Tibial fracture is the most common injury associated with ACS, and ACS develops in 2%–8% of the cases.[2, 4] The cardinal symptom of ACS is pain, which can be very difficult to differentiate from the pain caused by a fracture. The treatment for ACS is urgent surgical decompression of the affected muscle compartments through fasciotomy. However, fasciotomy involves a large incision of the skin and compromises the soft tissue envelope around the fractured bone. Therefore, fasciotomy has a profound negative impact on the possibilities for orthopaedic treatment of the fracture, increases the risk for complications, prolongs hospital stays, and

drives up costs.[5-7] Therefore, correct and timely diagnosis is of the outmost importance for these patients.

Currently, ACS is diagnosed using a combination of physical findings and intra-compartmental pressure measurements. Each of these measures has inherent drawbacks in terms of making the correct diagnosis.[1, 8, 9] Specific pressure thresholds at which fasciotomy should be performed have been proposed.[9, 10] However, studies have shown that up to 84% of patients with a tibial fracture exceed this pressure threshold without developing ACS.[11] A recent study has shown that perfusion pressure has low specificity,[12] and, furthermore, the pressure varies with distance to the fracture, making the interpretation even more difficult.[9] Inappropriate use of the method may, therefore, lead to unnecessary fasciotomies. Other promising diagnostic modalities include near-infrared spectroscopy (NIRS). Decreased tissue oxygenation levels correlate with increased intramuscular pressure.[13] NIRS can detect a sudden decrease in tissue oxygenation in patients with ACS,[14] although the reliability of NIRS in an injured leg remains uncertain and its role in the diagnosis of ACS has not been defined.[15] Biomarkers, including measurements of pH and intramuscular glucose might allow the identification of patients with impaired muscle metabolism due to ACS. Also, circulating microRNAs might be a potential tool for the future.[16] However, none of these techniques is used in clinical routine today. In the absence of good objective measures and a clear definition of when ACS is present, decision-making regarding fasciotomy relies on the judgment of the individual doctor, leading to significant variability (2%–24%) in the percentage of fasciotomies performed for ACS per surgeon in fracture patients.[17]

Similar to the diagnosis of acute myocardial infarction, whereby heart muscle-specific Troponin T is measured, markers of muscle cell damage (e.g., P-myoglobin) may be used to diagnose objectively or to monitor ACS. The use of such markers has previously been deemed

1  
2  
3 119 unfeasible on the basis that pathological levels of these markers could be attributed to sources  
4  
5 120 other than the actual muscle compartment, e.g., traumatic muscle damage and heart  
6  
7 121 contusion.[18] Nonetheless, some studies have shown the potential of biomarkers to improve  
8  
9 122 the diagnosis of ACS,[19, 20] albeit never in the presence of a fracture.  
10  
11  
12 123 We have recently shown that high intramuscular pressure coincides with high P-myoglobin  
13  
14 124 levels and that myoglobin may be a relevant, yet unexplored diagnostic tool in ACS  
15  
16  
17 125 associated with trauma.[21]  
18  
19 126  
20  
21  
22

23  
24 127 **Hypothesis**

25  
26 128 P-myoglobin and P-CK can be used to predict ACS in patients with traumatic tibial fractures.  
27  
28 129  
29

30 130 **Aims**

31 131 *Primary aim*

32  
33 132 To describe the diagnostic performances of P-myoglobin and P-CK to predict ACS in patients  
34  
35 133 with traumatic tibial fractures.  
36  
37 134

38  
39 135 *Secondary aim*

40  
41 136 To compare the pathological changes in circulating microRNA (miRNA) and muscle biopsies  
42  
43 137 with the levels of P-myoglobin and P-CK in fracture patients.  
44  
45 138

46  
47 139 To compare changes in P-myoglobin and P-CK between fracture patients with ACS and non-  
48  
49 fracture patients with ACS.  
50  
51 140

52  
53 141 **METHODS AND ANALYSIS**

54  
55 142 The study is a prospective, multi-national, multi-centre study. The study is currently running  
56  
57 143 in Sweden at Linköping University Hospital, Vrinnevi Hospital in Norrköping,  
58  
59  
60



Högländssjukhuset in Eksjö and Kalmar Hospital, as well as in Finland at Töölö Hospital in Helsinki. Additional hospitals may be included in the future.

## Study Population

Patients in the age range of 15–65 years who have suffered traumatic fractures of the tibia will be included. Exclusion criteria are listed in Table 1.

Table 1. Inclusion and exclusion criteria.

<i>Inclusion criteria</i>	<i>Exclusion criteria</i>
Traumatic tibial fracture*	Malignancy
15–65 years old	Acute myocardial infarction
	Kidney failure (GFR $\leq$ 35 ml/min)
	Muscle disease
	Paraplegia/tetraplegia

\*Any anatomical location of the tibia, excluding solitary ankle fractures.

## Outcome measures

### Primary

Myoglobin is an important myocyte-produced compound that is released into the bloodstream as the first enzyme to show increased levels following any type of muscle injury. Myoglobin becomes measurable when the protein binding capacity is exceeded after 1–3 hours. It peaks after 8–12 hours, and it is cleared from the plasma within 24 hours.[22]

The levels of CK in plasma increase within 12 hours of muscle injury, peak after 24–36 hours, and decrease at a rate of 30%–40% per day.[22] Both P-myoglobin and P-CK are routinely measured in the diagnosis and monitoring of rhabdomyolysis.[22] Therefore, facilities for the

162 analysis of these enzymes are available in most clinical settings. The analyses are relatively  
163 cheap (roughly €30 per sample) and can be performed in less than 1 hour.  
164 For the present study, we chose a combination of these enzymes and a 6-hourly interval over a  
165 24-hour or 48-hour period so as to be able to detect the post-injury peak of at least one of  
166 these enzymes (Figure 1). Considering the urgency of the diagnosis, we will primarily focus  
167 on an increase in the level of myoglobin as an early sign of muscle injury.

168

### 169 *Secondary*

170 We will collect blood samples for later analysis of muscle-specific miRNA at the same time  
171 intervals as for the primary outcome. Due to the rapidly evolving field of miRNA research,  
172 we abstain to prespecify specific miRNA's that will be used as objective measures of muscle  
173 damage.[23] At the time of surgery (internal fixation or fasciotomy), biopsies are taken at  
174 some centres for further histological analyses. Two biopsies are taken from the tibialis  
175 anterior muscle in the fractured leg, one close to the injury and one at a distance. One biopsy  
176 is taken from the same muscle in the uninjured leg (control). Biopsies are frozen within 30  
177 minutes using liquid nitrogen-cooled isopentane.[24] For storage, the samples are kept at -  
178 80°C. Since P-myoglobin and P-CK can be affected by renal function,[25] we will analyse the  
179 level of serum creatine and the glomerular filtration rate (GFR). As sex, body weight and age  
180 can affect muscle volume, these parameters will also be registered. We will record the  
181 mechanism of injury (high or low energy), type of fracture (proximal, mid-shaft or distal and  
182 AO/OTA classification), trauma severity (solitary tibial fracture (with or without fibula  
183 fracture), tibial fracture with concomitant long bone fracture, tibial fracture in combination  
184 with multi-trauma), peri-operative findings of muscle viability (colour, consistency,  
185 contractility, capacity to bleed), and whether the muscle bulges at the point of incision.  
186 Stratified analyses for these subgroups will be performed if feasible.

187  
188 *Timing of blood samples*  
189 Plasma for assaying the levels of myoglobin and CK will be collected at those time-points at  
190 which the patient is at high risk to develop ACS (Figure 1):

- 191 1) After the trauma, at admission to hospital: at 6-hourly intervals for a maximum of 48  
192 hours or until definitive surgical fixation.
  - 193 2) After definitive surgical fracture treatment: at 6-hourly intervals for 24 hours.
  - 194 3) If there is suspicion of ACS, blood samples will be collected at 6-hourly intervals until  
195 fasciotomy is performed or the suspicion is dismissed, although no longer than 48 hours.
  - 196 4) After fasciotomy, blood sampling will be continued at 6-hourly intervals for 24 hours.
- 197 Fasciotomy will be performed according to clinical routine praxis, as deemed feasible by the  
198 responsible surgeon.

## 199 200 **Exploratory analyses**

201 We recruit *non-fracture patients* with ACS of the lower leg to enable exploratory  
202 comparisons with *fracture patients* with ACS. Non-fracture patients will undergo the same  
203 sampling algorithm as fracture patients (Figure 1). Patients with malignancy, acute  
204 myocardial infarction, kidney failure ( $\text{GFR} \leq 35 \text{ ml/min}$ ), muscle disease, acute myocardial  
205 infarction, muscle disease, paraplegia/tetraplegia, any acute fracture or acute vascular events,  
206 will be excluded.

## 207 208 **Blood sample analysis**

209 The levels of P-myoglobin, P-CK and S-creatinine in patients at Linköping, Norrköping and  
210 Helsinki will be analysed at the respective hospital. Blood samples from Eksjö and Kalmar  
211 will be centrifuged, stored refrigerated, and sent to Linköping for analysis. EDTA-tubes (for

1  
2  
3 212 analysis of miRNA) will be centrifuged at 2,500 rpm at room temperature for 10 minutes at  
4  
5 213 each hospital. In the Swedish centres, the supernatant will be immediately and carefully  
6  
7 214 removed, frozen in cryo-tubes, and sent to and stored at -80°C until further processing. In  
8  
9  
10 215 Helsinki, the samples will be stored at the premises of the Helsinki Biobank prior to further  
11  
12 216 analysis.  
13

14 217  
15  
16  
17 218 **Expert panel assessment of ACS in patients with fasciotomy**  
18  
19 219 Once collected, the data will be anonymised, uniformly compiled and reviewed by an  
20  
21 220 independent expert panel of senior orthopaedic surgeons. The panel will retrospectively assess  
22  
23 221 the clinical data (injury characteristics, radiographs, pain medication administered, details as  
24  
25 222 to the surgical procedures). Thereafter, the panel will:  
26  
27  
28 223 1) Specify whether or not the patient had an ACS;  
29  
30 224 2) Indicate the two most important factors contributing to that decision; and  
31  
32 225 3) Determine if there are any missing data that might have influenced their decision.  
33  
34 226 The numbers of patients with ACS adjudged by the expert panel will be compared with the  
35  
36 227 numbers of patients undergoing fasciotomy for ACS.  
37  
38

39 228 **Follow-up**  
40  
41  
42 229 Patients will be followed clinically with individualised intervals according to routine care and  
43  
44 230 at 1 year. The 1-year follow-up will include completion of the Lower Extremity Functional  
45  
46 231 Score (LEFS) questionnaire and a clinical examination. The findings will be used for  
47  
48 232 descriptive purposes, in particular to detect patients who present with functional deficits that  
49  
50 233 might be related to an undetected ACS. In all fasciotomized patients and those with suspected  
51  
52 234 ACS-related functional impairment found during the clinical examination, we aim to perform  
53  
54 235 bilateral MRI examinations of the lower leg to explore the possibility of MRI as an objective  
55  
56 236 measure of ACS induced tissue damage.  
57  
58  
59  
60

237

**238 Sample size calculation and statistical analysis plan**

239 We have performed two separate sample size calculations. The first calculation is based on  
240 pilot studies on patients with tibial fractures and patients with tibial fractures complicated by  
241 fasciotomy due to suspected ACS. The mean pre-operative values for P-myoglobin were 289  
242 and 1449 µg/L, respectively, with standard deviations of 249 and 1044 µg/L, respectively.  
243 This corresponds to an effect size of 1.1 if we are conservative and use the largest standard  
244 deviation of 1044 µg/L from both groups. Using this effect size of 1.1 with a 2-sided test and  
245 alpha of 0.05, we need a sample size of 14 fracture patients undergoing fasciotomy due to  
246 suspected ACS to achieve 80% power. If we expect a fasciotomy prevalence of 5% of patients  
247 with tibial fractures, the expected number of non-fasciotomised patients is 266. In the second  
248 calculation, we assumed an improvement of the area under the curve (AUC) value from 0.5  
249 (as good as chance) to 0.7 (acceptable diagnostic accuracy [26]), with an alpha of 0.05, a  
250 power of 80% and ACS prevalence of 5% in the population. Under these assumptions, we  
251 require 16 fracture patients with fasciotomy due to suspected ACS and 311 non-fasciotomised  
252 patients. With 3–5 patients with fasciotomies due to suspected ACS presenting within the  
253 current study network every year, we estimate an inclusion period of 3 years. Inclusion will  
254 continue until 16 patients with tibial fractures and ACS have been recruited, irrespective of  
255 the total number of recruited patients.

256 The difference in P-myoglobin levels between patients with fractures and suspected ACS, and  
257 non-fasciotomised fracture patients will be calculated with an independent samples Welch's t-  
258 test assuming different standard deviations. The AUC will be used to identify the success of  
259 these biomarkers to discriminate fracture patients with ACS from those without.

260

1  
2  
3 261 Logistic regression analyses will be used to assess the discriminative ability of a combination  
4  
5 262 of the two biomarkers to predict ACS, using correction for the following covariates: GFR;  
6  
7 263 sex; body weight; age; trauma mechanism (high or low energy); fracture type (proximal, mid-  
8  
9 264 shaft or distal and AO/OTA classification); and trauma severity (solitary tibial fracture (with  
10  
11 265 or without fibula fracture), tibial fracture with concomitant long bone fracture, tibial fracture  
12  
13 266 in combination with multi-trauma). Due to the limited sample size, we decide to use Leave-  
14  
15  
16  
17 267 one-out cross-validation (LOOCV) for internal validation, which corresponds to analysis type  
18  
19 268 1b in the TRIPOD checklist.[27] A professional statistician (blinded to the clinical  
20  
21 269 parameters) will perform the statistical analyses. All data will be anonymised and stored in  
22  
23  
24 270 secure servers within Region Östergötland, Sweden.

25  
26 271  
27  
28 272 All primary comparisons will be performed between fracture patients with and without ACS.  
29  
30 273 Exploratory analysis will be performed between patients with suspected ACS, with or without  
31  
32 274 a fracture. Separate statistical analyses will be performed for those patients who have  
33  
34 275 undergone fasciotomies due to suspected ACS and those deemed as true ACS cases by the  
35  
36 276 expert panel. When feasible, stratified subgroup analyses based on the abovementioned  
37  
38  
39 277 covariates will be performed.

40  
41  
42 278  
43  
44 279 **Trial status**

45  
46 280 Patient recruitment started at Linköping University Hospital on April 1<sup>st</sup>, 2018. September 1<sup>st</sup>,  
47  
48 281 2019 a final study protocol was implemented at Linköping University Hospital and in  
49  
50 282 subsequent months at Höglandssjukhuset district hospital Eksjö, Vrinnevi Hospital in  
51  
52 283 Norrköping, and Kalmar Hospital. In January 2021, Helsinki University Hospital, Finland  
53  
54 284 started to recruit patients. Currently, approximately 200 patients have been included.

55  
56 285  
57  
58  
59  
60

286 *Study time schedule*

287 2021: Preliminary analysis of blood samples, muscle biopsies and miRNA to assess the  
288 quality levels of these samples.

289 2022: 300 patients included.

290 2023: Data analysis and manuscript writing.

291

## 292 **Patient and public involvement**

293 Neither patients nor members of the public are involved in the design, conduct, reporting, or  
294 dissemination plans of this study.

295

296

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

**ETHICS AND DISSEMINATION**

The study was approved by the Regional Ethical Review board in Linköping (Dnr. 2017/514-31) and the Ethical Review Board of the Hospital District of Helsinki and Uusimaa (HUS/2500/2000) and has been registered at *Clinicaltrials.gov* (Identifier: NCT04674592). Oral and written explanations will be provided to all eligible patients, and written consent will be obtained as soon as possible but no later than during clinical rounds the morning after enrolment. Patients might feel compelled to participate in the study as the treating surgeon usually is the same person that seeks the patient’s consent. This is a problem that cannot be avoided in the context of recruitment performed in emergency situations. As all the samples will be analysed in a blinded fashion, there is no risk that these values will disrupt the clinical decision-making process in the emergency situation. The risk for complications associated with study participation is low, and there is a potential diagnostic benefit. The study will increase the level of awareness and knowledge of medical staff. Therefore, regardless of the results, the implementation of the study will increase patient safety and, thereby, balance out any risks that study participation may entail.

If we will be able to define threshold values of P-myoglobin and P-CK for the detection of ACS with good diagnostic accuracy, these values could be implemented in clinical practice without delay. Specifically, these threshold values would allow the individual surgeon to decide to abstain from fasciotomy and instead observe the patient and follow the biomarker dynamics.



## **Acknowledgements**

To patients and personnel at all study sites for their invaluable contributions. Stefan Wetterstad for assistance in study organisation and patient recruitment at Kalmar Hospital, Sweden.

## **Author contributions**

JS conceptualised the study idea.  
AN and JS planned and implemented the study at all the study sites.  
TI and LR planned and implemented the study at the Helsinki University Hospital.  
JL, AN and JS planned the statistical analysis.  
BA provided advice on biomarker handling and analysis and planned and implemented the study at Eksjö Hospital.  
FW provided advice on biomarker handling and analysis.  
LF planned and implemented the study at Norrköping hospital.  
AS contributed to the overall design of the study.  
All the authors contributed to the design of the study and revised and approved the final manuscript.

## **Funding**

This work was supported by Linköping University, Region Östergötland (grant no. RÖ-969530, RÖ-941258, RÖ-892721, RÖ-785751), the Medical Research Council of Southeast Sweden (grant no. FORSS-852501, FORSS-939750, FORSS-706601 and RALF-0600911), the Wallenberg Foundation, Futurum – the Academy for Health and Care, Region Jönköping

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

345 County, Sweden (grant no. FUTURUM-937508 and 870471) and Finnish state funding for  
346 university-level health research.  
347

For peer review only

## Availability of data and material

The datasets generated and/or analysed during the current study may be obtained from the corresponding author on reasonable request.

## Competing interests

The authors declare that they have no competing interests.

## FULL REFERENCES

1. Schmidt AH. Continuous compartment pressure monitoring-better than clinical assessment? *J Bone Joint Surg Am*. 2013;95(8):e52.
2. Schmidt AH. Acute compartment syndrome. *Injury*. 2017;48 Suppl 1:S22-S5.
3. Matsen FA, 3rd, Krugmire RB, Jr. Compartmental syndromes. *Surg Gynecol Obstet*. 1978;147(6):943-9.
4. Park S, Ahn J, Gee AO, Kuntz AF, Esterhai JL. Compartment syndrome in tibial fractures. *J Orthop Trauma*. 2009;23(7):514-8.
5. Dover M, Memon AR, Marafi H, Kelly G, Quinlan JF. Factors associated with persistent sequelae after fasciotomy for acute compartment syndrome. *J Orthop Surg (Hong Kong)*. 2012;20(3):312-5.
6. Schmidt AH. The impact of compartment syndrome on hospital length of stay and charges among adult patients admitted with a fracture of the tibia. *J Orthop Trauma*. 2011;25(6):355-7.
7. Lollo L, Grabinsky A. Clinical and functional outcomes of acute lower extremity compartment syndrome at a Major Trauma Hospital. *Int J Crit Illn Inj Sci*. 2016;6(3):133-42.
8. Shuler FD, Dietz MJ. Physicians' ability to manually detect isolated elevations in leg intracompartmental pressure. *J Bone Joint Surg Am*. 2010;92(2):361-7.
9. Heckman MM, Whitesides TE, Jr., Grewe SR, Rooks MD. Compartment pressure in association with closed tibial fractures. The relationship between tissue pressure, compartment, and the distance from the site of the fracture. *J Bone Joint Surg Am*. 1994;76(9):1285-92.
10. McQueen MM, Court-Brown CM. Compartment monitoring in tibial fractures. The pressure threshold for decompression. *J Bone Joint Surg Br*. 1996;78(1):99-104.
11. Prayson MJ, Chen JL, Hampers D, Vogt M, Fenwick J, Meredick R. Baseline compartment pressure measurements in isolated lower extremity fractures without clinical compartment syndrome. *J Trauma*. 2006;60(5):1037-40.
12. Schmidt AH, Di J, Zipunnikov V, Frey KP, Scharfstein DO, O'Toole RV, et al. Perfusion Pressure Lacks Diagnostic Specificity for the Diagnosis of Acute Compartment Syndrome. *J Orthop Trauma*. 2020;34(6):287-93.
13. Shuler MS, Reisman WM, Kinsey TL, Whitesides TE, Jr., Hammerberg EM, Davila MG, et al. Correlation between muscle oxygenation and compartment pressures in acute compartment syndrome of the leg. *J Bone Joint Surg Am*. 2010;92(4):863-70.
14. Shuler MS, Reisman WM, Cole AL, Whitesides TE, Jr., Moore TJ. Near-infrared spectroscopy in acute compartment syndrome: Case report. *Injury*. 2011;42(12):1506-8.
15. Schmidt AH, Bosse MJ, Obrebsky WT, O'Toole RV, Carroll EA, Stinner DJ, et al. Continuous Near-Infrared Spectroscopy Demonstrates Limitations in Monitoring the Development of

Acute Compartment Syndrome in Patients with Leg Injuries. *J Bone Joint Surg Am*. 2018;100(19):1645-52.

16. Etheridge A, Lee I, Hood L, Galas D, Wang K. Extracellular microRNA: a new source of biomarkers. *Mutat Res*. 2011;717(1-2):85-90.

17. O'Toole RV, Whitney A, Merchant N, Hui E, Higgins J, Kim TT, et al. Variation in diagnosis of compartment syndrome by surgeons treating tibial shaft fractures. *J Trauma*. 2009;67(4):735-41.

18. Shadgan B, Menon M, O'Brien PJ, Reid WD. Diagnostic techniques in acute compartment syndrome of the leg. *J Orthop Trauma*. 2008;22(8):581-7.

19. Valdez C, Schroeder E, Amdur R, Pascual J, Sarani B. Serum creatine kinase levels are associated with extremity compartment syndrome. *J Trauma Acute Care Surg*. 2013;74(2):441-5; discussion 5-7.

20. Hefler F. Compartment syndrome after gynecologic laparoscopy: systematic review of the literature and establishment of normal values for postoperative serum creatine kinase and myoglobin levels. *Arch Gynecol Obstet*. 2017;296.2:285-93.

21. Nilsson A, Alkner B, Wetterlov P, Wetterstad S, Palm L, Schilcher J. Low compartment pressure and myoglobin levels in tibial fractures with suspected acute compartment syndrome. *BMC Musculoskelet Disord*. 2019;20(1):15.

22. Giannoglou GD, Chatzizisis YS, Misirli G. The syndrome of rhabdomyolysis: Pathophysiology and diagnosis. *Eur J Intern Med*. 2007;18(2):90-100.

23. Siracusa J, Koulmann N, Bourdon S, Goriot ME, Banzet S. Circulating miRNAs as Biomarkers of Acute Muscle Damage in Rats. *Am J Pathol*. 2016;186(5):1313-27.

24. Dubowitz V, A. Sewry C, Oldfors A. Muscle biopsy: a practical approach: expert consult; online and print.: Elsevier Health Sciences; 2013. 8-10 p.

25. Hällgren R, Karlsson FA, Roxin LE, Venge P. Myoglobin turnover--influence of renal and extrarenal factors. *Journal of Laboratory and Clinical Medicine*. 1978;91(2).

26. Mandrekar JN. Receiver Operating Characteristic Curve in Diagnostic Test Assessment. *Journal of Thoracic Oncology*. 2010;5(9):1315-6.

27. Collins GS, Reitsma JB, Altman DG, Moons KG. Transparent reporting of a multivariable prediction model for Individual Prognosis or Diagnosis (TRIPOD): the TRIPOD statement. *J Clin Epidemiol*. 2015;68(2):134-43.

## Legends

Figure 1. Flowchart of patient recruitment.

For peer review only

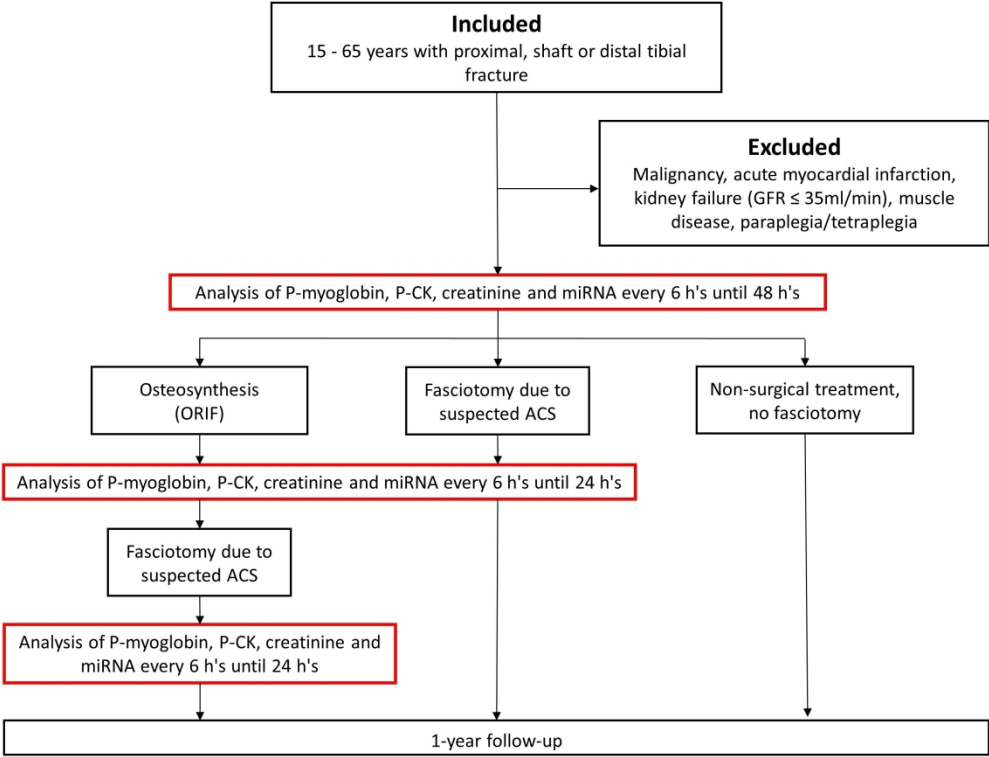


Figure 1. Flowchart of patient recruitment.

249x249mm (300 x 300 DPI)