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Neuroimmune responses following joint mobilisation and manipulation in people with persistent neck pain: A protocol for a randomised placebo-controlled trial

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2021-055748
Article Type:	Protocol
Date Submitted by the Author:	22-Jul-2021
Complete List of Authors:	Lutke Schipholt, Ivo J; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences; Amsterdam UMC Locatie VUmc, Department of Clinical Chemistry, Laboratory Medical Immunology Scholten-Peeters, Gwendolijne ; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences Bontkes, Hetty; Amsterdam UMC VUMC Site, Department of Clinical Chemistry, Laboratory Medical Immunology Coppieters, Michel ; Griffith University Menzies Health Institute Queensland; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences
Keywords:	IMMUNOLOGY, Musculoskeletal disorders < ORTHOPAEDIC & TRAUMA SURGERY, Spine < ORTHOPAEDIC & TRAUMA SURGERY, REHABILITATION MEDICINE, CLINICAL PHYSIOLOGY

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Manuscripts

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3 **Neuroimmune responses following joint mobilisation and manipulation in people with**
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5 **persistent neck pain: A protocol for a randomised placebo-controlled trial**
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Key words: Neuroimmune, musculoskeletal physiotherapy, pain, cytokine, immune system, manual therapy, non-pharmacological treatment, neck pain.

Contributorship statement

All authors were involved in the design of the study and the acquisition of the research funds. ILS drafted the initial versions of the article. All authors critically revised the various drafts and approved the final version.

Competing interests

The authors have no known conflict of interest and have no commercial interest in this study.

Funding

This study is funded by the Dutch Association for Manual Therapy (NVMT, grant ID. Top-down_2018) and by the Faculty of Behavioural and Movement Sciences (grant ID. Lab Fund_2019) of Vrije Universiteit Amsterdam. The MSG Science Network (<https://www.msg-sciencenetwerk.nl/>) (grant ID. N/A) will support participant recruitment.

Data sharing statement

Data are available upon reasonable request.

Word count 3536/4000

ABSTRACT

Introduction: Joint mobilisation and manipulation often results in immediate pain relief in people with neck pain. However, the biological mechanisms behind pain relief are largely unknown. There is preliminary evidence that joint mobilisation and manipulation lessens the upregulated neuroimmune responses in people with persistent neck pain.

Methods and analysis: This study protocol describes a randomised placebo-controlled trial to investigate whether joint mobilisation and manipulation influence neuroimmune responses in people with persistent neck pain. People with persistent neck pain (N=100) will be allocated, in a randomised and concealed manner, to the experimental or control group (ratio 3:1).

Short-term (i.e., baseline, immediately after and two-hours after the intervention) neuroimmune responses will be assessed, such as inflammatory marker concentration following *in-vitro* stimulation of whole blood cells, systemic inflammatory marker concentrations directly from blood samples, phenotypic analysis of peripheral blood mononuclear cells, and serum cortisol. Participants assigned to the experimental group (N=75) will receive cervical mobilisations targeting the painful and/or restricted cervical segments and a distraction manipulation of the cervico-thoracic junction. Participants assigned to the control group (N=25) will receive a placebo mobilisation and placebo manipulation. Using linear mixed models, the short-term neuroimmune responses will be compared 1) between people in the experimental and control group, 2) within the experimental group, between people who experience a good outcome and those with a poor outcome. Furthermore, the association between the short-term neuroimmune responses and pain relief following joint mobilisation and manipulation will be tested in the experimental group.

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3 **Ethics and dissemination:** This trial is approved by the Medical Ethics Committee of
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5 Amsterdam University Medical Centre, location VUmc (Approval number: 2018.181).
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7 **Trial registration number and status:** The study protocol is registered at trialregister.nl with
8
9 study ID: NL6575 registered on 18-01-2018; Recruitment commenced 26 February 2019. All
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11 data are anticipated to be collected by January 2022, when data analysis and interpretation are
12
13 anticipated to commence.
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19 **Strengths and limitations of this study**

- 21 ❖ This study provides insight in the interplay between joint mobilisation and
22 manipulation, neuroimmune responses, and pain relief in people with persistent neck
23 pain.
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- 26 ❖ By adding a placebo-control group, possible working mechanisms of joint
27 mobilisation and manipulation on neuroimmune responses may be revealed.
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- 30 ❖ The interventions will be delivered by two musculoskeletal physiotherapists, which
31 may limit the generalisability.
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- 34 ❖ Due to the small control group, it is not feasible to divide the control participants
35 according to outcome.
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1. INTRODUCTION

The disruption of the bidirectional communication pathways between the central nervous system and the immune system may play an important role in persistent pain.(1) Over the last two decades, it has become apparent that neuroimmune crosstalk is present in musculoskeletal pain, and may play a mediating role in the transition from acute to persistent pain.(1) For people with persistent neck pain, aberrant neuroimmune responses may be present, such as systemically elevated levels of inflammatory markers.(2, 3) These increased neuroimmune responses may be relevant to understand and manage persistent spinal pain.(3) A growing body of literature suggests that these neuroimmune responses are associated with pain intensity,(4-6) disability(7) and recovery,(8) and can be influenced by musculoskeletal physiotherapy, such as joint mobilisation and manipulation,(9-11) nerve mobilisation(12, 13) and exercise.(14-16)

Several meta-analyses indicate that musculoskeletal physiotherapy for people with spinal pain may provide immediately pain relief and improvements in functional activities compared to no treatment, placebo or other treatments.(17-19) Nevertheless, unravelling the mechanism of how joint mobilisation and manipulation results in pain relief remains an area for further investigation.(20, 21) There are various explanations of how joint mobilisation and manipulation might cause pain relief, including neurophysiological,(22, 23) neuromuscular,(20) neuroimmune(24, 25) and non-specific responses.(26)

Recent studies suggest a possible neuroimmune-mediated mechanism of pain relief following joint mobilisation and manipulation.(9-11) For example, a reduction in systemic inflammatory marker concentration directly from blood samples(9, 11) and a reduction in inflammatory marker concentration following *in-vitro* stimulation of whole blood cells(10, 27) were found immediately following the intervention. These studies have however

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3 important methodological limitations, such as inclusion of healthy participants,(27) modest
4 sample sizes,(9, 10) a narrow selection of inflammatory markers,(9-11) lack of correction for
5
6 potential confounding variables,(9, 10, 28) and lack of a placebo-control group.(9, 10)
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10 Therefore, we will conduct an adequately powered, placebo-controlled randomised clinical
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12 trial in people with persistent neck pain, which will evaluate a broad range of inflammatory
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14 markers. The purpose of this paper is to describe the study protocol to investigate the short-
15
16 term effects of joint mobilisation and manipulation on neuroimmune responses in people with
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18 persistent neck pain.
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24 **2. METHODS**

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28 This manuscript followed the guidelines for clinical trial protocols (SPIRIT
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30 statement),(29) for reporting randomised trials (CONSORT statement),(30) and for
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32 intervention description and replication (TIDieR checklist).(31)
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38 **Aim**

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40 The overall aim of this clinical trial is to gain insights in the relation between short-
41
42 term neuroimmune responses following joint mobilisation and manipulation and pain relief in
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44 people with persistent neck pain. The specific aims are: 1) to compare the short-term
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46 neuroimmune responses between the experimental and control group; 2) to compare the short-
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48 term neuroimmune responses of those in the experimental group with a good outcome (i.e.,
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50 immediately pain relief) with those in the experimental group with a poor outcome; and 3) to
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52 assess the association between short-term neuroimmune responses and pain relief in the
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54 experimental group.
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Study design and setting

The study is a placebo-controlled randomised trial with follow-up at three time points:

baseline, immediately, and two-hours and two-days following the intervention (Fig. 1).

Participants will be recruited from GP clinics, primary care physiotherapy practices and outpatient services (neurology and orthopaedic departments) at secondary care hospitals. Data are anticipated to be collected between February 2019 and January 2022, when data analysis and interpretation are anticipated to commence.

Selection criteria

Individuals meeting the following inclusion criteria are eligible to participate: age: 18-65 years; non-specific neck pain for at least six weeks(32) with a minimum pain intensity of 40/100 on a visual analogue scale (VAS), and a sufficient speaking and reading level of the Dutch language to complete the study. Exclusion criteria are contra-indications for cervical mobilisation or cervico-thoracic manipulation,(33, 34) pregnancy or less than 9 months postpartum, contra-indications for venipuncture (e.g., phlebitis), treatment for the current neck pain episode during the preceding two weeks, taken corticosteroids or cytokine modulatory medication (e.g., methotrexate, infliximab) in the preceding 6 weeks, use of botulinum toxin (Botox) injection during the preceding 3 months, non-steroid anti-inflammatory drug medication within the past 7 days (e.g., diclofenac, ibuprofen, naproxen), long-distance flight within the past 7 days, ongoing shift work, having a known comorbid condition with immune/endocrine malfunction (e.g., ankylosing spondylitis, Crohn's disease, sarcoidosis, Cushing syndrome, cancer, diabetes), medical red flags suggestive of serious pathology,(35, 36) and a diagnosed psychological condition (e.g., clinical depression).

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3 Consecutive participants who meet all selection criteria and are willing to participate
4 will be admitted to the study. All participants will provide written informed consent prior to
5 participation. Initial screening for eligibility will be conducted via telephone calls.
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10 11 12 **Randomisation, concealed allocation and blinding**

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14 Block randomisation will be used to allocate participants to the experimental or
15 control group with an allocation ratio of 3:1 (experimental : control). A computer random
16 number generator will create block sizes of 4 and 8 participants. To conceal the allocation
17 sequence, an independent person not involved in the study will assign eligible people to the
18 groups on the day the participant will enrol in the study. Blood samples will be coded to blind
19 the research assistant and laboratory investigators to the study groups. The participant,
20 research assistant and the investigator who includes the participants will be blinded for group
21 assignment. The treating clinicians, research assistant and laboratory investigators will be
22 unaware whether participants experienced a good outcome or not. All laboratory and data
23 analyses will be performed by blinded investigators.
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40 **Interventions**

41 42 ***Experimental intervention***

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44 Spinal mobilisation will consist of low-velocity, low-amplitude mobilisations at the
45 painful cervical segmental levels (Fig. 2 – Panels A-C); spinal manipulation will consist of a
46 high-velocity, low-amplitude distraction manipulation at the cervico-thoracic junction (Fig. 2
47 – Panel D).(37) These techniques aim to restore motion and reduce pain. They are commonly
48 used and are conform to the Dutch guidelines for musculoskeletal physiotherapy for treating
49 neck pain.(35) All interventions will be performed by two musculoskeletal physiotherapists
50 with more than 5 years of relevant clinical experience.
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Cervical mobilisation

Painful and restricted cervical segments will be identified by passive side-bending of the neck targeting each segmental level separately.(38) Reproduction of the participant's pain will be considered to identify the involved level(s). The inter-tester reliability for these tests is fair to substantial.(38, 39)

Depending on the identified painful or restricted spinal levels, the treating clinician may select from different mobilisation techniques: mobilisation targeting the atlanto-axial segment (Fig. 2 – Panel A); segmental zygapophyseal joint mobilisation (C2 to C7) (Fig. 2 – Panel B) and occipital-atlanto-axial joint mobilisation (Fig. 2 – Panel C). Three series of oscillations (~1Hz) will be applied for 30 seconds; with 30 seconds rest in between the series.

Cervico-thoracic junction distraction manipulation

Irrespective of the level of their neck pain, all participants will receive a distraction manipulation of the cervico-thoracic junction (Fig. 2– Panel D).(40) If there is no audible cavitation sound during the first attempt, the manipulation will be repeated once.

Control (placebo) intervention

The control group will receive a placebo mobilisation and placebo manipulation. Procedures, including the instructions, will be identical as for the experimental intervention, except that the clinician will only apply hand contact and no pressure or movement will occur. Participants will be informed that an audible popping sound may or may not occur, and that this sound is not necessary to restore motion and reduce pain.

The credibility of a control intervention can interact with participant expectations in complex ways.(41) To account for differences in intervention expectations, participants will

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3 indicate the extent to which they agree (using a four-point Likert scale) with four statements
4 regarding their intervention expectations (Table 1). These statements will be presented before
5 the delivery of the experimental and control intervention.(42)
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10 Based on the short-term changes in pain intensity score (i.e., immediately and two-
11 hours following the intervention), participants in the experimental group will be categorised
12 into those with a good outcome ($\geq 50\%$ improvement in pain intensity at both time points), a
13 poor outcome ($\leq 20\%$ improvement in pain intensity score at both time points) or an unclear
14 outcome (not fitting the criteria for a good or poor outcome).(43) Based on these cut-off
15 scores, we anticipate to have a minimum of 25 participants in both the good outcome and poor
16 outcome group. If our *a-priori* determined minimum of 25 participants in either group is not
17 achieved, the good outcome group and the poor outcome group will be supplemented with
18 respectively the best responders and poorest responders from the uncertain outcome group in
19 order to obtain 25 participants in both groups.
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35 **Outcomes**

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37 A broad range of neuroimmune responses will be monitored: a) inflammatory marker
38 concentration following *in-vitro* stimulation of whole blood cells, b) systemic inflammatory
39 marker concentrations directly from blood samples, c) phenotypic analysis of peripheral blood
40 mononuclear cells and d) *ex-vivo* serum cortisol (Table 1). To create an inflammatory
41 profile,(44) a range of pro-inflammatory and anti-inflammatory markers will be used. Ex-vivo
42 serum and supernatants after stimulation will be stored at minus 80°C and will be analysed
43 upon completion of data collection. The laboratory methodology and sample handling prior to
44 stimulation will be tightly monitored and reported, because inconsistency in interlaboratory
45 methodology and reporting impairs interpretation, comparability and reproducibility.(45)
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Primary outcomes

The primary outcomes are the short-term (i.e., immediately and two-hours following the intervention) differences in interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α) following *in-vitro* stimulation of whole blood cells. These cytokines will be determined using Meso Scale Discovery (MSD, Maryland United States) at baseline, immediately and two-hours following the intervention. These cytokines are selected because previous research has indicated that those cytokines might play a role in spinal pain.(11, 27, 46-48)

To induce cytokine production, whole blood cultures will be stimulated for 24 hours with lipopolysaccharide (LPS) from *Escherichia coli* O55:B5 (Sigma-Aldrich Chemie GmbH, Schnelldorf, Germany) at a concentration of 1 nanogram LPS/millilitre whole blood (ng/ml) and 10 microgram LPS/millilitre whole blood (μ g/ml) at 37°C in a humidified 5% CO₂ incubator. At baseline (Fig. 1) blood samples for neuroimmune measurements (one sodium heparin vacutainer without gel and one serum vacutainer without gel for each time point) will be drawn between 8:00 and 9:00 AM.(49) The cytokine levels will be determined using a custom-made U-plex MSD and expressed in picogram/millilitre (pg/ml). The entire blood stimulation procedure and MSD will be performed by an experienced laboratory technician at Amsterdam University Medical Centre, location VUmc, Department of Clinical Chemistry, Medical Immunology Laboratory.

Secondary outcomes

Several additional neuroimmune responses will be quantified as secondary outcomes at various time points (Table 1).

The levels of interleukin-1 receptor antagonist (IL-1RA), interleukin-4 (IL-4), interleukin-10 (IL-10), c-c motif chemokine ligand 2 (CCL2), c-c motif chemokine ligand 3

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3 (CCL3) and c-c motif chemokine ligand 4 (CCL4) will be determined following *in-vitro*
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5 stimulation of whole blood cells.
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8 Systemic inflammatory markers directly from blood samples (tumor necrosis factor –
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10 receptor antagonist II (TNF-RII), IL-1 β and IL-1RA) will be measured using multianalyte
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12 assay Ella (R&D systems, Minneapolis, United States) and high-sensitive c-reactive protein
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14 (hsCRP), using Roche/Hitachi cobas c systems (Indianapolis, United States).
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17 Phenotypic analysis of peripheral blood mononuclear cells will be determined. The
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19 absolute number of lymphocyte subsets (NK cells, B-cells, CD4⁺ and CD8⁺ T-cells and
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21 CD25^{hi} regulatory T-cells), monocytes, as well as activation status of these cells, HLA-DR
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23 and TLR-4 expression, will be determined by 10-color flowcytometry (FCM, Gallios Flow
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25 Cytometer, Beckman Coulter, Indianapolis, United States; Analyse software: Kaluza).
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27 Differences between all groups in serum cortisol concentration will be determined using
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29 conventional electrochemiluminescence immunoassay (ECLIA) from Roche (Cobas Cortisol,
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31 2nd generation, Indianapolis, United States) in agreement with the manufacturer's protocol.
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38 **Procedures**

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40 Once consent is obtained, baseline measurements will be taken (Fig. 1). At baseline,
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42 participants will undergo physical tests to determine pain characteristics, physical functioning
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44 and body composition (Table 2 & Table 3). After this, participants will complete an electronic
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46 survey to collect sociodemographic and clinical information (Table 1) and intervention
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48 expectations (Appendix A). Participants will then undergo one venipuncture from the cubital
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50 vein to fill two vacutainers which will be used to quantify the neuroimmune responses (Table
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52 1). Collection of all baseline data will take 30-45 minutes and will take place at the
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54 Amsterdam University Medical Centre, location VUmc, or at a participating primary care
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56 physiotherapy practice, under the supervision of a research assistant.
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3 Participants will then be randomly allocated to the experimental and control group,
4 and treated accordingly. Immediately and two-hours following the intervention, participants
5 will undergo another venipuncture to fill two vacutainers. Between the immediate and two-
6 hours follow-up measures, questionnaires will be completed to collect psychosocial
7 information such as sleep, disability and kinesiophobia (Table 2).
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12 Immediately and two-hours following the intervention, participants will undergo
13 physical tests (Fig. 1) and will rate their pain intensity on a VAS. Two-hours following the
14 intervention, participant will rate their perceived recovery on a 7-point Global Perceived
15 Effect scale (GPE) (Table 2). Two-days following the intervention, participants will receive
16 an electronic survey regarding potential adverse events, GPE and pain intensity. Figure 1
17 shows the planned flow of participants through the study.
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30 **Sample size**

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32 Based on the sample size calculation(50) (longitudinal analysis; three time points
33 (baseline, immediately follow-up, two-hours follow-up) with 80% power to detect a mean
34 difference of 550 (SD 933) for TNF- α levels with a 0.05 two-sided significance level,
35 correlation of 0.6 among repeated measures, ratio between groups of 0.25, a total sample size
36 of 91 is needed.(27) Allowing for a drop-out rate of ~10%, a total sample size of 100
37 participants is required.
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49 **Statistical analyses**

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51 Data will be checked for normality by the Kolmogorov-Smirnov test and visual
52 inspection of Q-Q plots, box plots and histograms. In case of no normality of data, the data
53 will be log transformation. Data will be presented as means with standard deviations unless
54 otherwise noted. For the analyses, statistical significance will be set at $p < 0.05$. Intention-to-
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3 treat analyses using mixed models will be performed to analyse differences between the
4 experimental group and control group. Linear mixed model analyses with fixed factor (time),
5 covariate (group) and interaction (time*group) will be used to detect differences between the
6 groups at the three time points (baseline, immediately follow-up, two-hours follow-up) for
7 TNF- α and IL- β following *in-vitro* stimulation of whole blood cells. A random intercept will
8 be selected to account for the correlated nature of multiple measurements from the same
9 participant. The regression coefficient (B), p-value and confidence intervals (95%CI) will be
10 computed for the crude models, as well as for the adjusted models.(28, 51) Linear regression
11 analysis will be used to test for differences in phenotypic analysis of peripheral blood
12 mononuclear cells and cortisol between the experimental and control group and of those in the
13 experimental group with a good outcome (i.e., immediate pain relief) with those in the
14 experimental group with a poor outcome.

33 **Adverse events**

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35 Serious and non-serious adverse events related to the experimental and control
36 intervention, and all other aspects of the study, will be documented. At the three post-
37 intervention time points, potential adverse events will be recorded using an online survey.
38 Adverse events will be followed-up as needed by an independent clinician. Depending on the
39 nature of the event, participants may be referred to a GP or a medical specialist, and
40 additional tests or procedures may be proposed. The experimental intervention has been
41 shown to be safe(11, 27) and it is considered unlikely that serious adverse events due to the
42 interventions will occur. Therefore, installing a data monitoring safety board was not
43 requested by the Ethics Committee.

58 **Patient and public involvement**

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3 A panel of four people with persistent neck pain co-developed and evaluated the study
4 design, research questions, choice of experimental and control intervention, and burden of
5 study participation for the participants. Two of these people and two representatives from the
6 public reviewed the Patient information letter and their feedback was used to improve the
7 letter.
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17 **Data management and monitoring**

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19 The data will be collected at the Department of Rehabilitation of the Amsterdam
20 University Medical Centre, location VUmc, and/or in physiotherapy practices. The collected
21 data will be securely stored at Vrije Universiteit Amsterdam, Faculty of Behavioural and
22 Movement Sciences. All data are de-identified by using unique participant ID numbers in
23 such a way that the data cannot be traced back to the individual participants without the key.
24 The participants code will exist of a random code of three numbers. The electronically key
25 connecting participant names with codes will be kept in a secure location in the principal
26 investigator's office. The key will be kept for six months after the final publication, and will
27 then be destroyed. Data will be stored in a de-identified manner for fifteen years after the final
28 publication.
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45 **Role of funding source**

46
47 This study is funded by the Dutch Association for Manual Therapy (NVMT, grant ID.
48 Top-down_2018) and by the Faculty of Behavioural and Movement Sciences (grant ID. Lab
49 Fund_2019) of Vrije Universiteit Amsterdam. The MSG Science Network ([https://www.msg-](https://www.msg-sciencenetwerk.nl/)
50 [sciencenetwerk.nl/](https://www.msg-sciencenetwerk.nl/)) (grant ID. N/A) will support participant recruitment. The funding sources
51 have no role in the study design and will not have any roles in data collection, analysis and
52 interpretation of the data, nor in the reporting of the results.
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Ethics and dissemination

The study is registered at trialregister.nl with study identification NL6575. The results of the study will be published in peer-reviewed journals and disseminated at conferences, in newsletters and social media. The trial is approved by the Medical Ethics Committee of Amsterdam University Medical Centre, location VUmc (Approval number: 2018.181). All procedures will be conducted in accordance with the Declaration of Helsinki.(52) Amendment to this protocol will be submitted for approval to the Medical Ethical Committee and deviations from the protocol will be reported to the trial registration.

3. DISCUSSION

There is considerable debate in the literature regarding the possibility of meaningful neuroimmune-mediated pain relief following joint mobilisation and manipulation.(2, 7, 53, 54) We described a protocol for a randomised placebo-controlled study that will assess potential neuroimmune-mediated pain relief following joint mobilisation and manipulation in people with persistent neck pain. The aim of this study is to gain insights in the relation between changes in neuroimmune responses and pain relief, rather than in the clinical efficacy or effectiveness of joint mobilisation and manipulation for people with persistent neck pain.

Recent data suggest that the production of pro-inflammatory cytokines is higher and production of anti-inflammatory cytokines is lower in patients with persistent-pain compared to healthy people following *in-vitro* stimulation of whole blood cells.(44) Additionally, a specific, coordinated inflammatory processes may be important for patient recovery.(3) Contrary to the other studies we are aware of that measured neuroimmune responses following joint mobilisation and manipulation,(7, 44, 55) we will assess a comprehensive

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3 range of inflammatory markers. Our approach to measure pro-inflammatory cytokines and
4 their antagonists provides insight into the activation of immunocompetent cells.(56)
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8 We believe the design of our study allows to assess the specific effects of joint
9 mobilisation and manipulation on neuroimmune responses. For instance, rather than
10 comparing the joint mobilisation and manipulation with a wait-and-see approach, we will
11 compare responses with a placebo-control intervention that resembles joint mobilisation and
12 manipulation. Additionally, the verbal instructions between the experimental and control
13 groups will be comparable and standardised, which reduces differences in intervention
14 efficacy due to non-specific intervention effects.(57) Differences in verbal instructions have
15 been shown to be associated with differences in endocrine responses following joint
16 manipulation in people with neck pain.(58) Finally, we will record the participant's
17 intervention expectations and beliefs regarding joint mobilisation and manipulation as a
18 treatment method to alleviate neck pain.(42)
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33 Previous research revealed a non-linearity of the VAS to measure pain intensity, that
34 responsiveness varies along the spectrum of pain intensity and the importance of taking
35 baseline pain into account when evaluating change scores.(43, 59, 60) Consequently,
36 categorising good, unclear and poor outcome using raw data, or change scores in general, are
37 invalid as these will either underestimate or overestimate true change.(60) To overcome this
38 problem, we follow the initiative on methods, measurement and pain assessment in clinical
39 trials (IMMPACT) recommendation to identify those with a good, poor outcome, or unclear
40 outcome.(43)
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51 Besides the strengths, the proposed study has some potential limitations. First, we
52 assume a linear association between neuroimmune responses and musculoskeletal pain. A
53 linear association between neuroimmune responses and musculoskeletal pain is a prerequisite
54 for the justification of the statistics proposed in this protocol. However, one study suggests
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3 that an initial threshold of neuroimmune responses might be required, which would suggest a
4 non-linear relationship between neuroimmune responses and musculoskeletal pain.(61) In that
5 study, elevated IL-6 levels were only present in the group of people with pain > 40/100 VAS
6 compared to control.(61) Therefore, a minimal pain intensity of 40/100 on the VAS will be a
7 prerequisite for participating in this study.
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14 Another limitation is that only a single session of joint mobilisation and manipulation
15 will be provided together with a short follow-up. While a single session of joint mobilisation
16 and manipulation may induce a pain-relieving effect,(17) the clinical relevance of
17 immediately pain relief is unclear. Nonetheless, our aim is not to examine the efficacy of joint
18 mobilisation and manipulation but rather to understand the biological mechanisms behind
19 pain relief following joint mobilisation and manipulation. In studying the mechanism of
20 action, a short follow-up has the advantage that potential confounding variables can be
21 controlled, such as food intake, stress, physical exercise and health status.
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35 **Contributorship statement**

36 All authors were involved in the design of the study and the acquisition of the research funds.
37 ILS drafted the initial versions of the article. All authors critically revised the various drafts
38 and approved the final version.
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47 **Competing interests**

48 The authors have no known conflict of interest and have no commercial interest in this study.
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53 **Funding**

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3 This study is funded by the Dutch Association for Manual Therapy (NVMT, grant ID. Top-
4 down grant_2018) and by the Faculty of Behavioural and Movement Sciences (grant ID. Lab
5 Fund_2019) of Vrije Universiteit Amsterdam.
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11 **Data sharing statement**

12 Data are available upon reasonable request.
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18 **REFERENCES**

- 19 1. Sawicki CM, Humeidan ML, Sheridan JF. Neuroimmune Interactions in Pain and Stress:
20 An Interdisciplinary Approach. *Neuroscientist*. 2020:1073858420914747.
21
- 22 2. Gold JE, Hallman DM, Hellstrom F, et al. Systematic review of biochemical biomarkers
23 for neck and upper-extremity musculoskeletal disorders. *Scand J Work Environ Health*.
24 2016;42(2):103-24.
25
- 26 3. Farrell SF, de Zoete R, Cabot PJ, Sterling M. Systemic inflammatory markers in neck pain:
27 a systematic review with meta-analysis. *Eur J Pain*. 2020.
28
- 29 4. Carp SJ, Barbe MF, Winter KA, Amin M, Barr AE. Inflammatory biomarkers increase
30 with severity of upper-extremity overuse disorders. *Clin Sci (Lond)*. 2007;112(5):305-14.
31
- 32 5. Albrecht DS, Ahmed SU, Kettner NW, et al. Neuroinflammation of the spinal cord and
33 nerve roots in chronic radicular pain patients. *Pain*. 2018;159(5):968-77.
34
- 35 6. Loggia ML, Chonde DB, Akeju O, et al. Evidence for brain glial activation in chronic pain
36 patients. *Brain*. 2015;138(Pt 3):604-15.
37
- 38 7. Jungen MJ, Ter Meulen BC, van Osch T, Weinstein HC, Ostelo R. Inflammatory
39 biomarkers in patients with sciatica: a systematic review. *BMC Musculoskelet Disord*.
40 2019;20(1):156.
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3 8. Sterling M, Elliott JM, Cabot PJ. The course of serum inflammatory biomarkers following
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
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44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
whiplash injury and their relationship to sensory and muscle measures: a longitudinal cohort
study. *PLoS One*. 2013;8(10):e77903.
9. Roy RA, Boucher JP, Comtois AS. Inflammatory response following a short-term course
of chiropractic treatment in subjects with and without chronic low back pain. *J Chiropr Med*.
2010;9(3):107-14.
10. Teodorczyk-Injeyan JA, McGregor M, Triano JJ, Injeyan SH. Elevated Production of
Nociceptive CC Chemokines and sE-Selectin in Patients With Low Back Pain and the Effects
of Spinal Manipulation: A Nonrandomized Clinical Trial. *Clin J Pain*. 2018;34(1):68-75.
11. Michalis K, Anastasios P, Theodoros B, Konstantinos Z. The Impact of Manual Therapy
Techniques on Pain, Disability and Il-1b Levels in Patients with Chronic Cervical Pain.
International Journal of Physiotherapy. 2019;6(6):268-76.
12. Martins DF, Mazzardo-Martins L, Gadotti VM, et al. Ankle joint mobilization reduces
axonotmesis-induced neuropathic pain and glial activation in the spinal cord and enhances
nerve regeneration in rats. *Pain*. 2011;152(11):2653-61.
13. Santos FM, Silva JT, Giardini AC, et al. Neural mobilization reverses behavioral and
cellular changes that characterize neuropathic pain in rats. *Mol Pain*. 2012;8:57.
14. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA. The anti-
inflammatory effects of exercise: mechanisms and implications for the prevention and
treatment of disease. *Nat Rev Immunol*. 2011;11(9):607-15.
15. McGee SL, Hargreaves M. Exercise adaptations: molecular mechanisms and potential
targets for therapeutic benefit. *Nat Rev Endocrinol*. 2020.
16. Lutke Schipholt IJ, Coppieters MW, Meijer OG, Tompra N, de Vries RBM, Scholten-
Peeters GGM. Effects of joint and nerve mobilisation on neuroimmune responses in animals

1
2
3 and humans with neuromusculoskeletal conditions: a systematic review and meta-analysis.

4
5 Pain Rep. 2021;6(2):e927.

6
7 17. Scholten-Peeters GG, Thoomes E, Konings S, et al. Is manipulative therapy more
8
9 effective than sham manipulation in adults : a systematic review and meta-analysis. *Chiropr
10
11
12 Man Therap.* 2013;21(1):34.

13
14 18. Coulter ID, Crawford C, Vernon H, et al. Manipulation and Mobilization for Treating
15
16 Chronic Nonspecific Neck Pain: A Systematic Review and Meta-Analysis for an
17
18 Appropriateness Panel. *Pain Physician.* 2019;22(2):E55-e70.

19
20 19. Paige NM, Miake-Lye IM, Booth MS, et al. Association of Spinal Manipulative Therapy
21
22 With Clinical Benefit and Harm for Acute Low Back Pain: Systematic Review and Meta-
23
24 analysis. *Jama.* 2017;317(14):1451-60.

25
26 20. Bialosky JE, Beneciuk JM, Bishop MD, et al. Unraveling the Mechanisms of Manual
27
28 Therapy: Modeling an Approach. *J Orthop Sports Phys Ther.* 2018;48(1):8-18.

29
30 21. Mintken PE, Rodeghero J, Cleland JA. Manual therapists - Have you lost that loving
31
32 feeling?! *J Man Manip Ther.* 2018;26(2):53-54.

33
34 22. Randoll C, Gagnon-Normandin V, Tessier J, et al. The mechanism of back pain relief by
35
36 spinal manipulation relies on decreased temporal summation of pain. *Neuroscience.*
37
38 2017;349:220-28.

39
40 23. Courtney CA, Steffen AD, Fernandez-de-Las-Penas C, Kim J, Chmell SJ. Joint
41
42 Mobilization Enhances Mechanisms of Conditioned Pain Modulation in Individuals With
43
44 Osteoarthritis of the Knee. *J Orthop Sports Phys Ther.* 2016;46(3):168-76.

45
46 24. Song X, Gan Q, Cao J, Wang Z, Rupert RL. Spinal manipulation reduces pain and
47
48 hyperalgesia after lumbar intervertebral foramen inflammation in the rat. *Journal of
49
50 Manipulative & Physiological Therapeutics.* 2006;29(1):5-13.

- 1
2
3 25. Song XJ, Huang ZJ, Song WB, et al. Attenuation Effect of Spinal Manipulation on
4 Neuropathic and Postoperative Pain Through Activating Endogenous Anti-Inflammatory
5 Cytokine Interleukin 10 in Rat Spinal Cord. *Journal of Manipulative and Physiological*
6
7
8
9
10 Therapeutics. 2016;39(1):42-53.
11
- 12 26. Bialosky JE, Bishop MD, Penza CW. Placebo Mechanisms of Manual Therapy: A Sheep
13
14
15 in Wolf's Clothing? *J Orthop Sports Phys Ther.* 2017;47(5):301-04.
16
- 17 27. Teodorczyk-Injeyan JA, Injeyan HS, Ruegg R. Spinal manipulative therapy reduces
18
19
20
21
22
23 inflammatory cytokines but not substance P production in normal subjects. *J Manipulative*
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
28. Lutke Schipholt IJ, Scholten-Peeters GGM, Bontkes HJ, Coppieters MW. Multiple
confounders influence the association between low-grade systemic inflammation and
musculoskeletal pain. A call for a prudent interpretation of the literature. *Spine J.*
2018;18(11):2162-63.
29. Chan AW, Tetzlaff JM, Altman DG, et al. SPIRIT 2013 statement: defining standard
protocol items for clinical trials. *Ann Intern Med.* 2013;158(3):200-7.
30. Cuschieri S. The CONSORT statement. *Saudi J Anaesth.* 2019;13(Suppl 1):S27-s30.
31. Hoffmann TC, Glasziou PP, Boutron I, et al. Better reporting of interventions: template
for intervention description and replication (TIDieR) checklist and guide. *Bmj.*
2014;348:g1687.
32. Haldeman S, Carroll L, Cassidy JD, Schubert J, Nygren Å. The Bone and Joint Decade
2000–2010 Task Force on Neck Pain and Its Associated Disorders: Executive Summary.
Spine. 2008;33(4S):S5-S7.
33. Hutting N, Kerry R, Coppieters MW, Scholten-Peeters GGM. Considerations to improve
the safety of cervical spine manual therapy. *Musculoskelet Sci Pract.* 2018;33:41-45.

- 1
2
3 34. Rushton A, Rivett D, Carlesso L, Flynn T, Hing W, Kerry R. International framework for
4 examination of the cervical region for potential of Cervical Arterial Dysfunction prior to
5 Orthopaedic Manual Therapy intervention. *Man Ther.* 2014;19(3):222-8.
6
7
8
9
10 35. Bier JD, Scholten-Peeters WGM, Staal JB, et al. Clinical Practice Guideline for Physical
11 Therapy Assessment and Treatment in Patients With Nonspecific Neck Pain. *Phys Ther.*
12 2018;98(3):162-71.
13
14
15
16 36. Finucane LM, Downie A, Mercer C, et al. International Framework for Red Flags for
17 Potential Serious Spinal Pathologies. *J Orthop Sports Phys Ther.* 2020;50(7):350-72.
18
19
20
21 37. Jull G, Moore A, Falla D, Lewis J, McCarthy C, Sterling M. *Grieve's Modern*
22 *Musculoskeletal Physiotherapy*: Elsevier; 2015.
23
24
25
26 38. Manning DM, Dedrick GS, Sizer PS, Brismee JM. Reliability of a seated three-
27 dimensional passive intervertebral motion test for mobility, end-feel, and pain provocation in
28 patients with cervicgia. *J Man Manip Ther.* 2012;20(3):135-41.
29
30
31
32 39. van Trijffel E, Anderegg Q, Bossuyt PM, Lucas C. Inter-examiner reliability of passive
33 assessment of intervertebral motion in the cervical and lumbar spine: a systematic review.
34 *Man Ther.* 2005;10(4):256-69.
35
36
37
38
39 40. van der El A. *Orthopaedic Manual Therapy Diagnosis: Spine and Temporomandibular*
40 *Joints*. Gartside M, editor: Jones and Barlett Publishers; 2010.
41
42
43
44 41. Licciardone JC, Russo DP. Blinding protocols, treatment credibility, and expectancy:
45 methodologic issues in clinical trials of osteopathic manipulative treatment. *J Am Osteopath*
46 *Assoc.* 2006;106(8):457-63.
47
48
49
50 51 42. Fulda KG, Slichon T, Stoll ST. Patient expectations for placebo treatments commonly used
52 in osteopathic manipulative treatment (OMT) clinical trials: a pilot study. *Osteopath Med*
53 *Prim Care.* 2007;1:3.
54
55
56
57
58
59
60

- 1
2
3 43. Dworkin RH, Turk DC, Wyrwich KW, et al. Interpreting the clinical importance of
4 treatment outcomes in chronic pain clinical trials: IMMPACT recommendations. *J Pain*.
5 2008;9(2):105-21.
6
7
8
9
10 44. Teodorczyk-Injeyan JA, Triano JJ, Injeyan HS. Nonspecific Low Back Pain:
11 Inflammatory Profiles of Patients With Acute and Chronic Pain. *Clin J Pain*.
12 2019;35(10):818-25.
13
14
15
16 45. Segre E, Fullerton JN. Stimulated Whole Blood Cytokine Release as a Biomarker of
17 Immunosuppression in the Critically Ill: The Need for a Standardized Methodology. *Shock*.
18 2016;45(5):490-4.
19
20
21
22
23 46. Teodorczyk-Injeyan JA, Triano JJ, McGregor M, Woodhouse L, Injeyan HS. Effect of
24 Interactive Neurostimulation Therapy on Inflammatory Response in Patients With Chronic
25 and Recurrent Mechanical Neck Pain. *J Manipulative Physiol Ther*. 2015;38(8):545-54.
26
27
28
29
30 47. Teodorczyk-Injeyan JA, Triano JJ, McGregor M, Woodhouse L, Injeyan HS. Elevated
31 production of inflammatory mediators including nociceptive chemokines in patients with neck
32 pain: a cross-sectional evaluation. *J Manipulative Physiol Ther*. 2011;34(8):498-505.
33
34
35
36
37 48. Kwok YH, Hutchinson MR, Gentgall MG, Rolan PE. Increased responsiveness of
38 peripheral blood mononuclear cells to in vitro TLR 2, 4 and 7 ligand stimulation in chronic
39 pain patients. *PLoS One*. 2012;7(8):e44232.
40
41
42
43
44 49. Habbal OA, Al-Jabri AA. Circadian rhythm and the immune response: a review. *Int Rev*
45 *Immunol*. 2009;28(1):93-108.
46
47
48
49 50. Twisk JWR. Inleiding in de toegepaste biostatistiek: Bohn Stafleu van Loghum; 2016.
50
51 51. Lutke Schipholt IJ, Scholten-Peeters GGM, Bontkes HJ, Coppieters MW. Authors' Reply:
52 Confounding and mediation to reveal the true association between systemic inflammation and
53 musculoskeletal pain. *Spine J*. 2019;19(11):1901.
54
55
56
57
58
59
60

- 1
2
3 52. World Medical Association Declaration of Helsinki: ethical principles for medical
4
5 research involving human subjects. *Jama*. 2013;310(20):2191-4.
6
7
8 53. van den Berg R, Jongbloed EM, de Schepper EIT, Bierma-Zeinstra SMA, Koes BW,
9
10 Luijsterburg PAJ. The association between pro-inflammatory biomarkers and nonspecific low
11
12 back pain: a systematic review. *Spine J*. 2018;18(11):2140-51.
13
14
15 54. Morris P, Ali K, Merritt M, Pelletier J, Macedo LG. A systematic review of the role of
16
17 inflammatory biomarkers in acute, subacute and chronic non-specific low back pain. *BMC*
18
19 *Musculoskelet Disord*. 2020;21(1):142.
20
21
22 55. Klyne DM, Hodges PW. Letter to the editor concerning “Multiple confounders influence
23
24 the association between low-grade systemic inflammation and musculoskeletal pain. A call
25
26 for a prudent interpretation of the literature”. *Spine J*. 2019.
27
28
29 56. Li Y, Liu J, Liu ZZ, Duan DP. Inflammation in low back pain may be detected from the
30
31 peripheral blood: suggestions for biomarker. *Biosci Rep*. 2016;36(4).
32
33
34 57. Rossetini G, Camerone EM, Carlino E, Benedetti F, Testa M. Context matters: the
35
36 psychoneurobiological determinants of placebo, nocebo and context-related effects in
37
38 physiotherapy. *Archives of physiotherapy*. 2020;10:11-11.
39
40
41 58. Malfliet A, Lluch Girbés E, Pecos-Martin D, Gallego-Izquierdo T, Valera-Calero A. The
42
43 Influence of Treatment Expectations on Clinical Outcomes and Cortisol Levels in Patients
44
45 With Chronic Neck Pain: An Experimental Study. *Pain Pract*. 2019;19(4):370-81.
46
47
48 59. Emshoff R, Bertram S, Emshoff I. Clinically important difference thresholds of the visual
49
50 analog scale: a conceptual model for identifying meaningful intraindividual changes for pain
51
52 intensity. *Pain*. 2011;152(10):2277-82.
53
54
55 60. Kersten P, White PJ, Tennant A. Is the pain visual analogue scale linear and responsive to
56
57 change? An exploration using Rasch analysis. *PloS one*. 2014;9(6):e99485-e85.
58
59
60

- 1
2
3 61. Klyne DM, Barbe MF, Hodges PW. Systemic inflammatory profiles and their
4 relationships with demographic, behavioural and clinical features in acute low back pain.
5 Brain Behav Immun. 2017;60:84-92.
6
7
8
9
10 62. Jorritsma W, de Vries GE, Dijkstra PU, Geertzen JH, Reneman MF. Neck Pain and
11 Disability Scale and Neck Disability Index: validity of Dutch language versions. Eur Spine J.
12 2012;21(1):93-100.
13
14
15
16 63. Kamper SJ, Ostelo RW, Knol DL, Maher CG, de Vet HC, Hancock MJ. Global Perceived
17 Effect scales provided reliable assessments of health transition in people with musculoskeletal
18 disorders, but ratings are strongly influenced by current status. J Clin Epidemiol.
19 2010;63(7):760-66.e1.
20
21
22
23
24 64. Sleijser-Koehorst MLS, Bijker L, Cuijpers P, Scholten-Peters GGM, Coppieters MW.
25 Preferred self-administered questionnaires to assess fear of movement, coping, self-efficacy,
26 and catastrophizing in patients with musculoskeletal pain-A modified Delphi study. Pain.
27 2019;160(3):600-06.
28
29
30
31
32
33 65. Freynhagen R, Baron R, Gockel U, Tolle TR. painDETECT: a new screening
34 questionnaire to identify neuropathic components in patients with back pain. Curr Med Res
35 Opin. 2006;22(10):1911-20.
36
37
38
39
40
41 66. Neblett R, Cohen H, Choi Y, et al. The Central Sensitization Inventory (CSI): establishing
42 clinically significant values for identifying central sensitivity syndromes in an outpatient
43 chronic pain sample. J Pain. 2013;14(5):438-45.
44
45
46
47 67. Bijker L, Sleijser-Koehorst MLS, Coppieters MW, Cuijpers P, Scholten-Peters GGM.
48 Preferred Self-Administered Questionnaires to Assess Depression, Anxiety and Somatization
49 in People With Musculoskeletal Pain - A Modified Delphi Study. J Pain. 2019.
50
51
52
53
54 68. Blikman T, Stevens M, Bulstra SK, van den Akker-Scheek I, Reininga IH. Reliability and
55 validity of the Dutch version of the International Physical Activity Questionnaire in patients
56
57
58
59
60

1
2
3 after total hip arthroplasty or total knee arthroplasty. *J Orthop Sports Phys Ther.*

4
5 2013;43(9):650-9.

6
7 69. Poppel MNMv, Paw MCA, Mechelen WV, editors. *Reproduceerbaarheid en validiteit van*
8
9 *de Nederlandse versie van de International Physical Activity Questionnaire (IPAQ)2004.*

10
11 70. Buysse DJ, Reynolds CF, Monk TH, Berman SR, Kupfer DJ. *The Pittsburgh sleep quality*
12
13 *index: A new instrument for psychiatric practice and research. Psychiatry Research.*

14
15 1989;28(2):193-213.

16
17 71. Kovacs FM, Abraira V, Royuela A, et al. *Minimum detectable and minimal clinically*
18
19 *important changes for pain in patients with nonspecific neck pain. BMC Musculoskelet*
20
21 *Disord.* 2008;9:43.

22
23 72. Cuijpers P, Smits N, Donker T, ten Have M, de Graaf R. *Screening for mood and anxiety*
24
25 *disorders with the five-item, the three-item, and the two-item Mental Health Inventory.*
26
27 *Psychiatry Research.* 2009;168(3):250-55.

28
29 73. Hoeymans N, Garssen AA, Westert GP, Verhaak PF. *Measuring mental health of the*
30
31 *Dutch population: a comparison of the GHQ-12 and the MHI-5. Health Qual Life Outcomes.*
32
33 2004;2:23.

34
35 74. Fletcher JP, Bandy WD. *Intrarater reliability of CROM measurement of cervical spine*
36
37 *active range of motion in persons with and without neck pain. J Orthop Sports Phys Ther.*
38
39 2008;38(10):640-5.

40
41 75. Walton DM, Macdermid JC, Nielson W, Teasell RW, Chiasson M, Brown L. *Reliability,*
42
43 *standard error, and minimum detectable change of clinical pressure pain threshold testing in*
44
45 *people with and without acute neck pain. J Orthop Sports Phys Ther.* 2011;41(9):644-50.

46
47 76. Rolke R, Baron R, Maier C, et al. *Quantitative sensory testing in the German Research*
48
49 *Network on Neuropathic Pain (DFNS): standardized protocol and reference values. Pain.*
50
51 2006;123(3):231-43.

- 1
2
3 77. Schlecht I, Wiggermann P, Behrens G, et al. Reproducibility and validity of ultrasound for
4 the measurement of visceral and subcutaneous adipose tissues. *Metabolism*.
5
6 2014;63(12):1512-9.
7
8
9 78. Park HS, Park JY, Yu R. Relationship of obesity and visceral adiposity with serum
10 concentrations of CRP, TNF-alpha and IL-6. *Diabetes Res Clin Pract*. 2005;69(1):29-35.
11
12 79. Bouman A, Moes H, Heineman MJ, de Leij LF, Faas MM. The immune response during
13 the luteal phase of the ovarian cycle: increasing sensitivity of human monocytes to endotoxin.
14
15 *Fertil Steril*. 2001;76(3):555-9.
16
17
18 80. Myriantsefs P, Karatzas S, Venetsanou K, et al. Seasonal variation in whole blood
19 cytokine production after LPS stimulation in normal individuals. *Cytokine*. 2003;24(6):286-
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
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36
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3 **Figure 1:** Anticipated flow of the study
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6 **Abbreviations:** GPE: global perceived effect; VAS: visual analogue scale
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3 **Figure 2.** Spinal mobilisation and manipulation techniques.
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6 Depending on the identified painful segmental levels, the clinician can select from different
7 cervical mobilisation techniques (A-C); For techniques A-C, the participant will be seated on
8 a chair, leaning against the upper leg or shoulder of the clinician. **Panel A:** Mobilisation
9 targeting the atlanto-axial joints. The cervical segments below the second cervical vertebrae
10 are submaximal rotated and lateroflexed. With the clinician's hypothenar region of the hand
11 over the structures overlying the arcus of the first vertebrae, the clinician moved the head
12 further in rotation.⁽³⁹⁾ **Panel B:** Segmental zygapophyseal joint mobilisation (C2 to C7; the
13 image shows the technique for C3-C4). First, the occipital-atlanto-axial joint is maximally
14 rotated in the direction of the facet joint being mobilised. Subsequently, the head is moved to
15 extension, ipsilateral lateroflexion and rotation until pressure from the thumb is felt. This
16 technique is repeated on the lower level until the painful cervical segment is reached (C3-C4).
17 Next, on the painful cervical segment, pressure will be given in a cranio-ventral direction.
18 **Panel C:** Mobilisation technique targeting the occipital-atlanto-axial joints. The
19 clinician's hypothenar region is placed against the mastoid process. C2 to C7 are
20 submaximally locked in flexion, rotation and lateroflexion. The head is then moved in a
21 medio-caudal direction.⁽³⁹⁾ **Panel D:** Spinal manipulation technique targeting the cervico-
22 thoracic junction. The participant will be seated on a treatment table. The height of the table
23 will be adjusted to the level of the clinician's abdomen. The participant's hands will be placed
24 on the back of their head (with one hand placed over the other hand, rather than with
25 interlocking fingers), and with the shoulders slightly retracted. The clinician's hands will be
26 placed over the hands of the participant, with the clinician's forearms ventral to the shoulder
27 of the participant. Then, a high-velocity, low-amplitude movement will be applied in a dorsal-
28 cranial direction.⁽³⁹⁾ Green arrows represent the direction of the mobilisation (Panel A-C) or
29 manipulation (Panel D).
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Table 1: Overview of the neuroimmune responses

Domain	Neuroimmune parameters	Timing of measurements			
		T0	T1	T2	T3
Systemic inflammatory marker directly from blood samples ^a	TNF- α , TNF-RII, IL-1 β , IL-1RA, hsCRP ^b	√	√	√	-
Inflammatory marker concentration after <i>in-vitro</i> stimulation of whole blood cells ^c	TNF- α , IL-1 β , IL-1RA, IL-4, IL-10, CCL2, CCL3, CCL4	√	√	√	-
Ex-vivo serum cortisol ^d	Cortisol	√	√	-	-
Phenotypic analysis of peripheral blood mononuclear cells ^e	CD45 ⁺ , CD3 ⁺ , CD4 ⁺ , CD25 ^{hi} , CD8 ⁺ , CD56 ⁺ , CD19 ⁺ , CD14 ⁺ , HLA-DR, TLR-4	√	-	√	-

a) Measured using multianalyte assay Ella (R&D systems, Minneapolis, United States)

b) Cardiac C-Reactive Protein (Latex) High Sensitive using Roche/Hitachi cobas c systems.

c) Stimulated for 24 hours at 37°C, in a humidified 5% CO₂ incubator, with lipopolysaccharide (LPS) from *Escherichia coli* O55:B5 at a concentration of 1ng/ml and 10 μ g/ml. Determined using a custom-made U-plex (MSD, Maryland, United States)

d) Using conventional electrochemiluminescence immunoassay (ECLIA), Roche (Cobas Cortisol, 2nd generation).

e) Determined by 10-color flowcytometry (FCM): CD45⁺ = General Leukocyte marker; CD3⁺ = T-cell marker; CD3⁺CD4⁺ = CD4⁺ T-helper marker; CD3⁺CD4⁺CD25^{hi} = T-regulator cell marker; CD3⁺CD8⁺ = Cytotoxic T-cell marker; CD3⁺CD56⁺ = Natural Killer cell marker; CD19⁺ = B-cell marker; CD14⁺ = monocyte marker; HLA-DR = activation marker for T-cells and monocytes; TLR-4 = Toll-like receptor 4 marker.

Abbreviations: T0: baseline; T1: immediately following the intervention; T2: two-hours following the intervention; T3: two-days following the intervention; TNF- α : Tumor Necrosis Factor- α ; TNF-RII: Tumor Necrosis Factor Receptor Antagonist 2; IL-1 β : Interleukin-1 β ; IL-1RA: Interleukin-1 receptor antagonist; hsCRP: High sensitive C-Reactive Protein; IL-4: Interleukin-4; IL-10: Interleukin-10; CCL2: c-c-motif chemokine ligand 2; CCL3: c-c-motif chemokine ligand 3; CCL4: c-c-motif chemokine ligand 4; CD: Cluster of Differentiation

Table 2: Self-reported questionnaires and physical tests

Domain	Self-reported questionnaires	Timing of measurements			
		T0	T1	T2	T3
Disability	Neck Disability Index (NDI) ^a	-	√	-	-
Perceived effect	Global Perceived Effect (GPE) ^b	-	-	√	√
Fear of movement	Tampa Scale of Kinesiophobia ^c	-	√	-	-
Type of pain	PAIN Detect Questionnaire (PDQ) ^d	-	√	-	-
Type of pain	Central Sensitisation Inventory (CSI) ^e	-	√	-	-
Depression, Anxiety, Stress	Depression Anxiety Stress Scale (DASS21) ^f	-	√	-	-
Physical activity	International Physical Activity Questionnaire (IPAQ) ^g	-	√	-	-
Catastrophising	Pain Catastrophising Scale (PCS) ^h	-	√	-	-
Sleep Quality	Pittsburgh Sleep Quality Index (PSQI) ⁱ	-	√	-	-
Pain Intensity	Visual Analogue Scale (VAS) ^j	√	√	√	√
Mental health	Mental health inventory (MHI-5) ^k	√	-	-	-

Domain	Physical tests	Timing of measurement			
		T0	T1	T2	T3
Range of motion	Cervical Range of Motion (CROM) ^l	√	√	√	-
Pain intensity	CROM-VAS test ^m	-	√	√	-
Quantitative sensory testing	Pressure Pain Threshold (PPT) ⁿ	√	√	√	-
Quantitative sensory testing	Wind-up ratio ^o	√	√	√	-

a) The Dutch version of the NDI is a valid and responsive measure of disability.(61)

b) The GPE is a validated and reliable tool to assess health transitions in patients with musculoskeletal disorders.(62)

c) Preferred self-administrated questionnaire to asses fear of movement in musculoskeletal pain.(63)

d) Persistent pain will be categorised in two-mechanism based groups: nociceptive and neuropathic pain using the PDQ. The PD-Q is a reliable screening tool with high specificity.(64)

e) The Dutch Central Sensitization Inventory (CSI) has good internal consistency, good discriminative power and excellent test-retest reliability. A cut-off score of 40/100 provides a sensitivity of 81% and specificity of 75%.(65)

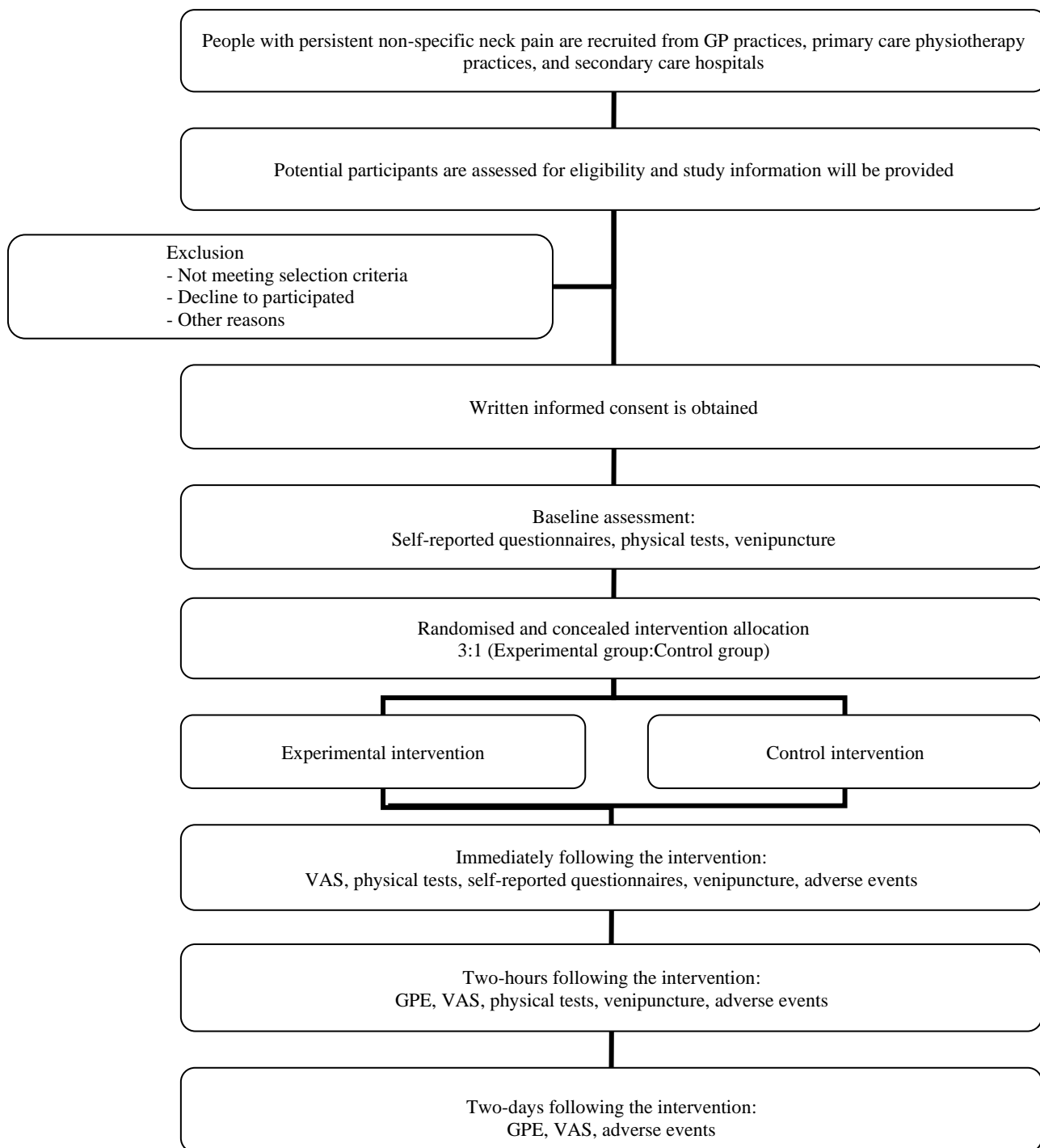
- 1
2
3 f) Preferred self-administrated questionnaire to assess depression, anxiety and stress in
4 musculoskeletal pain.(63, 66)
5
6 g) Expressed in 1000 metabolic equivalent minutes per week (Dutch-language
7 version).(67) The IPAQ has good reliability (intraclass correlation coefficient [ICC] =
8 0.70-0.96) and moderate validity ($r = 0.36-0.49$) of the IPAQ compared with an
9 accelerometer.(68)
10
11 h) Preferred self-administrated questionnaire to assess pain catastrophising in
12 musculoskeletal pain.(63)
13
14 i) Score above 5 yield a sensitivity of 89.6% and specificity of 86.5% in distinguishing
15 good and poor sleepers.(69)
16
17 j) The reliability and validity of the VAS as a measure of pain for neck pain patients is
18 good.(70)
19
20 k) General psychological status will be assessed using the MHI-5.(71) A higher score
21 indicates better mental health. Cronbach's alpha for the MHI-5 scale is 0.85.(72)
22
23 l) The CROM is a clinically reliable tool to measure active cervical range of motion
24 people with neck pain and healthy participants.(73)
25
26 m) This novel test consists of two parts. In Part 1, the participant is asked to perform
27 maximal active right and left cervical rotation and the degrees of rotation are reordered
28 using the CROM device. In this position, the pain intensity is measured with the VAS
29 following intervention. After the intervention, Part 2 of the test is performed . The
30 participant is again asked to actively rotate (left and right) to the same position as in
31 Part 1 and the pain intensity is recorded. The difference on VAS scores is the outcome
32 of the CROM-VAS test.
33
34 n) Pressure algometry over the cervical spine has shown excellent intrarater and good-to-
35 excellent interrater reliability in individuals with acute neck pain.(74) This study
36 reported that the MDC for PPT over the cervical spine and tibialis anterior muscle in
37 patients with acute neck pain was 47.2 and 97.9 kPa, respectively.(74) To determine
38 changes in widespread pressure pain sensitivity, PPTs will be assessed bilaterally over
39 the mid-point trapezius (pars descendens), second metacarpal, and tibialis anterior
40 muscle.
41
42 o) Using a pinprick 256 mN wind up ratio will be calculated bilaterally over the mid-
43 point trapezius (pars descendens) and tibiales anterior muscle.(75)
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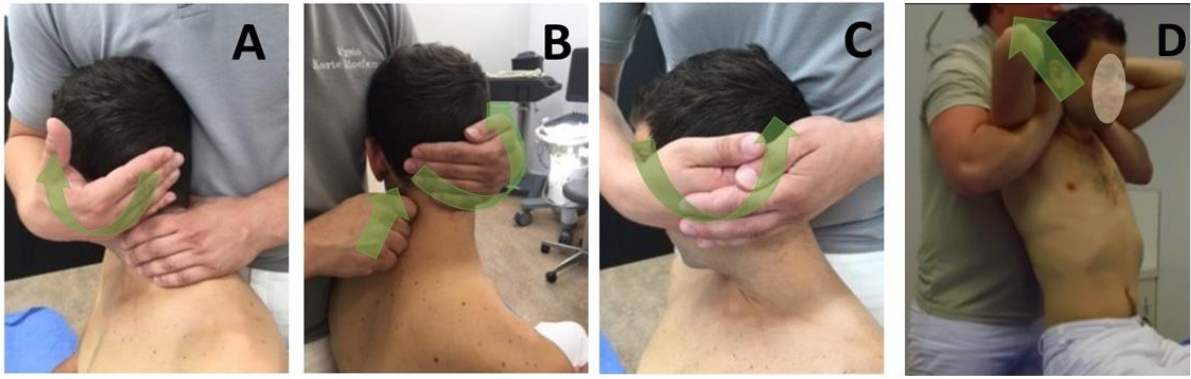
47 **Abbreviations:** T0: baseline; T1: immediately following the intervention; T2: two-hours
48 following the intervention; T3: two-days following the intervention
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Table 3: Potential confounding variables that will be assessed

Potential confounding variables	
Co-morbidities	Number of co-morbidities
Alcohol use	- Non-drinker - Moderate drinker (women: 1-14 glasses/week) (men: 1-21 glasses/week) - Heavy drinker (women: >14 glasses/week) (men: >21 glasses/week)
Smoking	- Never smoked - Former smoker - Current smoker
Body Mass Index	BMI calculated by dividing body weight (kg) by height (m ²)
Medication use	Type and number of medications used
Drugs use	Recreational drugs use - Yes - No
Visceral Adipose Tissue(76, 77)	Linear distance between abdominal peritoneum and ventral aspect of vertebrae will be assessed using ultrasonography
Physical activity	International Physical Activity Questionnaire, expressed in 1000 metabolic equivalent minutes per week (Dutch version)
Menstrual cycle(78)	Regular menstrual cycle (yes/no), whether women are in the luteal or follicular stage (yes/no), menopause (yes/no) and post menopause (yes/no)
Season(79)	Timing of experiment (summer, autumn, spring or winter)
Age	Age in years
Psychological status(71)	Mental health inventory-5
Intervention expectations(41)	The extent to which they agree (using a four-point Likert scale) with four statements (Appendix A)

Abbreviation: BMI: Body Mass Index





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3 **Appendix A:** Four statements regarding the intervention expectation of the participants
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5 (modified from(41))
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7 I believe this intervention will allow me to get better quicker. Strongly agree
8 Agree
9 Disagree
10 Strongly disagree
11
12

13 I believe this intervention will decrease my neck pain. Strongly agree
14 Agree
15 Disagree
16 Strongly disagree
17
18

19 I believe this intervention will make me more able to do the things I want to do. Strongly agree
20 Agree
21 Disagree
22 Strongly disagree
23
24

25 This seems like a logical way to treat neck pain. Strongly agree
26 Agree
27 Disagree
28 Strongly disagree
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The TIDieR (Template for Intervention Description and Replication) Checklist*:

Information to include when describing an intervention and the location of the information

Item number	Item	Where located **	
		Primary paper (page or appendix number)	Other † (details)
1.	BRIEF NAME Provide the name or a phrase that describes the intervention.	4 & P.6	_____
2.	WHY Describe any rationale, theory, or goal of the elements essential to the intervention.	4	_____
3.	WHAT Materials: Describe any physical or informational materials used in the intervention, including those provided to participants or used in intervention delivery or in training of intervention providers. Provide information on where the materials can be accessed (e.g. online appendix, URL).	6, Figure 2 & 3	_____
4.	Procedures: Describe each of the procedures, activities, and/or processes used in the intervention, including any enabling or support activities.	4-6	_____
5.	WHO PROVIDED For each category of intervention provider (e.g. psychologist, nursing assistant), describe their expertise, background and any specific training given.	4	_____
6.	HOW Describe the modes of delivery (e.g. face-to-face or by some other mechanism, such as internet or telephone) of the intervention and whether it was provided individually or in a group.	4-6	_____
7.	WHERE Describe the type(s) of location(s) where the intervention occurred, including any necessary infrastructure or relevant features.	3	_____

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WHEN and HOW MUCH

8. Describe the number of times the intervention was delivered and over what period of time including the number of sessions, their schedule, and their duration, intensity or dose.

TAILORING

9. If the intervention was planned to be personalised, titrated or adapted, then describe what, why, when, and how.

MODIFICATIONS

- 10.* If the intervention was modified during the course of the study, describe the changes (what, why, when, and how).

HOW WELL

11. Planned: If intervention adherence or fidelity was assessed, describe how and by whom, and if any strategies were used to maintain or improve fidelity, describe them.

- 12.* Actual: If intervention adherence or fidelity was assessed, describe the extent to which the intervention was delivered as planned.

2, P4-6

4-5

A.

A.

A.

** **Authors** - use N/A if an item is not applicable for the intervention being described. **Reviewers** – use ‘?’ if information about the element is not reported/not sufficiently reported.

† If the information is not provided in the primary paper, give details of where this information is available. This may include locations such as a published protocol or other published papers (provide citation details) or a website (provide the URL).

‡ If completing the TIDieR checklist for a protocol, these items are not relevant to the protocol and cannot be described until the study is complete.

* We strongly recommend using this checklist in conjunction with the TIDieR guide (see *BMJ* 2014;348:g1687) which contains an explanation and elaboration for each item.

* The focus of TIDieR is on reporting details of the intervention elements (and where relevant, comparison elements) of a study. Other elements and methodological features of studies are covered by other reporting statements and checklists and have not been duplicated as part of the TIDieR checklist. When a **randomised trial** is being reported, the TIDieR checklist should be used in conjunction with the CONSORT statement (see www.consort-statement.org) as an extension of **Item 5 of the CONSORT 2010 Statement**. When a **clinical trial protocol** is being reported, the TIDieR checklist should be used in conjunction with the SPIRIT statement as an extension of **Item 11 of the SPIRIT 2013 Statement** (see www.spirit-statement.org). For alternate study designs, TIDieR can be used in conjunction with the appropriate checklist for that study design (see www.equator-network.org).

BMJ Open

Neuroimmune responses following joint mobilisation and manipulation in people with persistent neck pain: A protocol for a randomised placebo-controlled trial

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2021-055748.R1
Article Type:	Protocol
Date Submitted by the Author:	22-Dec-2021
Complete List of Authors:	Lutke Schipholt, Ivo J; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences; Amsterdam UMC VUMC Site, Department of Clinical Chemistry, Laboratory Medical Immunology Scholten-Peeters, Gwendolijne ; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences Bontkes, Hetty; Amsterdam UMC VUMC Site, Department of Clinical Chemistry, Laboratory Medical Immunology Coppieters, Michel ; Griffith University Menzies Health Institute Queensland; Vrije Universiteit Amsterdam, Faculty of Behavioural and Movement Sciences
Primary Subject Heading:	Rehabilitation medicine
Secondary Subject Heading:	Sports and exercise medicine, Immunology (including allergy)
Keywords:	Musculoskeletal disorders < ORTHOPAEDIC & TRAUMA SURGERY, Spine < ORTHOPAEDIC & TRAUMA SURGERY, REHABILITATION MEDICINE, CLINICAL PHYSIOLOGY, IMMUNOLOGY

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Manuscripts

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3 **Neuroimmune responses following joint mobilisation and manipulation in people with**
4
5 **persistent neck pain: A protocol for a randomised placebo-controlled trial**
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51 **Key words:** Neuroimmune, musculoskeletal health, pain, immune system, physiotherapy,
52 manual therapy, non-pharmacological treatment, neck pain.
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Word count 3745/4000

ABSTRACT

Introduction: Joint mobilisation and manipulation often results in immediate pain relief in people with neck pain. However, the biological mechanisms behind pain relief are largely unknown. There is preliminary evidence that joint mobilisation and manipulation lessens the upregulated neuroimmune responses in people with persistent neck pain.

Methods and analysis: This study protocol describes a randomised placebo-controlled trial to investigate whether joint mobilisation and manipulation influence neuroimmune responses in people with persistent neck pain. People with persistent neck pain (N=100) will be allocated, in a randomised and concealed manner, to the experimental or control group (ratio 3:1).

Short-term (i.e., baseline, immediately after and two-hours after the intervention) neuroimmune responses will be assessed, such as inflammatory marker concentration following *in-vitro* stimulation of whole blood cells, systemic inflammatory marker concentrations directly from blood samples, phenotypic analysis of peripheral blood mononuclear cells, and serum cortisol. Participants assigned to the experimental group (N=75) will receive cervical mobilisations targeting the painful and/or restricted cervical segments and a distraction manipulation of the cervico-thoracic junction. Participants assigned to the control group (N=25) will receive a placebo mobilisation and placebo manipulation. Using linear mixed models, the short-term neuroimmune responses will be compared 1) between people in the experimental and control group, 2) within the experimental group, between people who experience a good outcome and those with a poor outcome. Furthermore, the association between the short-term neuroimmune responses and pain relief following joint mobilisation and manipulation will be tested in the experimental group.

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3 **Ethics and dissemination:** This trial is approved by the Medical Ethics Committee of
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6 Amsterdam University Medical Centre, location VUmc (Approval number: 2018.181).

7 **Trial registration number and status:** The study protocol is registered at trialregister.nl with
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9
10 study ID: NL6575 registered on 18-01-2018; Recruitment commenced 26 February 2019. All
11
12 data are anticipated to be collected by January 2022, when data analysis and interpretation are
13
14 anticipated to commence.
15
16

17 18 19 **Strengths and limitations of this study**

- 20
21 ❖ This study provides insight in the interplay between joint mobilisation and
22
23 manipulation, neuroimmune responses, and pain relief in people with persistent neck
24
25 pain.
26
27
- 28
29 ❖ By adding a placebo-control group, possible working mechanisms of joint
30
31 mobilisation and manipulation on neuroimmune responses may be revealed.
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- 33
34 ❖ The interventions will be delivered by two musculoskeletal physiotherapists, which
35
36 may limit the generalisability.
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- 38
39 ❖ Due to the small control group, it is not feasible to divide the control participants
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41 according to outcome.
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44 ❖ Inflammatory indices will be calculated that combine overall inflammatory, pro-
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46 inflammatory, anti-inflammatory and ratio pro/anti-inflammatory markers.
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1. INTRODUCTION

The disruption of the bidirectional communication pathways between the central nervous system and the immune system may play an important role in persistent pain.(1) Over the last two decades, it has become apparent that neuroimmune crosstalk is present in musculoskeletal pain, and may play a mediating role in the transition from acute to persistent pain.(1) For people with persistent neck pain, aberrant neuroimmune responses may be present, such as systemically elevated levels of inflammatory markers.(2, 3) These increased neuroimmune responses may be relevant to understand and manage persistent spinal pain.(3) A growing body of literature suggests that these neuroimmune responses are associated with pain intensity,(4-6) disability(7) and recovery,(8) and can be influenced by musculoskeletal physiotherapy, such as joint mobilisation and manipulation,(9-11) nerve mobilisation(12, 13) and exercise.(14-16)

Several meta-analyses indicate that musculoskeletal physiotherapy for people with spinal pain may provide immediately pain relief and improvements in functional activities compared to no treatment, placebo or other treatments.(17-19) Nevertheless, unravelling the mechanism of how joint mobilisation and manipulation results in pain relief remains an area for further investigation.(20, 21) There are various explanations of how joint mobilisation and manipulation might cause pain relief, including neurophysiological,(22, 23) neuromuscular,(20) neuroimmune(24, 25) and non-specific responses.(26)

Recent studies suggest a possible neuroimmune-mediated mechanism of pain relief following joint mobilisation and manipulation.(9-11) For example, a reduction in systemic inflammatory marker concentration directly from blood samples(9, 11) and a reduction in inflammatory marker concentration following *in-vitro* stimulation of whole blood cells(10, 27) were found immediately following the intervention. These studies have however

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3 important methodological limitations, such as inclusion of healthy participants,(27) modest
4 sample sizes,(9, 10) a narrow selection of inflammatory markers,(9-11) lack of correction for
5
6 potential confounding variables,(9, 10, 28) and lack of a placebo-control group.(9, 10)
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10 Therefore, we will conduct an adequately powered, placebo-controlled randomised clinical
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12 trial in people with persistent neck pain, which will evaluate a broad range of inflammatory
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14 markers. The purpose of this paper is to describe the study protocol to investigate the short-
15
16 term effects of joint mobilisation and manipulation on neuroimmune responses in people with
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18 persistent neck pain.
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23 24 **2. METHODS**

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28 This manuscript followed the guidelines for clinical trial protocols (SPIRIT
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30 statement),(29) for reporting randomised trials (CONSORT statement),(30) and for
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32 intervention description and replication (TIDieR checklist).(31)
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38 **Aim**

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40 The overall aim of this clinical trial is to gain insights in the relation between short-
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42 term neuroimmune responses following joint mobilisation and manipulation and pain relief in
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44 people with persistent neck pain. The specific aims are: 1) to compare the short-term
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46 neuroimmune responses between the experimental and control group; 2) to compare the short-
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48 term neuroimmune responses of those in the experimental group with a good outcome (i.e.,
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50 immediately pain relief) with those in the experimental group with a poor outcome; and 3) to
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52 assess the association between short-term neuroimmune responses and pain relief in the
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54 experimental group.
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Study design and setting

The study is a placebo-controlled randomised trial with follow-up at three time points: baseline, immediately, and two-hours and two-days following the intervention (Fig. 1).

Participants will be recruited from GP clinics, primary care physiotherapy practices and outpatient services (neurology and orthopaedic departments) at secondary care hospitals. Data are anticipated to be collected between February 2019 and January 2022, when data analysis and interpretation are anticipated to commence.

Selection criteria

Individuals meeting the following inclusion criteria are eligible to participate: age: 18-65 years; non-specific neck pain for at least six weeks⁽³²⁾ with a minimum pain intensity of 40/100 on a visual analogue scale (VAS), and a sufficient speaking and reading level of the Dutch language to complete the study. Exclusion criteria are contra-indications for cervical mobilisation or cervico-thoracic manipulation,^(33, 34) pregnancy or less than 9 months postpartum, contra-indications for venipuncture (e.g., phlebitis), treatment for the current neck pain episode during the preceding two weeks, taken corticosteroids or cytokine modulatory medication (e.g., methotrexate, infliximab) in the preceding 6 weeks, use of botulinum toxin (Botox) injection during the preceding 3 months, non-steroid anti-inflammatory drug medication within the past 7 days (e.g., diclofenac, ibuprofen, naproxen), long-distance flight within the past 7 days, ongoing shift work, having a known comorbid condition with immune/endocrine malfunction (e.g., ankylosing spondylitis, Crohn's disease, sarcoidosis, Cushing syndrome, cancer, diabetes), medical red flags suggestive of serious pathology,^(35, 36) and a diagnosed psychological condition (e.g., clinical depression).

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3 Consecutive participants who meet all selection criteria and are willing to participate
4 will be admitted to the study. All participants will provide written informed consent prior to
5 participation. Initial screening for eligibility will be conducted via telephone calls.
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10 11 12 **Randomisation, concealed allocation and blinding**

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14 Block randomisation will be used to allocate participants to the experimental or
15 control group with an allocation ratio of 3:1 (experimental : control). A computer random
16 number generator will create block sizes of 4 and 8 participants. To conceal the allocation
17 sequence, an independent person not involved in the study will assign eligible people to the
18 groups on the day the participant will enrol in the study. Blood samples will be coded to blind
19 the research assistant and laboratory investigators to the study groups. The participant,
20 research assistant and the investigator who includes the participants will be blinded for group
21 assignment. The treating clinicians, research assistant and laboratory investigators will be
22 unaware whether participants experienced a good outcome or not. All laboratory and data
23 analyses will be performed by blinded investigators.
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40 **Interventions**

41 42 ***Experimental intervention***

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44 Spinal mobilisation will consist of low-velocity, low-amplitude mobilisations at the
45 painful cervical segmental levels (Fig. 2 – Panels A-C); spinal manipulation will consist of a
46 high-velocity, low-amplitude distraction manipulation at the cervico-thoracic junction (Fig. 2
47 – Panel D).(37) These techniques aim to restore motion and reduce pain. They are commonly
48 used and are conform to the Dutch guidelines for musculoskeletal physiotherapy for treating
49 neck pain.(35) All interventions will be performed by two musculoskeletal physiotherapists
50 with more than 5 years of relevant clinical experience.
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Cervical mobilisation

Painful and restricted cervical segments will be identified by passive side-bending of the neck targeting each segmental level separately.(38) Reproduction of the participant's pain will be considered to identify the involved level(s). The inter-tester reliability for these tests is fair to substantial.(38, 39)

Depending on the identified painful or restricted spinal levels, the treating clinician may select from different mobilisation techniques: mobilisation targeting the atlanto-axial segment (Fig. 2 – Panel A); segmental zygapophyseal joint mobilisation (C2 to C7) (Fig. 2 – Panel B) and occipital-atlanto-axial joint mobilisation (Fig. 2 – Panel C). Three series of oscillations (~1Hz) will be applied for 30 seconds; with 30 seconds rest in between the series.

Cervico-thoracic junction distraction manipulation

Irrespective of the level of their neck pain, all participants will receive a distraction manipulation of the cervico-thoracic junction (Fig. 2– Panel D).(40) If there is no audible cavitation sound during the first attempt, the manipulation will be repeated once.

Control (placebo) intervention

The control group will receive a placebo mobilisation and placebo manipulation. Procedures, including the instructions, will be identical as for the experimental intervention, except that the clinician will only apply hand contact and no pressure or movement will occur. Participants will be informed that an audible popping sound may or may not occur, and that this sound is not necessary to restore motion and reduce pain.

The credibility of a control intervention can interact with participant expectations in complex ways.(41) To account for differences in intervention expectations, participants will

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3 indicate the extent to which they agree (using a four-point Likert scale) with four statements
4 regarding their intervention expectations (Table 1). These statements will be presented before
5 the delivery of the experimental and control intervention.(42)
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10 Based on the short-term changes in pain intensity score (i.e., immediately and two-
11 hours following the intervention), participants in the experimental group will be categorised
12 into those with a good outcome ($\geq 50\%$ improvement in pain intensity at both time points), a
13 poor outcome ($\leq 20\%$ improvement in pain intensity score at both time points) or an unclear
14 outcome (not fitting the criteria for a good or poor outcome).(43) Based on these cut-off
15 scores, we anticipate to have a minimum of 25 participants in both the good outcome and poor
16 outcome group. If our *a-priori* determined minimum of 25 participants in either group is not
17 achieved, the good outcome group and the poor outcome group will be supplemented with
18 respectively the best responders and poorest responders from the uncertain outcome group in
19 order to obtain 25 participants in both groups.
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35 **Outcomes**

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37 A broad range of neuroimmune responses will be monitored: a) inflammatory marker
38 concentration following *in-vitro* stimulation of whole blood cells, b) systemic inflammatory
39 marker concentrations directly from blood samples, c) phenotypic analysis of peripheral blood
40 mononuclear cells and d) *ex-vivo* serum cortisol (Table 1). To create an inflammatory
41 profile,(44) a range of pro-inflammatory and anti-inflammatory markers will be used. Ex-vivo
42 serum and supernatants after stimulation will be stored at minus 80°C and will be analysed
43 upon completion of data collection. The laboratory methodology and sample handling prior to
44 stimulation will be tightly monitored and reported, because inconsistency in interlaboratory
45 methodology and reporting impairs interpretation, comparability and reproducibility.(45)
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Primary outcomes

The primary outcomes are the short-term (i.e., immediately and two-hours following the intervention) differences in interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α) following *in-vitro* stimulation of whole blood cells. These cytokines will be determined using Meso Scale Discovery (MSD, Maryland United States) at baseline, immediately and two-hours following the intervention. These cytokines are selected because previous research has indicated that those cytokines might play a role in spinal pain.(11, 27, 46-48)

To induce cytokine production, whole blood cultures will be stimulated for 24 hours with lipopolysaccharide (LPS) from *Escherichia coli* O55:B5 (Sigma-Aldrich Chemie GmbH, Schnelldorf, Germany) at a concentration of 1 nanogram LPS/millilitre whole blood (ng/ml) and 10 microgram LPS/millilitre whole blood (μ g/ml) at 37°C in a humidified 5% CO₂ incubator. At baseline (Fig. 1) blood samples for neuroimmune measurements (one sodium heparin vacutainer without gel and one serum vacutainer without gel for each time point) will be drawn between 8:00 and 9:00 AM.(49) The cytokine levels will be determined using a custom-made U-plex MSD and expressed in picogram/millilitre (pg/ml). The entire blood stimulation procedure and MSD will be performed by an experienced laboratory technician at Amsterdam University Medical Centre, location VUmc, Department of Clinical Chemistry, Medical Immunology Laboratory.

Secondary outcomes

Several additional neuroimmune responses will be quantified as secondary outcomes at various time points (Table 1).

The levels of interleukin-1 receptor antagonist (IL-1RA), interleukin-4 (IL-4), interleukin-10 (IL-10), c-c motif chemokine ligand 2 (CCL2), c-c motif chemokine ligand 3

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3 (CCL3) and c-c motif chemokine ligand 4 (CCL4) will be determined following *in-vitro*
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5 stimulation of whole blood cells.
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8 Systemic inflammatory markers directly from blood samples (tumor necrosis factor –
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10 receptor antagonist II (TNF-RII), IL-1 β and IL-1RA) will be measured using multianalyte
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12 assay Ella (R&D systems, Minneapolis, United States) and high-sensitive c-reactive protein
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14 (hsCRP), using Roche/Hitachi cobas c systems (Indianapolis, United States).
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17 To examine a general change in inflammatory marker production, we will calculate *in-*
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19 *vitro* and *ex-vivo* overall inflammatory, pro-inflammatory, anti-inflammatory and ratio
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21 pro/anti-inflammatory indices.(50, 51) The indices will be calculated as the mean value or the
22
23 Ln-transformed data in case of non-normality and z-score standardised levels (based on the
24
25 control group or poor outcome group) of the inflammatory markers (Appendix A).
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29 Phenotypic analysis of peripheral blood mononuclear cells will be determined. The
30
31 absolute number of lymphocyte subsets (NK cells, B-cells, CD4⁺ and CD8⁺ T-cells and
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33 CD25^{hi} regulatory T-cells), monocytes, as well as activation status of these cells, HLA-DR
34
35 and TLR-4 expression, will be determined by 10-color flowcytometry (FCM, Gallios Flow
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37 Cytometer, Beckman Coulter, Indianapolis, United States; Analyse software: Kaluza).
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39 Differences between all groups in serum cortisol concentration will be determined using
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41 conventional electrochemiluminescence immunoassay (ECLIA) from Roche (Cobas Cortisol,
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43 2nd generation, Indianapolis, United States) in agreement with the manufacturer's protocol.
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49 **Procedures**

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51 Once consent is obtained (Appendix B), baseline measurements will be taken (Fig. 1).
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53 At baseline, participants will undergo physical tests to determine pain characteristics, physical
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55 functioning and body composition (Table 2 & Table 3). After this, participants will complete
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57 an electronic survey to collect sociodemographic and clinical information (Table 1) and
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3 intervention expectations (Appendix C). Participants will then undergo one venipuncture from
4 the cubital vein to fill two vacutainers which will be used to quantify the neuroimmune
5 responses (Table 1). Collection of all baseline data will take 30-45 minutes and will take place
6 at the Amsterdam University Medical Centre, location VUmc, or at a participating primary
7 care physiotherapy practice, under the supervision of a research assistant.
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15 Participants will then be randomly allocated to the experimental and control group,
16 and treated accordingly. Immediately and two-hours following the intervention, participants
17 will undergo another venipuncture to fill two vacutainers. Between the immediate and two-
18 hours follow-up measures, questionnaires will be completed to collect psychosocial
19 information such as sleep, disability and kinesiophobia (Table 2).
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Immediately and two-hours following the intervention, participants will undergo
physical tests (Fig. 1) and will rate their pain intensity on a VAS. Two-hours following the
intervention, participant will rate their perceived recovery on a 7-point Global Perceived
Effect scale (GPE) (Table 2). Two-days following the intervention, participants will receive
an electronic survey regarding potential adverse events, GPE and pain intensity. Figure 1
shows the planned flow of participants through the study.

Sample size

Based on the sample size calculation⁽⁵²⁾ (longitudinal analysis; three time points
(baseline, immediately follow-up, two-hours follow-up) with 80% power to detect a mean
difference of 550 (SD 933) for TNF- α levels with a 0.05 two-sided significance level,
correlation of 0.6 among repeated measures, ratio between groups of 0.25, a total sample size
of 91 is needed.⁽²⁷⁾ Allowing for a drop-out rate of ~10%, a total sample size of 100
participants is required.

Statistical analyses

Data will be checked for normality by the Kolmogorov-Smirnov test and visual inspection of Q-Q plots, box plots and histograms. In case of no normality of data, the data will be log transformation. Data will be presented as means with standard deviations unless otherwise noted. For the analyses, statistical significance will be set at $p < 0.05$. Intention-to-treat analyses using mixed models will be performed to analyse differences between the experimental group and control group. Linear mixed model analyses with fixed factor (time), covariate (group) and interaction (time*group) will be used to detect differences between the groups at the three time points (baseline, immediately follow-up, two-hours follow-up) for TNF- α and IL- β following *in-vitro* stimulation of whole blood cells. A random intercept will be selected to account for the correlated nature of multiple measurements from the same participant. The regression coefficient (B), p-value and confidence intervals (95%CI) will be computed for the crude models, as well as for the adjusted models.(28, 53) Linear regression analysis will be used to test for differences in phenotypic analysis of peripheral blood mononuclear cells and cortisol between the experimental and control group and of those in the experimental group with a good outcome (i.e., immediate pain relief) with those in the experimental group with a poor outcome.

Adverse events

Serious and non-serious adverse events related to the experimental and control intervention, and all other aspects of the study, will be documented. At the three post-intervention time points, potential adverse events will be recorded using an online survey. Adverse events will be followed-up as needed by an independent clinician. Depending on the nature of the event, participants may be referred to a GP or a medical specialist, and additional tests or procedures may be proposed. The experimental intervention has been

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3 shown to be safe(11, 27) and it is considered unlikely that serious adverse events due to the
4 interventions will occur. Therefore, installing a data monitoring safety board was not
5 requested by the Ethics Committee.
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10 11 12 **Patient and public involvement** 13

14 A panel of four people with persistent neck pain co-developed and evaluated the study
15 design, research questions, choice of experimental and control intervention, and burden of
16 study participation for the participants. Two of these people and two representatives from the
17 public reviewed the Patient information letter and their feedback was used to improve the
18 letter.
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28 **Data management and monitoring** 29

30 The data will be collected at the Department of Rehabilitation of the Amsterdam
31 University Medical Centre, location VUmc, and/or in physiotherapy practices. The collected
32 data will be securely stored at Vrije Universiteit Amsterdam, Faculty of Behavioural and
33 Movement Sciences. All data are de-identified by using unique participant ID numbers in
34 such a way that the data cannot be traced back to the individual participants without the key.
35 The participants code will exist of a random code of three numbers. The electronically key
36 connecting participant names with codes will be kept in a secure location in the principal
37 investigator's office. The key will be kept for six months after the final publication, and will
38 then be destroyed. Data will be stored in a de-identified manner for fifteen years after the final
39 publication.
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Role of funding source

This study is funded by the Dutch Association for Manual Therapy (NVMT, grant ID. Top-down_2018) and by the Faculty of Behavioural and Movement Sciences (grant ID. Lab Fund_2019) of Vrije Universiteit Amsterdam. The MSG Science Network (<https://www.msg-sciencenetwerk.nl/>) (grant ID. N/A) will support participant recruitment. The funding sources have no role in the study design and will not have any roles in data collection, analysis and interpretation of the data, nor in the reporting of the results.

Ethics and dissemination

The study is registered at trialregister.nl with study identification NL6575. The results of the study will be published in peer-reviewed journals and disseminated at conferences, in newsletters and social media. The trial is approved by the Medical Ethics Committee of Amsterdam University Medical Centre, location VUmc (Approval number: 2018.181). All procedures will be conducted in accordance with the Declaration of Helsinki.(54) Amendment to this protocol will be submitted for approval to the Medical Ethical Committee and deviations from the protocol will be reported to the trial registration.

3. DISCUSSION

There is considerable debate in the literature regarding the possibility of meaningful neuroimmune-mediated pain relief following joint mobilisation and manipulation.(2, 7, 55, 56) We described a protocol for a randomised placebo-controlled study that will assess potential neuroimmune-mediated pain relief following joint mobilisation and manipulation in people with persistent neck pain. The aim of this study is to gain insights in the relation

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3 between changes in neuroimmune responses and pain relief, rather than in the clinical efficacy
4 or effectiveness of joint mobilisation and manipulation for people with persistent neck pain.
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8 Recent data suggest that the production of pro-inflammatory cytokines is higher and
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10 production of anti-inflammatory cytokines is lower in patients with persistent-pain compared
11 to healthy people following *in-vitro* stimulation of whole blood cells.(44) Additionally, a
12 specific, coordinated inflammatory processes may be important for patient recovery.(3)
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14 Contrary to the other studies we are aware of that measured neuroimmune responses
15 following joint mobilisation and manipulation,(7, 44, 57) we will assess a comprehensive
16 range of inflammatory markers. Our approach to measure pro-inflammatory cytokines and
17 their antagonists provides insight into the activation of immunocompetent cells.(58)
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21 We believe the design of our study allows to assess the specific effects of joint
22 mobilisation and manipulation on neuroimmune responses. For instance, rather than
23 comparing the joint mobilisation and manipulation with a wait-and-see approach, we will
24 compare responses with a placebo-control intervention that resembles joint mobilisation and
25 manipulation. Additionally, the verbal instructions between the experimental and control
26 groups will be comparable and standardised, which reduces differences in intervention
27 efficacy due to non-specific intervention effects.(59) Differences in verbal instructions have
28 been shown to be associated with differences in endocrine responses following joint
29 manipulation in people with neck pain.(60) Finally, we will record the participant's
30 intervention expectations and beliefs regarding joint mobilisation and manipulation as a
31 treatment method to alleviate neck pain.(42)
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51 Previous research revealed a non-linearity of the VAS to measure pain intensity, that
52 responsiveness varies along the spectrum of pain intensity and the importance of taking
53 baseline pain into account when evaluating change scores.(43, 61, 62) Consequently,
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3 invalid as these will either underestimate or overestimate true change.(62) To overcome this
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5 problem, we follow the initiative on methods, measurement and pain assessment in clinical
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7 trials (IMMPACT) recommendation to identify those with a good, poor outcome, or unclear
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9 outcome.(43)
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12 Besides the strengths, the proposed study has some potential limitations. First, we
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14 assume a linear association between neuroimmune responses and musculoskeletal pain. A
15
16 linear association between neuroimmune responses and musculoskeletal pain is a prerequisite
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18 for the justification of the statistics proposed in this protocol. However, one study suggests
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20 that an initial threshold of neuroimmune responses might be required, which would suggest a
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22 non-linear relationship between neuroimmune responses and musculoskeletal pain.(63) In that
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24 study, elevated IL-6 levels were only present in the group of people with pain > 40/100 VAS
25
26 compared to control.(63) Therefore, a minimal pain intensity of 40/100 on the VAS will be a
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28 prerequisite for participating in this study.
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33 Another limitation is that only a single session of joint mobilisation and manipulation
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35 will be provided together with a short follow-up. While a single session of joint mobilisation
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37 and manipulation may induce a pain-relieving effect,(17) the clinical relevance of
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39 immediately pain relief is unclear. Nonetheless, our aim is not to examine the efficacy of joint
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41 mobilisation and manipulation but rather to understand the biological mechanisms behind
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43 pain relief following joint mobilisation and manipulation. In studying the mechanism of
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45 action, a short follow-up has the advantage that potential confounding variables can be
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47 controlled, such as food intake, stress, physical exercise and health status.
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Contributorship statement

All authors contributed to the design of this protocol. ILS, GGM and MWC initiated the protocol. The protocol was drafted by ILS, GGM, HJB and MWC. Statistical advice was provided by GGM and MWC. ILS, GGM and MWC were responsible for ethical board approval. ILS was responsible for drafting the manuscript. All authors contributed to the manuscript and read and approved the final manuscript.

Competing interests

The authors have no known conflict of interest and have no commercial interest in this study.

Data sharing statement

Individual deidentified participant data that underlie the results will be shared. Investigators whose proposed use of the data had been approved by an independent review committee identified for this purpose can access the data for individual participant data meta-analysis. Data will be available beginning 9 months and ending 36 months following the result article publication. Proposals may be submitted up to 36 months following article publication. After 36 months the data will be available in our University's data warehouse but without investigator support other than deposited metadata. Information regarding submitting proposals and accessing data may be found at <https://research.vu.nl>.

REFERENCES

1. Sawicki CM, Humeidan ML, Sheridan JF. Neuroimmune Interactions in Pain and Stress: An Interdisciplinary Approach. *Neuroscientist*. 2020:1073858420914747.
2. Gold JE, Hallman DM, Hellstrom F, et al. Systematic review of biochemical biomarkers for neck and upper-extremity musculoskeletal disorders. *Scand J Work Environ Health*. 2016;42(2):103-24.
3. Farrell SF, de Zoete R, Cabot PJ, Sterling M. Systemic inflammatory markers in neck pain: a systematic review with meta-analysis. *Eur J Pain*. 2020.
4. Carp SJ, Barbe MF, Winter KA, Amin M, Barr AE. Inflammatory biomarkers increase with severity of upper-extremity overuse disorders. *Clin Sci (Lond)*. 2007;112(5):305-14.
5. Albrecht DS, Ahmed SU, Kettner NW, et al. Neuroinflammation of the spinal cord and nerve roots in chronic radicular pain patients. *Pain*. 2018;159(5):968-77.
6. Loggia ML, Chonde DB, Akeju O, et al. Evidence for brain glial activation in chronic pain patients. *Brain*. 2015;138(Pt 3):604-15.
7. Jungen MJ, Ter Meulen BC, van Osch T, Weinstein HC, Ostelo R. Inflammatory biomarkers in patients with sciatica: a systematic review. *BMC Musculoskelet Disord*. 2019;20(1):156.
8. Sterling M, Elliott JM, Cabot PJ. The course of serum inflammatory biomarkers following whiplash injury and their relationship to sensory and muscle measures: a longitudinal cohort study. *PLoS One*. 2013;8(10):e77903.
9. Roy RA, Boucher JP, Comtois AS. Inflammatory response following a short-term course of chiropractic treatment in subjects with and without chronic low back pain. *J Chiropr Med*. 2010;9(3):107-14.
10. Teodorczyk-Injeyan JA, McGregor M, Triano JJ, Injeyan SH. Elevated Production of Nociceptive CC Chemokines and sE-Selectin in Patients With Low Back Pain and the

- 1
2
3 Effects of Spinal Manipulation: A Nonrandomized Clinical Trial. *Clin J Pain*.
4
5 2018;34(1):68-75.
6
7
8 11. Michalis K, Anastasios P, Theodoros B, Konstantinos Z. The Impact of Manual Therapy
9
10 Techniques on Pain, Disability and II-1b Levels in Patients with Chronic Cervical Pain.
11
12 *International Journal of Physiotherapy*. 2019;6(6):268-76.
13
14
15 12. Martins DF, Mazzardo-Martins L, Gadotti VM, et al. Ankle joint mobilization reduces
16
17 axonotmesis-induced neuropathic pain and glial activation in the spinal cord and
18
19 enhances nerve regeneration in rats. *Pain*. 2011;152(11):2653-61.
20
21
22 13. Santos FM, Silva JT, Giardini AC, et al. Neural mobilization reverses behavioral and
23
24 cellular changes that characterize neuropathic pain in rats. *Mol Pain*. 2012;8:57.
25
26
27 14. Gleeson M, Bishop NC, Stensel DJ, Lindley MR, Mastana SS, Nimmo MA. The anti-
28
29 inflammatory effects of exercise: mechanisms and implications for the prevention and
30
31 treatment of disease. *Nat Rev Immunol*. 2011;11(9):607-15.
32
33
34 15. McGee SL, Hargreaves M. Exercise adaptations: molecular mechanisms and potential
35
36 targets for therapeutic benefit. *Nat Rev Endocrinol*. 2020.
37
38
39 16. Lutke Schipholt IJ, Coppieters MW, Meijer OG, Tompra N, de Vries RBM, Scholten-
40
41 Peeters GGM. Effects of joint and nerve mobilisation on neuroimmune responses in
42
43 animals and humans with neuromusculoskeletal conditions: a systematic review and
44
45 meta-analysis. *Pain Rep*. 2021;6(2):e927.
46
47
48 17. Scholten-Peeters GG, Thoomes E, Konings S, et al. Is manipulative therapy more
49
50 effective than sham manipulation in adults : a systematic review and meta-analysis.
51
52 *Chiropr Man Therap*. 2013;21(1):34.
53
54
55 18. Coulter ID, Crawford C, Vernon H, et al. Manipulation and Mobilization for Treating
56
57 Chronic Nonspecific Neck Pain: A Systematic Review and Meta-Analysis for an
58
59 Appropriateness Panel. *Pain Physician*. 2019;22(2):E55-e70.
60

19. Paige NM, Miake-Lye IM, Booth MS, et al. Association of Spinal Manipulative Therapy With Clinical Benefit and Harm for Acute Low Back Pain: Systematic Review and Meta-analysis. *Jama*. 2017;317(14):1451-60.
20. Bialosky JE, Beneciuk JM, Bishop MD, et al. Unraveling the Mechanisms of Manual Therapy: Modeling an Approach. *J Orthop Sports Phys Ther*. 2018;48(1):8-18.
21. Mintken PE, Rodeghero J, Cleland JA. Manual therapists - Have you lost that loving feeling?! *J Man Manip Ther*. 2018;26(2):53-54.
22. Randoll C, Gagnon-Normandin V, Tessier J, et al. The mechanism of back pain relief by spinal manipulation relies on decreased temporal summation of pain. *Neuroscience*. 2017;349:220-28.
23. Courtney CA, Steffen AD, Fernandez-de-Las-Penas C, Kim J, Chmell SJ. Joint Mobilization Enhances Mechanisms of Conditioned Pain Modulation in Individuals With Osteoarthritis of the Knee. *J Orthop Sports Phys Ther*. 2016;46(3):168-76.
24. Song X, Gan Q, Cao J, Wang Z, Rupert RL. Spinal manipulation reduces pain and hyperalgesia after lumbar intervertebral foramen inflammation in the rat. *Journal of Manipulative & Physiological Therapeutics*. 2006;29(1):5-13.
25. Song XJ, Huang ZJ, Song WB, et al. Attenuation Effect of Spinal Manipulation on Neuropathic and Postoperative Pain Through Activating Endogenous Anti-Inflammatory Cytokine Interleukin 10 in Rat Spinal Cord. *Journal of Manipulative and Physiological Therapeutics*. 2016;39(1):42-53.
26. Bialosky JE, Bishop MD, Penza CW. Placebo Mechanisms of Manual Therapy: A Sheep in Wolf's Clothing? *J Orthop Sports Phys Ther*. 2017;47(5):301-04.
27. Teodorczyk-Injeyan JA, Injeyan HS, Ruegg R. Spinal manipulative therapy reduces inflammatory cytokines but not substance P production in normal subjects. *J Manipulative Physiol Ther*. 2006;29(1):14-21.

- 1
2
3 28. Lutke Schipholt IJ, Scholten-Peeters GGM, Bontkes HJ, Coppieters MW. Multiple
4
5 confounders influence the association between low-grade systemic inflammation and
6
7 musculoskeletal pain. A call for a prudent interpretation of the literature. *Spine J*.
8
9 2018;18(11):2162-63.
10
11
- 12 29. Chan AW, Tetzlaff JM, Altman DG, et al. SPIRIT 2013 statement: defining standard
13
14 protocol items for clinical trials. *Ann Intern Med*. 2013;158(3):200-7.
15
16
- 17 30. Cuschieri S. The CONSORT statement. *Saudi J Anaesth*. 2019;13(Suppl 1):S27-s30.
18
- 19 31. Hoffmann TC, Glasziou PP, Boutron I, et al. Better reporting of interventions: template
20
21 for intervention description and replication (TIDieR) checklist and guide. *Bmj*.
22
23 2014;348:g1687.
24
25
- 26 32. Haldeman S, Carroll L, Cassidy JD, Schubert J, Nygren Å. The Bone and Joint Decade
27
28 2000–2010 Task Force on Neck Pain and Its Associated Disorders: Executive Summary.
29
30 *Spine*. 2008;33(4S):S5-S7.
31
32
- 33 33. Hutting N, Kerry R, Coppieters MW, Scholten-Peeters GGM. Considerations to improve
34
35 the safety of cervical spine manual therapy. *Musculoskelet Sci Pract*. 2018;33:41-45.
36
37
- 38 34. Rushton A, Rivett D, Carlesso L, Flynn T, Hing W, Kerry R. International framework for
39
40 examination of the cervical region for potential of Cervical Arterial Dysfunction prior to
41
42 Orthopaedic Manual Therapy intervention. *Man Ther*. 2014;19(3):222-8.
43
44
- 45 35. Bier JD, Scholten-Peeters WGM, Staal JB, et al. Clinical Practice Guideline for Physical
46
47 Therapy Assessment and Treatment in Patients With Nonspecific Neck Pain. *Phys Ther*.
48
49 2018;98(3):162-71.
50
51
- 52 36. Finucane LM, Downie A, Mercer C, et al. International Framework for Red Flags for
53
54 Potential Serious Spinal Pathologies. *J Orthop Sports Phys Ther*. 2020;50(7):350-72.
55
56
- 57 37. Jull G, Moore A, Falla D, Lewis J, McCarthy C, Sterling M. *Grieve's Modern*
58
59 *Musculoskeletal Physiotherapy*: Elsevier; 2015.
60

- 1
2
3 38. Manning DM, Dedrick GS, Sizer PS, Brismee JM. Reliability of a seated three-
4
5 dimensional passive intervertebral motion test for mobility, end-feel, and pain
6
7 provocation in patients with cervicgia. *J Man Manip Ther.* 2012;20(3):135-41.
8
9
- 10 39. van Trijffel E, Anderegg Q, Bossuyt PM, Lucas C. Inter-examiner reliability of passive
11
12 assessment of intervertebral motion in the cervical and lumbar spine: a systematic review.
13
14 *Man Ther.* 2005;10(4):256-69.
15
16
- 17 40. van der El A. *Orthopaedic Manual Therapy Diagnosis: Spine and Temporomandibular*
18
19 *Joints.* Gartside M, editor: Jones and Barlett Publishers; 2010.
20
21
- 22 41. Licciardone JC, Russo DP. Blinding protocols, treatment credibility, and expectancy:
23
24 methodologic issues in clinical trials of osteopathic manipulative treatment. *J Am*
25
26 *Osteopath Assoc.* 2006;106(8):457-63.
27
28
- 29 42. Fulda KG, Slichon T, Stoll ST. Patient expectations for placebo treatments commonly used
30
31 in osteopathic manipulative treatment (OMT) clinical trials: a pilot study. *Osteopath Med*
32
33 *Prim Care.* 2007;1:3.
34
35
- 36 43. Dworkin RH, Turk DC, Wyrwich KW, et al. Interpreting the clinical importance of
37
38 treatment outcomes in chronic pain clinical trials: IMMPACT recommendations. *J Pain.*
39
40 2008;9(2):105-21.
41
42
- 43 44. Teodorczyk-Injeyan JA, Triano JJ, Injeyan HS. Nonspecific Low Back Pain:
44
45 Inflammatory Profiles of Patients With Acute and Chronic Pain. *Clin J Pain.*
46
47 2019;35(10):818-25.
48
49
- 50 45. Segre E, Fullerton JN. Stimulated Whole Blood Cytokine Release as a Biomarker of
51
52 Immunosuppression in the Critically Ill: The Need for a Standardized Methodology.
53
54 *Shock.* 2016;45(5):490-4.
55
56
- 57 46. Teodorczyk-Injeyan JA, Triano JJ, McGregor M, Woodhouse L, Injeyan HS. Effect of
58
59 Interactive Neurostimulation Therapy on Inflammatory Response in Patients With
60

- 1
2
3 Chronic and Recurrent Mechanical Neck Pain. *J Manipulative Physiol Ther.*
4
5 2015;38(8):545-54.
6
7
8 47. Teodorczyk-Injeyan JA, Triano JJ, McGregor M, Woodhouse L, Injeyan HS. Elevated
9
10 production of inflammatory mediators including nociceptive chemokines in patients with
11
12 neck pain: a cross-sectional evaluation. *J Manipulative Physiol Ther.* 2011;34(8):498-
13
14 505.
15
16
17 48. Kwok YH, Hutchinson MR, Gentall MG, Rolan PE. Increased responsiveness of
18
19 peripheral blood mononuclear cells to in vitro TLR 2, 4 and 7 ligand stimulation in
20
21 chronic pain patients. *PLoS One.* 2012;7(8):e44232.
22
23
24 49. Habbal OA, Al-Jabri AA. Circadian rhythm and the immune response: a review. *Int Rev*
25
26 *Immunol.* 2009;28(1):93-108.
27
28
29 50. van Eeden WA, van Hemert AM, Carlier IVE, et al. Basal and LPS-stimulated
30
31 inflammatory markers and the course of individual symptoms of depression. *Transl*
32
33 *Psychiatry.* 2020;10(1):235.
34
35
36 51. Generaal E, Vogelzangs N, Macfarlane GJ, et al. Basal inflammation and innate immune
37
38 response in chronic multisite musculoskeletal pain. *Pain.* 2014;155(8):1605-12.
39
40
41 52. Twisk JWR. *Inleiding in de toegepaste biostatistiek*: Bohn Stafleu van Loghum; 2016.
42
43
44 53. Lutke Schipholt IJ, Scholten-Peeters GGM, Bontkes HJ, Coppieters MW. Authors'
45
46 Reply: Confounding and mediation to reveal the true association between systemic
47
48 inflammation and musculoskeletal pain. *Spine J.* 2019;19(11):1901.
49
50
51 54. World Medical Association Declaration of Helsinki: ethical principles for medical
52
53 research involving human subjects. *Jama.* 2013;310(20):2191-4.
54
55
56 55. van den Berg R, Jongbloed EM, de Schepper EIT, Bierma-Zeinstra SMA, Koes BW,
57
58 Luijsterburg PAJ. The association between pro-inflammatory biomarkers and nonspecific
59
60 low back pain: a systematic review. *Spine J.* 2018;18(11):2140-51.

- 1
2
3 56. Morris P, Ali K, Merritt M, Pelletier J, Macedo LG. A systematic review of the role of
4
5 inflammatory biomarkers in acute, subacute and chronic non-specific low back pain.
6
7 BMC Musculoskelet Disord. 2020;21(1):142.
8
9
10 57. Klyne DM, Hodges PW. Letter to the editor concerning “Multiple confounders influence
11
12 the association between low-grade systemic inflammation and musculoskeletal pain. A
13
14 call for a prudent interpretation of the literature”. Spine J. 2019.
15
16
17 58. Li Y, Liu J, Liu ZZ, Duan DP. Inflammation in low back pain may be detected from the
18
19 peripheral blood: suggestions for biomarker. Biosci Rep. 2016;36(4).
20
21
22 59. Rossetini G, Camerone EM, Carlino E, Benedetti F, Testa M. Context matters: the
23
24 psychoneurobiological determinants of placebo, nocebo and context-related effects in
25
26 physiotherapy. Archives of physiotherapy. 2020;10:11-11.
27
28
29 60. Malfliet A, Lluch Girbés E, Pecos-Martin D, Gallego-Izquierdo T, Valera-Calero A. The
30
31 Influence of Treatment Expectations on Clinical Outcomes and Cortisol Levels in
32
33 Patients With Chronic Neck Pain: An Experimental Study. Pain Pract. 2019;19(4):370-
34
35 81.
36
37
38 61. Emshoff R, Bertram S, Emshoff I. Clinically important difference thresholds of the visual
39
40 analog scale: a conceptual model for identifying meaningful intraindividual changes for
41
42 pain intensity. Pain. 2011;152(10):2277-82.
43
44
45 62. Kersten P, White PJ, Tennant A. Is the pain visual analogue scale linear and responsive to
46
47 change? An exploration using Rasch analysis. PloS one. 2014;9(6):e99485-e85.
48
49
50 63. Klyne DM, Barbe MF, Hodges PW. Systemic inflammatory profiles and their
51
52 relationships with demographic, behavioural and clinical features in acute low back pain.
53
54 Brain Behav Immun. 2017;60:84-92.
55
56
57
58
59
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- 1
2
3 64. Jorritsma W, de Vries GE, Dijkstra PU, Geertzen JH, Reneman MF. Neck Pain and
4
5 Disability Scale and Neck Disability Index: validity of Dutch language versions. *Eur*
6
7 *Spine J.* 2012;21(1):93-100.
8
9
- 10 65. Kamper SJ, Ostelo RW, Knol DL, Maher CG, de Vet HC, Hancock MJ. Global Perceived
11
12 Effect scales provided reliable assessments of health transition in people with
13
14 musculoskeletal disorders, but ratings are strongly influenced by current status. *J Clin*
15
16 *Epidemiol.* 2010;63(7):760-66.e1.
17
18
- 19 66. Sleijser-Koehorst MLS, Bijker L, Cuijpers P, Scholten-Peeters GGM, Coppieters MW.
20
21 Preferred self-administered questionnaires to assess fear of movement, coping, self-
22
23 efficacy, and catastrophizing in patients with musculoskeletal pain-A modified Delphi
24
25 study. *Pain.* 2019;160(3):600-06.
26
27
- 28 67. Freynhagen R, Baron R, Gockel U, Tolle TR. painDETECT: a new screening
29
30 questionnaire to identify neuropathic components in patients with back pain. *Curr Med*
31
32 *Res Opin.* 2006;22(10):1911-20.
33
34
- 35 68. Neblett R, Cohen H, Choi Y, et al. The Central Sensitization Inventory (CSI):
36
37 establishing clinically significant values for identifying central sensitivity syndromes in
38
39 an outpatient chronic pain sample. *J Pain.* 2013;14(5):438-45.
40
41
- 42 69. Bijker L, Sleijser-Koehorst MLS, Coppieters MW, Cuijpers P, Scholten-Peeters GGM.
43
44 Preferred Self-Administered Questionnaires to Assess Depression, Anxiety and
45
46 Somatization in People With Musculoskeletal Pain - A Modified Delphi Study. *J Pain.*
47
48 2019.
49
50
- 51 70. Blikman T, Stevens M, Bulstra SK, van den Akker-Scheek I, Reininga IH. Reliability and
52
53 validity of the Dutch version of the International Physical Activity Questionnaire in
54
55 patients after total hip arthroplasty or total knee arthroplasty. *J Orthop Sports Phys Ther.*
56
57 2013;43(9):650-9.
58
59
60

- 1
2
3 71. Poppel MNMv, Paw MCA, Mechelen WV, editors. Reproduceerbaarheid en validiteit
4 van de Nederlandse versie van de International Physical Activity Questionnaire
5 (IPAQ)2004.
6
7
8
9
10 72. Buysse DJ, Reynolds CF, Monk TH, Berman SR, Kupfer DJ. The Pittsburgh sleep
11 quality index: A new instrument for psychiatric practice and research. *Psychiatry*
12 *Research*. 1989;28(2):193-213.
13
14
15
16 73. Kovacs FM, Abaira V, Royuela A, et al. Minimum detectable and minimal clinically
17 important changes for pain in patients with nonspecific neck pain. *BMC Musculoskelet*
18 *Disord*. 2008;9:43.
19
20
21
22
23 74. Cuijpers P, Smits N, Donker T, ten Have M, de Graaf R. Screening for mood and anxiety
24 disorders with the five-item, the three-item, and the two-item Mental Health Inventory.
25 *Psychiatry Research*. 2009;168(3):250-55.
26
27
28
29 75. Hoeymans N, Garssen AA, Westert GP, Verhaak PF. Measuring mental health of the
30 Dutch population: a comparison of the GHQ-12 and the MHI-5. *Health Qual Life*
31 *Outcomes*. 2004;2:23.
32
33
34
35 76. Fletcher JP, Bandy WD. Intrarater reliability of CROM measurement of cervical spine
36 active range of motion in persons with and without neck pain. *J Orthop Sports Phys Ther*.
37 2008;38(10):640-5.
38
39
40
41 77. Walton DM, Macdermid JC, Nielson W, Teasell RW, Chiasson M, Brown L. Reliability,
42 standard error, and minimum detectable change of clinical pressure pain threshold testing
43 in people with and without acute neck pain. *J Orthop Sports Phys Ther*. 2011;41(9):644-
44 50.
45
46
47
48 78. Rolke R, Baron R, Maier C, et al. Quantitative sensory testing in the German Research
49 Network on Neuropathic Pain (DFNS): standardized protocol and reference values. *Pain*.
50 2006;123(3):231-43.
51
52
53
54
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2
3 79. Schlecht I, Wiggermann P, Behrens G, et al. Reproducibility and validity of ultrasound
4
5 for the measurement of visceral and subcutaneous adipose tissues. *Metabolism*.
6
7 2014;63(12):1512-9.
8
9
10 80. Park HS, Park JY, Yu R. Relationship of obesity and visceral adiposity with serum
11
12 concentrations of CRP, TNF-alpha and IL-6. *Diabetes Res Clin Pract*. 2005;69(1):29-35.
13
14 81. Bouman A, Moes H, Heineman MJ, de Leij LF, Faas MM. The immune response during
15
16 the luteal phase of the ovarian cycle: increasing sensitivity of human monocytes to
17
18 endotoxin. *Fertil Steril*. 2001;76(3):555-9.
19
20
21 82. Myriantefs P, Karatzas S, Venetsanou K, et al. Seasonal variation in whole blood
22
23 cytokine production after LPS stimulation in normal individuals. *Cytokine*.
24
25 2003;24(6):286-92.
26
27
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3 **Figure 1:** Anticipated flow of the study
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6 **Abbreviations:** GPE: global perceived effect; VAS: visual analogue scale
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3 **Figure 2.** Spinal mobilisation and manipulation techniques.
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6 Depending on the identified painful segmental levels, the clinician can select from different
7 cervical mobilisation techniques (A-C); For techniques A-C, the participant will be seated on
8 a chair, leaning against the upper leg or shoulder of the clinician. **Panel A:** Mobilisation
9 targeting the atlanto-axial joints. The cervical segments below the second cervical vertebrae
10 are submaximal rotated and lateroflexed. With the clinician's hypothenar region of the hand
11 over the structures overlying the arcus of the first vertebrae, the clinician moved the head
12 further in rotation.(40) **Panel B:** Segmental zygapophyseal joint mobilisation (C2 to C7; the
13 image shows the technique for C3-C4). First, the occipital-atlanto-axial joint is maximally
14 rotated in the direction of the facet joint being mobilised. Subsequently, the head is moved to
15 extension, ipsilateral lateroflexion and rotation until pressure from the thumb is felt. This
16 technique is repeated on the lower level until the painful cervical segment is reached (C3-C4).
17 Next, on the painful cervical segment, pressure will be given in a cranio-ventral direction.
18 (40) **Panel C:** Mobilisation technique targeting the occipital-atlanto-axial joints. The
19 clinician's hypothenar region is placed against the mastoid process. C2 to C7 are
20 submaximally locked in flexion, rotation and lateroflexion. The head is then moved in a
21 medio-caudal direction.(40) **Panel D:** Spinal manipulation technique targeting the cervico-
22 thoracic junction. The participant will be seated on a treatment table. The height of the table
23 will be adjusted to the level of the clinician's abdomen. The participant's hands will be placed
24 on the back of their head (with one hand placed over the other hand, rather than with
25 interlocking fingers), and with the shoulders slightly retracted. The clinician's hands will be
26 placed over the hands of the participant, with the clinician's forearms ventral to the shoulder
27 of the participant. Then, a high-velocity, low-amplitude movement will be applied in a dorsal-
28 cranial direction.(40) Green arrows represent the direction of the mobilisation (Panel A-C) or
29 manipulation (Panel D).
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Table 1: Overview of the neuroimmune responses

Domain	Neuroimmune parameters	Timing of measurements			
		T0	T1	T2	T3
Systemic inflammatory marker directly from blood samples ^a	TNF- α , TNF-RII, IL-1 β , IL-1RA, hsCRP ^b	√	√	√	-
Inflammatory marker concentration after <i>in-vitro</i> stimulation of whole blood cells ^c	TNF- α , IL-1 β , IL-1RA, IL-4, IL-10, CCL2, CCL3, CCL4	√	√	√	-
Ex-vivo serum cortisol ^d	Cortisol	√	√	-	-
Phenotypic analysis of peripheral blood mononuclear cells ^e	CD45 ⁺ , CD3 ⁺ , CD4 ⁺ , CD25 ^{hi} , CD8 ⁺ , CD56 ⁺ , CD19 ⁺ , CD14 ⁺ , HLA-DR, TLR-4	√	-	√	-

a) Measured using multianalyte assay Ella (R&D systems, Minneapolis, United States)

b) Cardiac C-Reactive Protein (Latex) High Sensitive using Roche/Hitachi cobas c systems.

c) Stimulated for 24 hours at 37°C, in a humidified 5% CO₂ incubator, with lipopolysaccharide (LPS) from *Escherichia coli* O55:B5 at a concentration of 1ng/ml and 10 μ g/ml. Determined using a custom-made U-plex (MSD, Maryland, United States)

d) Using conventional electrochemiluminescence immunoassay (ECLIA), Roche (Cobas Cortisol, 2nd generation).

e) Determined by 10-color flowcytometry (FCM): CD45⁺ = General Leukocyte marker; CD3⁺ = T-cell marker; CD3⁺CD4⁺ = CD4⁺ T-helper marker; CD3⁺CD4⁺CD25^{hi} = T-regulator cell marker; CD3⁺CD8⁺ = Cytotoxic T-cell marker; CD3⁺CD56⁺ = Natural Killer cell marker; CD19⁺ = B-cell marker; CD14⁺ = monocyte marker; HLA-DR = activation marker for T-cells and monocytes; TLR-4 = Toll-like receptor 4 marker.

Abbreviations: T0: baseline; T1: immediately following the intervention; T2: two-hours following the intervention; T3: two-days following the intervention; TNF- α : Tumor Necrosis Factor- α ; TNF-RII: Tumor Necrosis Factor Receptor Antagonist 2; IL-1 β : Interleukin-1 β ; IL-1RA: Interleukin-1 receptor antagonist; hsCRP: High sensitive C-Reactive Protein; IL-4: Interleukin-4; IL-10: Interleukin-10; CCL2: c-c-motif chemokine ligand 2; CCL3: c-c-motif chemokine ligand 3; CCL4: c-c-motif chemokine ligand 4; CD: Cluster of Differentiation

Table 2: Self-reported questionnaires and physical tests

Domain	Self-reported questionnaires	Timing of measurements			
		T0	T1	T2	T3
Disability	Neck Disability Index (NDI) ^a	-	√	-	-
Perceived effect	Global Perceived Effect (GPE) ^b	-	-	√	√
Fear of movement	Tampa Scale of Kinesiophobia ^c	-	√	-	-
Type of pain	PAIN Detect Questionnaire (PDQ) ^d	-	√	-	-
Type of pain	Central Sensitisation Inventory (CSI) ^e	-	√	-	-
Depression, Anxiety, Stress	Depression Anxiety Stress Scale (DASS21) ^f	-	√	-	-
Physical activity	International Physical Activity Questionnaire (IPAQ) ^g	-	√	-	-
Catastrophising	Pain Catastrophising Scale (PCS) ^h	-	√	-	-
Sleep Quality	Pittsburgh Sleep Quality Index (PSQI) ⁱ	-	√	-	-
Pain Intensity	Visual Analogue Scale (VAS) ^j	√	√	√	√
Mental health	Mental health inventory (MHI-5) ^k	√	-	-	-

Domain	Physical tests	Timing of measurement			
		T0	T1	T2	T3
Range of motion	Cervical Range of Motion (CROM) ^l	√	√	√	-
Pain intensity	CROM-VAS test ^m	-	√	√	-
Quantitative sensory testing	Pressure Pain Threshold (PPT) ⁿ	√	√	√	-
Quantitative sensory testing	Wind-up ratio ^o	√	√	√	-

a) The Dutch version of the NDI is a valid and responsive measure of disability.(64)

b) The GPE is a validated and reliable tool to assess health transitions in patients with musculoskeletal disorders.(65)

c) Preferred self-administrated questionnaire to asses fear of movement in musculoskeletal pain.(66)

d) Persistent pain will be categorised in two-mechanism based groups: nociceptive and neuropathic pain using the PDQ. The PD-Q is a reliable screening tool with high specificity.(67)

e) The Dutch Central Sensitization Inventory (CSI) has good internal consistency, good discriminative power and excellent test-retest reliability. A cut-off score of 40/100 provides a sensitivity of 81% and specificity of 75%.(68)

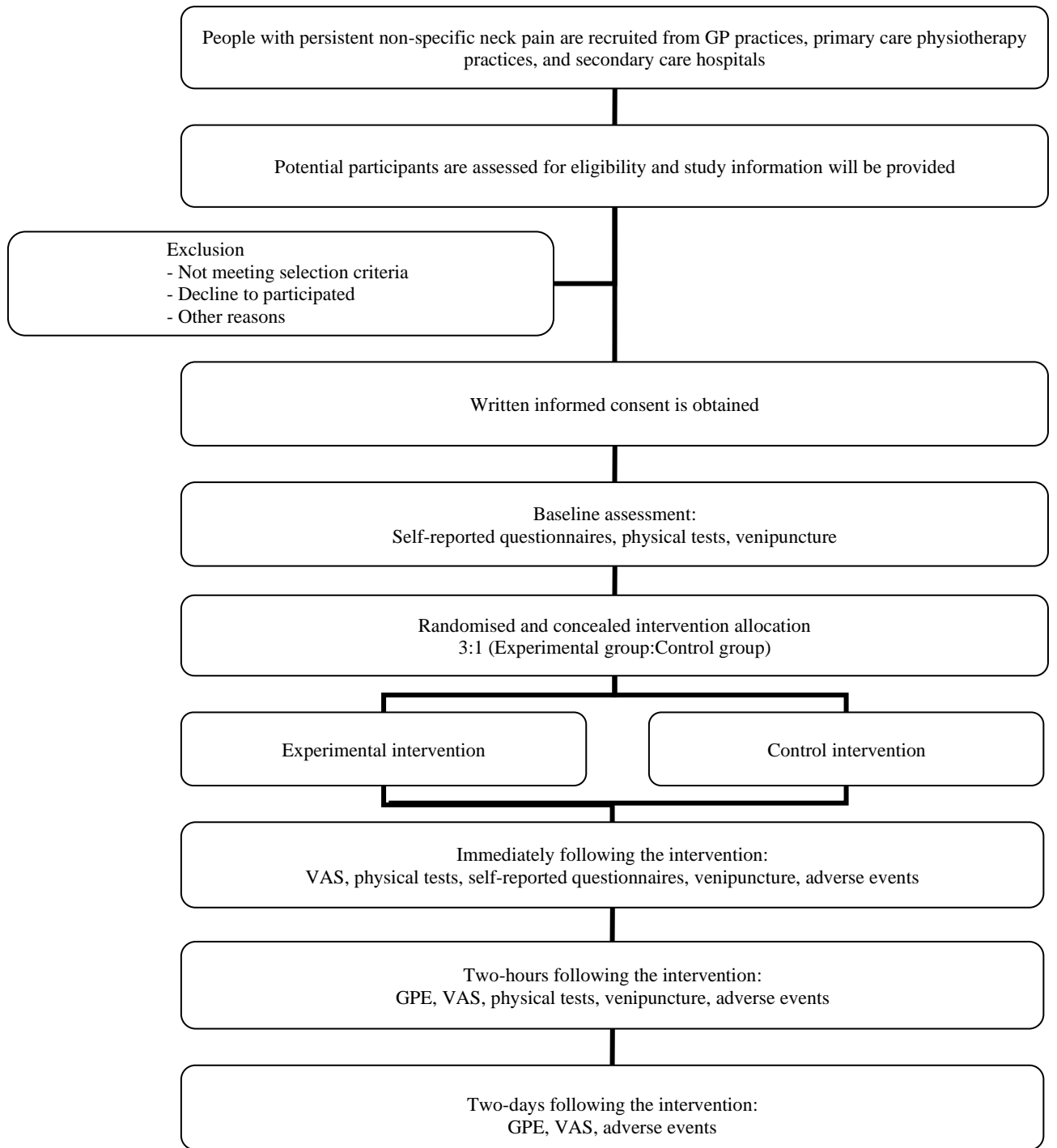
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3 f) Preferred self-administrated questionnaire to assess depression, anxiety and stress in
4 musculoskeletal pain.(66, 69)
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6 g) Expressed in 1000 metabolic equivalent minutes per week (Dutch-language
7 version).(70) The IPAQ has good reliability (intraclass correlation coefficient [ICC] =
8 0.70-0.96) and moderate validity ($r = 0.36-0.49$) of the IPAQ compared with an
9 accelerometer.(71)
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11 h) Preferred self-administrated questionnaire to assess pain catastrophising in
12 musculoskeletal pain.(66)
13
14 i) Score above 5 yield a sensitivity of 89.6% and specificity of 86.5% in distinguishing
15 good and poor sleepers.(72)
16
17 j) The reliability and validity of the VAS as a measure of pain for neck pain patients is
18 good.(73)
19
20 k) General psychological status will be assessed using the MHI-5.(74) A higher score
21 indicates better mental health. Cronbach's alpha for the MHI-5 scale is 0.85.(75)
22
23 l) The CROM is a clinically reliable tool to measure active cervical range of motion
24 people with neck pain and healthy participants.(76)
25
26 m) This novel test consists of two parts. In Part 1, the participant is asked to perform
27 maximal active right and left cervical rotation and the degrees of rotation are reordered
28 using the CROM device. In this position, the pain intensity is measured with the VAS
29 following intervention. After the intervention, Part 2 of the test is performed . The
30 participant is again asked to actively rotate (left and right) to the same position as in
31 Part 1 and the pain intensity is recorded. The difference on VAS scores is the outcome
32 of the CROM-VAS test.
33
34 n) Pressure algometry over the cervical spine has shown excellent intrarater and good-to-
35 excellent interrater reliability in individuals with acute neck pain.(77) This study
36 reported that the MDC for PPT over the cervical spine and tibialis anterior muscle in
37 patients with acute neck pain was 47.2 and 97.9 kPa, respectively.(77) To determine
38 changes in widespread pressure pain sensitivity, PPTs will be assessed bilaterally over
39 the mid-point trapezius (pars descendens), second metacarpal, and tibialis anterior
40 muscle.
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42 o) Using a pinprick 256 mN wind up ratio will be calculated bilaterally over the mid-
43 point trapezius (pars descendens) and tibiales anterior muscle.(78)
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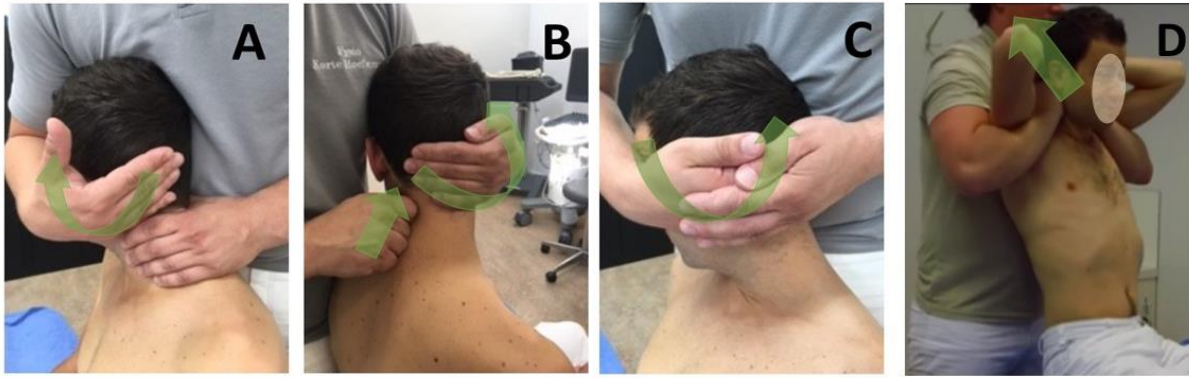
47 **Abbreviations:** T0: baseline; T1: immediately following the intervention; T2: two-hours
48 following the intervention; T3: two-days following the intervention
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Table 3: Potential confounding variables that will be assessed

Potential confounding variables	
Co-morbidities	Number of co-morbidities
Alcohol use	- Non-drinker - Moderate drinker (women: 1-14 glasses/week) (men: 1-21 glasses/week) - Heavy drinker (women: >14 glasses/week) (men: >21 glasses/week)
Smoking	- Never smoked - Former smoker - Current smoker
Body Mass Index	BMI calculated by dividing body weight (kg) by height (m ²)
Medication use	Type and number of medications used
Drugs use	Recreational drugs use - Yes - No
Visceral Adipose Tissue(79, 80)	Linear distance between abdominal peritoneum and ventral aspect of vertebrae will be assessed using ultrasonography
Physical activity	International Physical Activity Questionnaire, expressed in 1000 metabolic equivalent minutes per week (Dutch version)
Menstrual cycle(81)	Regular menstrual cycle (yes/no), whether women are in the luteal or follicular stage (yes/no), menopause (yes/no) and post menopause (yes/no)
Season(82)	Timing of experiment (summer, autumn, spring or winter)
Age	Age in years
Psychological status(74)	Mental health inventory-5
Intervention expectations(42)	The extent to which they agree (using a four-point Likert scale) with four statements (Appendix C)

Abbreviation: BMI: Body Mass Index





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Appendix A: Inflammatory indices

The inflammatory indices will be calculated as follows: The *ex-vivo* and *in-vitro* inflammatory markers will be z-score standardised, summed and divided by the total number of inflammatory markers used. Z-score standardisation for the experimental intervention and control intervention will be obtained using the T0-mean and T0-standard deviation of the control intervention. Z-score standardisation for those in the experimental group with a good outcome and those in the experimental group with a poor outcome will be obtained using the T0-mean and T0-standard deviation of the poor outcome group. Inflammatory markers will be Ln-transformed in the case of non-normality. The following calculations will be used to determine the separate indices:

$$ex-vivo \text{ inflammatory index} = \frac{zTNF\alpha + zTNFR2 + zIL1\beta + zIL1RA + zhsCRP}{5}$$

$$ex-vivo \text{ pro inflammatory index} = \frac{zTNF\alpha + zIL1\beta + zhsCRP}{3}$$

$$ex-vivo \text{ anti inflammatory index} = \frac{zTNFR2 + zIL1RA}{2}$$

$$ex-vivo \text{ ratio } \frac{\text{pro}}{\text{anti}} \text{ inflammatory index} = \frac{ex-vivo \text{ pro inflammatory index}}{ex-vivo \text{ anti inflammatory index}}$$

$$in-vitro \text{ inflammatory index} = \frac{zTNF\alpha + zIL1\beta + zIL1RA + zIL4 + zIL10 + zCCL2 + zCCL3 + zCCL4 + zhsCRP}{9}$$

$$in-vitro \text{ pro inflammatory index} = \frac{zTNF\alpha + zIL1\beta + zCCL2 + zCCL3 + zCCL4 + zhsCRP}{6}$$

$$in-vitro \text{ anti inflammatory index} = \frac{zIL1RA + zIL4 + zIL10}{3}$$

$$in-vitro \text{ ratio } \frac{\text{pro}}{\text{anti}} \text{ inflammatory index} = \frac{in-vitro \text{ pro inflammatory index}}{in-vitro \text{ anti inflammatory index}}$$

Appendix B: Patient informed consent form

- *Ik heb de informatiebrief gelezen. Ook kon ik vragen stellen. Mijn vragen zijn voldoende beantwoord. Ik had genoeg tijd om te beslissen of ik meedoe.*
- *Ik weet dat meedoen vrijwillig is. Ook weet ik dat ik op ieder moment kan beslissen om toch niet mee te doen of te stoppen met het onderzoek. Daarvoor hoef ik geen reden te geven.*
- *Ik geef toestemming voor het informeren van mijn huisarts dat ik meedoe aan dit onderzoek.*
- *Ik geef toestemming voor het verzamelen en gebruiken van mijn gegevens en bloedmonsters voor de beantwoording van de onderzoeksvraag in dit onderzoek.*
- *Ik weet dat voor de controle van het onderzoek sommige mensen toegang tot al mijn gegevens kunnen krijgen. Die mensen staan vermeld in deze informatiebrief. Ik geef toestemming voor die inzage door deze personen.*
- *Ik geef toestemming voor het informeren van mijn huisarts en/of behandelend specialist van onverwachte bevindingen die van belang (kunnen) zijn voor mijn gezondheid.*
- *Ik geef toestemming dat mijn huisarts mij mag informeren over onverwachte bevindingen die van belang (kunnen) zijn voor mijn gezondheid.*
- *Ik geef toestemming om mijn gegevens nog 15 jaar na dit onderzoek te bewaren. Mogelijk kan dit later nog voor [ander/meer] onderzoek worden gebruikt.*
- *Ik ben me ervan bewust dat de gegeven interventie geen vervanging biedt voor een volledig manueel therapeutische behandeling*
- *Ik geef* **wel**
 geen
Toestemming om drie extra bloedsamples (3 keer 5 ml) af te nemen en te bewaren (5 jaar) en om dit later nog voor ander/meer onderzoek te gebruiken, zoals in de informatiebrief staat.
- *Ik wil* **wel**
 niet
Geïnformeerd worden over de uitkomsten van het onderzoek. Dit is informatie over het hele onderzoek en niet specifiek op mij toegespitst.

Toestemmingsformulier

Voor- en achternaam: _____

Straatnaam en huisnummer: _____

Postcode en Woonplaats: _____

E-mailadres:

Handtekening: _____ Datum: __/__/__

In te vullen door de uitvoerende onderzoeker

- *Ik verklaar dat ik deze proefpersoon volledig heb geïnformeerd over het genoemde onderzoek.*
- *Als er tijdens het onderzoek informatie bekend wordt die de toestemming van de proefpersoon zou kunnen beïnvloeden, dan breng ik hem/haar daarvan tijdig op de hoogte.*
- *De proefpersoon krijgt een volledige informatiebrief mee, samen met een kopie van het getekende toestemmingsformulier.*

Naam onderzoeker: _____ Datum : __/__/__

Handtekening: _____

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3 **Appendix C:** Four statements regarding the intervention expectation of the participants
4
5 (modified from(41))
6

7 I believe this intervention will allow me to get better quicker. Strongly agree
8 Agree
9 Disagree
10 Strongly disagree
11
12

13
14 I believe this intervention will decrease my neck pain. Strongly agree
15 Agree
16 Disagree
17 Strongly disagree
18
19

20
21 I believe this intervention will make me more able to do the things I want to do.
22 Strongly agree
23 Agree
24 Disagree
25 Strongly disagree
26
27

28
29 This seems like a logical way to treat neck pain. Strongly agree
30 Agree
31 Disagree
32 Strongly disagree
33
34
35

The TIDieR (Template for Intervention Description and Replication) Checklist*:

Information to include when describing an intervention and the location of the information

Item number	Item	Where located **	
		Primary paper (page or appendix number)	Other † (details)
1.	BRIEF NAME Provide the name or a phrase that describes the intervention.	4 & P.6	_____
2.	WHY Describe any rationale, theory, or goal of the elements essential to the intervention.	4	_____
3.	WHAT Materials: Describe any physical or informational materials used in the intervention, including those provided to participants or used in intervention delivery or in training of intervention providers. Provide information on where the materials can be accessed (e.g. online appendix, URL).	6, Figure 2 & 3	_____
4.	Procedures: Describe each of the procedures, activities, and/or processes used in the intervention, including any enabling or support activities.	4-6	_____
5.	WHO PROVIDED For each category of intervention provider (e.g. psychologist, nursing assistant), describe their expertise, background and any specific training given.	4	_____
6.	HOW Describe the modes of delivery (e.g. face-to-face or by some other mechanism, such as internet or telephone) of the intervention and whether it was provided individually or in a group.	4-6	_____
7.	WHERE Describe the type(s) of location(s) where the intervention occurred, including any necessary infrastructure or relevant features.	3	_____

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WHEN and HOW MUCH

8. Describe the number of times the intervention was delivered and over what period of time including the number of sessions, their schedule, and their duration, intensity or dose.

TAILORING

9. If the intervention was planned to be personalised, titrated or adapted, then describe what, why, when, and how.

MODIFICATIONS

10.* If the intervention was modified during the course of the study, describe the changes (what, why, when, and how).

HOW WELL

11. Planned: If intervention adherence or fidelity was assessed, describe how and by whom, and if any strategies were used to maintain or improve fidelity, describe them.

12.* Actual: If intervention adherence or fidelity was assessed, describe the extent to which the intervention was delivered as planned.

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2, P4-6

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** **Authors** - use N/A if an item is not applicable for the intervention being described. **Reviewers** – use ‘?’ if information about the element is not reported/not sufficiently reported.

† If the information is not provided in the primary paper, give details of where this information is available. This may include locations such as a published protocol or other published papers (provide citation details) or a website (provide the URL).

‡ If completing the TIDieR checklist for a protocol, these items are not relevant to the protocol and cannot be described until the study is complete.

* We strongly recommend using this checklist in conjunction with the TIDieR guide (see *BMJ* 2014;348:g1687) which contains an explanation and elaboration for each item.

* The focus of TIDieR is on reporting details of the intervention elements (and where relevant, comparison elements) of a study. Other elements and methodological features of studies are covered by other reporting statements and checklists and have not been duplicated as part of the TIDieR checklist. When a **randomised trial** is being reported, the TIDieR checklist should be used in conjunction with the CONSORT statement (see www.consort-statement.org) as an extension of **Item 5 of the CONSORT 2010 Statement**. When a **clinical trial protocol** is being reported, the TIDieR checklist should be used in conjunction with the SPIRIT statement as an extension of **Item 11 of the SPIRIT 2013 Statement** (see www.spirit-statement.org). For alternate study designs, TIDieR can be used in conjunction with the appropriate checklist for that study design (see www.equator-network.org).



SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	ItemNo	Description	Page / line number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	Page 1; lines 1-2
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	Page 3; lines 3-6
	2b	All items from the World Health Organization Trial Registration Data Set	
Protocol version	3	Date and version identifier	Page 3; line 4
Funding	4	Sources and types of financial, material, and other support	Page 18; lines 10-12
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	Page 1; lines 4-17 Page 18; lines 1-4
	5b	Name and contact information for the trial sponsor	Page 1; lines 26-35
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	Page 15; lines 1-7
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	Page 15; lines 10-16

Introduction

Background and rationale

6a Description of research question and justification for undertaking the trial including summary of relevant studies (published and unpublished) examining benefits and harms for each intervention

Page 4-5; lines 21-25; 1-3

6b Explanation for choice of comparators

Page 5; line 3

Objectives

7 Specific objectives or hypotheses

Page 5; lines 17-24

Trial design

8 Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group), allocation ratio, and framework (eg, superiority, equivalence, non-inferiority, exploratory)

Page 5; lines 4-5

Methods: Participants, interventions, and outcomes

Study setting

9 Description of study settings (eg, community clinic, academic hospital) and list of countries where data will be collected. Reference to where list of study sites can be obtained

Page 6; 2-5

Eligibility criteria

10 Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and individuals who will perform the interventions (eg, surgeons, psychotherapists)

Page 6; lines 10-23

Interventions

11a Interventions for each group with sufficient detail to allow replication, including how and when they will be administered

Pages 7-8; lines 19-25;
Lines 2-23

11b Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose change in response to harms, participant request, or improving/worsening disease)

n.a.

11c Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence (eg, drug tablet return, laboratory tests)

n.a.

11d Relevant concomitant care and interventions that are permitted or prohibited during the trial

n.a.

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4	Outcomes	12	Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended	Page 10-11; lines 1-24 Lines 1-19
5				
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9	Participant timeline	13	Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure)	Page 29; line 2 (Figure 1)
10				
11				
12	Sample size	14	Estimated number of participants needed to achieve study objectives and how it was determined, including clinical and statistical assumptions supporting any sample size calculations	Page 12; lines 19-24
13				
14				
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16				
17	Recruitment	15	Strategies for achieving adequate participant enrolment to reach target sample size	Page 6; lines 4-5
18				
19	Methods: Assignment of interventions (for controlled trials)			
20				
21	Allocation:			
22				
23	Sequence generation	16a	Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any factors for stratification. To reduce predictability of a random sequence, details of any planned restriction (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants or assign interventions	Page 7; lines 6-8
24				
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29	Allocation concealment mechanism	16b	Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned	Page 7; lines 8-10
30				
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33	Implementation	16c	Who will generate the allocation sequence, who will enrol participants, and who will assign participants to interventions	Page 7; lines 8-10
34				
35				
36	Blinding (masking)	17a	Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome assessors, data analysts), and how	Page 7; lines 11-15
37				
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17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's allocated intervention during the trial n.a.

Methods: Data collection, management, and analysis

Data collection methods 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known. Reference to where data collection forms can be found, if not in the protocol Page 9-10; lines 16-14
Lines 1-18

18b Plans to promote participant retention and complete follow-up, including list of any outcome data to be collected for participants who discontinue or deviate from intervention protocols n.a.

Data management 19 Plans for data entry, coding, security, and storage, including any related processes to promote data quality (eg, double data entry; range checks for data values). Reference to where details of data management procedures can be found, if not in the protocol Page 14; lines 13-22

Statistical methods 20a Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the statistical analysis plan can be found, if not in the protocol Page 13; lines 2-17

20b Methods for any additional analyses (eg, subgroup and adjusted analyses) Page 13; line 13

20c Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation) n.a.

Methods: Monitoring

Data monitoring 21a Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed Page 14; lines 2-3

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4		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	n.a.
5				
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7	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse events and other unintended effects of trial interventions or trial conduct	Page 13; lines 20-25
8				
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10	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	n.a.
11				
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14	Ethics and dissemination			
15				
16	Research ethics approval	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	Page 15; lines 10-16
17				
18				
19	Protocol amendments	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals, regulators)	Page 15; lines 10-16
20				
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23	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and how (see Item 32)	Page 7; line 12
24				
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26		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary studies, if applicable	n.a.
27				
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29	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained in order to protect confidentiality before, during, and after the trial	Page 14; lines 13-22
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33	Declaration of interests	28	Financial and other competing interests for principal investigators for the overall trial and each study site	Page 15; lines 10-16 Page 18; line 7
34				
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36	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that limit such access for investigators	Page 18; lines 10-17
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Ancillary and post-trial care	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial participation	Page 13; line 23
Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals, the public, and other relevant groups (eg, via publication, reporting in results databases, or other data sharing arrangements), including any publication restrictions	Page 14; line 10
	31b	Authorship eligibility guidelines and any intended use of professional writers	Page 18; lines 2-4
	31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	Page 18; lines 10-17
Appendices			
Informed consent materials	32	Model consent form and other related documentation given to participants and authorised surrogates	Page 35
Biological specimens	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular analysis in the current trial and for future use in ancillary studies, if applicable	n.a.

*It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items. Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons "[Attribution-NonCommercial-NoDerivs 3.0 Unported](https://creativecommons.org/licenses/by-nc-nd/3.0/)" license.