BMJ Open  Vitamin D levels in critically ill patients with acute kidney injury: a protocol for a prospective cohort study (VID-AKI)

Lynda Katherine Cameron,1,2,3 Katie Lei,3 Samantha Smith,3 Nanci Leigh Doyle,4 James F Doyle,4 Kate Flynn,3 Nicola Purchase,3 John Smith,3 Kathryn Chan,3 Farida Kamara,3 Nardos Ghebremedhin Kidane,3 Lui G Forni,4 Dominic Harrington,5 Geeta Hampson,6 Marlies Ostermann3,7

ABSTRACT

Introduction  Acute kidney injury (AKI) affects more than 50% of critically ill patients. The formation of calcitriol, the active vitamin D metabolite, from the main inactive circulating form, 25-hydroxyvitamin D (25(OH)D), occurs primarily in the proximal renal tubules. This results in a theoretical basis for reduction in levels of calcitriol over the course of an AKI. Vitamin D deficiency is highly prevalent in critically ill adults, and has been associated with increased rates of sepsis, longer hospital stays and increased mortality. The primary objective of this study is to perform serial measurements of 25(OH)D and calcitriol (1,25(OH)2D), as well as parathyroid hormone (PTH) and fibroblast growth factor 23 (FGF23) levels, in critically ill adult patients with and without AKI, and to determine whether patients with AKI have significantly lower vitamin D metabolite concentrations. The secondary objectives are to describe dynamic changes in vitamin D metabolites, PTH and FGF23 during critical illness; to compare vitamin D metabolite concentrations in patients with AKI with and without renal replacement therapy; and to investigate whether there is an association between vitamin D status and outcomes.

Methods and analysis  230 general adult intensive care patients will be recruited. The AKI arm will include 115 critically ill patients with AKI Kidney Disease Improving Global Outcome stage II or stage III. The comparison group will include 115 patients who require cardiovascular or respiratory support, but who do not have AKI. Serial measurements of vitamin D metabolites and associated hormones will be taken on prespecified days. Patients will be recruited from two large teaching Trusts in England. Data will be analysed using standard statistical methods.

Ethics and dissemination  Ethical approval was obtained. Upon completion, the study team will submit the study report for publication in a peer-reviewed scientific journal and for conference presentation.

Trial registration number  NCT02869919; Pre-results.

INTRODUCTION

Vitamin D deficiency is common, affecting 25% of the general UK adult population in summer and up to 30%–40% in winter months.1 There is increasing awareness that vitamin D deficiency is associated with a number of important health consequences. The role of vitamin D metabolites in calcium homeostasis and the regulation of bone metabolism is well characterised. In the last decade it has been recognised that vitamin D status also impacts on infectious, immunological, neurological, cardiovascular, endothelial and respiratory disorders.2–5 25-Hydroxyvitamin D, termed 25(OH)D, is the main circulating metabolite of vitamin D and serves as a marker for evaluation of vitamin D status.1 Generally, although not universally, accepted cut-offs for defining vitamin D deficiency and severe deficiency are 25(OH)D levels of 50 nmol/L and 30 nmol/L, respectively.3,5

Intensive care units (ICUs) worldwide have reported vitamin D deficiency rates ranging from 60% to 100%.5 The causes may predate admission, or be a consequence of critical illness, interventions or therapies.5 25(OH) D concentrations of <50 nmol/L are associated with an increased rate of sepsis, a longer length of hospital stay and increased mortality both in-hospital and at 30 days in...
Patients with AKI may have altered vitamin D metabolism mechanisms for reasons that could include the following:

1. The kidneys generate the majority of 1α-hydroxylase CYP27B1 and other hydroxylases implicated in vitamin D metabolism, and are responsible for producing and regulating 1,25(OH)_2D and vitamin D-dependent proteins.  

2. FGF23 reduces circulating 1,25(OH)_2D levels. Concentrations of FGF23 are elevated in AKI due to increased production by bone and reduced clearance. 

3. Patients with AKI have lower VDBP concentrations than patients without AKI. 

4. Pre-existing vitamin D deficiency is highly prevalent in the general population and has been associated with the development of AKI during critical illness. 

5. 1,25(OH)_2D has an elimination half-life of only 4–15 hours. Circulating levels may fall over the first days of critical illness if renal or extrarenal production capability is compromised.

6. Previous studies examining the association between AKI and vitamin D deficiency have been inconclusive. A study including 200 critically ill patients revealed significantly lower 1,25(OH)_2D concentrations at the time of AKI diagnosis compared with controls without AKI, but there were no differences in 25(OH)_2D concentrations. PTH and FGF23 concentrations were not measured and serial measurements were not undertaken. 

A recent meta-analysis highlighted the lack of relevant studies that consider serial measurements of 25(OH)_2D and 1,25(OH)_2D alongside other markers of vitamin D metabolism. Others have emphasised the importance of conducting intervention studies in the most appropriate cohort. Our aim is to understand the dynamic changes in vitamin D metabolite concentrations that occur over the course of AKI, from which the design of a future intervention study could be informed.

OBJECTIVES

The primary objective is to perform serial measurements of 25(OH)_2D and 1,25(OH)_2D, PTH, FGF23 and VDBP concentrations in critically ill adult patients with and without AKI, to determine whether patients with AKI have significantly lower vitamin D metabolite concentrations. The following are the secondary objectives:

1. to describe the changes of 25(OH)_2D, 1,25(OH)_2D, PTH and FGF23 levels during critical illness

2. to compare total and bioavailable (ie, free plus albumin-bound, as opposed to that bound to VDBP; online supplementary appendix 1) concentrations of 25(OH)_2D and 1,25(OH)_2D in patients with AKI treated with and without RRT

3. to investigate whether there is an association between total and bioavailable vitamin D metabolite concentrations and outcome in critically ill patients
with and without AKI; outcomes reported will be hospital mortality, 30-day mortality and length of stay in ICU and in hospital
4. to explore the possible statistical interaction between biomarker levels and length of ICU stay.

**METHODS AND ANALYSIS**

**Setting**
The study will be conducted in critical care units in two National Health Service hospitals in the UK.

**Patient selection**
Adult patients admitted to a critical care unit will be screened for the presence of AKI, on the basis of either serum creatinine or urine output, as defined by the Kidney Disease Improving Global Outcome (KDIGO) consensus criteria (table 1). The critical care units are mixed medical and surgical.

The following are the criteria for inclusion into the study: (1) adult patient (18 years or older) admitted to a critical care unit and (2) presence of KDIGO AKI II or III for ≤36 hours (AKI arm), or presence of cardiovascular and/or respiratory failure requiring the use of invasive or non-invasive respiratory support and/or treatment with catecholamines (non-AKI arm), with a requirement anticipated to last for longer than 24 hours.

The following are the exclusion criteria: (1) AKI stage I (as defined by KDIGO criteria), (2) known vitamin D deficiency (recorded value of <50 nmol/L on the hospital’s electronic record system, diagnosis documented on a hospital clinic letter, or evidence of vitamin D supplementation prescribed in primary or secondary care in the last 3 months), (3) known vitamin D supplementation in the last 3 months (either as a single agent or in a combination product, including multivitamins), (4) pre-existing chronic kidney disease (CKD) stages 3b–5 (ie, estimated glomerular filtration rate <45 mL/min/1.73 m²), (5) renal transplant, (6) known hyperparathyroidism, (7) need for total parenteral nutrition (TPN), (8) anticipated life expectancy <48 hours, (9) haemoglobin concentration <70g/L (unless a decision has been made to administer blood transfusion), and (10) pregnancy.

**Sample size calculation**
Based on data from Lai et al, 126 patients with complete data are required to detect a difference of 25 pmol/L in 1,25(OH)₂D concentrations between the two arms, with a power of 80% and at a two-tailed significance level of 5% (assuming an SD of 50 in each group). Interim review of the case report forms of a small set of patients highlighted an attrition rate of 40%–50% between the time of eligibility screening and day 5, leading to the requirement to recruit a total of 230 patients to analyse at least 126 with complete data.

**Baseline data collection**
The following data will be collected: age, gender, ethnicity, weight, height, ideal body weight, place of residence prior to hospitalisation, admission diagnosis, comorbidities (CKD, chronic lung disease, chronic heart failure, coronary artery disease, cancer, chronic liver disease, diabetes, chronic gastrointestinal disease), premorbid serum creatinine concentration, C reactive protein (CRP), albumin, Acute Physiology and Chronic Health Evaluation (APACHE) II score on admission, and daily Sequential Organ Failure Assessment (SOFA) scores. We also plan to collect data on potential confounding factors, including daily nutritional intake, fluid balance, calcium and phosphate supplementation and treatment with extracorporeal therapy (ie, RRT and plasma exchange).

**Sampling**
On the day of enrolment, day 5 and the day of discharge from the critical care unit, blood will be taken for measurement of 25(OH)D, 1,25(OH)₂D, PTH, FGF23 and VDBP. Samples will be taken in the morning, between 09:00 and 12:00. In a yellow top SST vacutainer, 5.5 mL of blood will be collected, and 4 mL will be collected in a purple top EDTA vacutainer. Samples will be centrifuged at 3500 rpm for 10 min, then aliquots of 0.5 mL serum for 25(OH)D, 1 mL serum for 1,25(OH)₂D and 0.5 mL serum for VDBP will be taken from the SST tube, and 1 mL of plasma for PTH and 1 mL of plasma for FGF23 from the EDTA tube. Samples will then be stored at −70°C until analysis. Additional measurements of 1,25(OH)₂D and PTH will be made on day 2 (table 2). Calcium and phosphate are measured routinely on a daily basis for all ICU patients.
Table 2  Sampling schedule

<table>
<thead>
<tr>
<th>Serum concentration</th>
<th>Day 0 (day of enrolment)</th>
<th>Day 2</th>
<th>Day 5</th>
<th>Day of intensive care unit discharge</th>
</tr>
</thead>
<tbody>
<tr>
<td>25-hydroxyvitamin D</td>
<td>√</td>
<td>–</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>1,25-dihydroxyvitamin D</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Calcium (Ca)</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Phosphate (P)</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Parathyroid hormone</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Fibroblast growth factor 23</td>
<td>√</td>
<td>–</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Vitamin D binding protein</td>
<td>√</td>
<td>–</td>
<td>√</td>
<td>√</td>
</tr>
</tbody>
</table>

patients as part of preward round bloods, typically taken at 06:00. These results will be recorded.

All samples will be stored in dedicated research freezers until batch analysis.

25(OH)D will be measured by liquid chromatography-mass spectrometry, with a coefficient of variation of 7.3% for 25(OH)D2 and of 10.9% for 25(OH)D3. Total 1,25(OH)2D will be measured by radioimmunoassay. FGF23 will be measured using the Immutopics Human FGF23 (C-Term) ELISA assay.21 VDBP will be measured by ELISA assay, using the Quantikine Human Vitamin D BP Immunoassay.22 PTH will be measured using a Roche automated analyser.

Statistical analysis

Multiple comparisons will be performed to determine the difference in vitamin D metabolite concentrations between the two arms. The distribution of the data will be examined and appropriate statistical tests will be applied, accounting for potential confounding factors including age, ethnicity, CRP, calcium concentrations, season, severity of illness at admission using APACHE II scores and pattern of organ failures using SOFA scores. A longitudinal analysis will be presented, covering the time from enrolment to the date of critical care discharge, to describe changes in 25(OH)D, 1,25(OH)2D, FGF23 and PTH concentrations over the course of critical illness. An investigation of possible correlation between vitamin D metabolites and related hormones (PTH and FGF23) on day 0 and day 5 is planned. A subgroup analysis is planned for those participants receiving RRT and in patients with sepsis on the day of enrolment. An interim analysis is planned after recruitment of 126 participants.

DISCUSSION

This study aims to describe the dynamic changes in 25(OH)D and 1,25(OH)2D concentrations, and associated hormones, in a general population of critically unwell patients with and without AKI.

The strengths of this project are that (1) we plan to measure 25(OH)D and 1,25(OH)2D together with regulatory hormones and proteins, which will address an existing gap in knowledge. (2) Serial measurements will be undertaken to determine dynamic changes, including an additional measurement of 1,25(OH)2D on day 2. This will serve to provide data on the timing and magnitude of a drop in concentrations of this metabolite, which is predicted based on previous work23 and from the biological basis that 1,25(OH)2D has a circulating half-life of only 4–15 hours.1 (3) The project is adequately powered to provide meaningful data.

We are aware of potential limitations, which we will eliminate as much as possible.

1. The high attrition rate identified in early work comprises participants who did not have AKI at enrolment, but who went on to develop AKI during the first 5 days of their critical care stay; patients who went on to meet an exclusion criterion such as requirement for TPN; and patients for whom a pre-existing known vitamin D deficiency was identified after enrolment. The sample size was increased to 230 to maintain power to detect a clinically significant difference between arms if one exists. The increase in sample size was approved by a national research ethics committee and the institutional governance department.

2. We plan to perform serial measurements but will only undertake blood sampling once a day. Blood samples will be collected in the morning (between 09:00 and 12:00) on the days specified in table 2. Previous studies confirmed diurnal variation of 25(OH)D concentrations in intensive care patients over 24 hours with mean within-patient variation of 24.3 nmol/L over a 24-hour period.24 As such, single point in time measures may not be adequate to describe vitamin D status fully.

3. We plan to enrol patients with AKI independent of underlying aetiology. By not stratifying patients by aetiology of AKI, patients with a wide range of clinical diagnoses will be eligible for inclusion and the results of the study will remain generalisable. Stratification would, nonetheless, have been useful.
to identify a population who may benefit most from supplementation in a future trial.

4. Patients prescribed vitamin D supplements in primary care or from secondary or tertiary care clinics at the host institutions can be identified and excluded from the study. However, patients taking vitamin D prescribed by other institutions or those buying over-the-counter supplements cannot be identified in all cases. Medication histories do routinely prompt for this, acknowledging that information from relatives in cases where the patient cannot clarify for themselves may be incomplete.

5. Limited resources mean measurement of other associated proteins, including alpha Klotho, which is primarily generated by the kidneys and plays a role in bone mineral metabolism, is not possible. Further, the lack of a standardised assay to measure circulating Klotho precludes its measurement within this study. It could, however, represent an interesting area for future work.

6. The equations used to describe levels of bioavailable vitamin D metabolites (ie, free and albumin-bound; online supplementary appendix 1) have not been validated in the critical care population, but have been used in several other studies that included critical care patients. The bioavailable vitamin D values reported in this study should be viewed as indicative estimates, as direct assay of free vitamin D metabolites is not possible. While measurement of concentration of VDBP to inform the equations in online supplementary appendix 1 is possible within the available resources, assessment of genotype is not. Lack of this information, which is known to affect the affinity of vitamin D metabolites for VDBP, is a limitation of this work.

Contributors LKC is a co-investigator of the study and was involved in the design of the study protocol. She wrote the first draft of the manuscript, designed the case report form and is the primary data collector at the lead site. She revised the manuscript in response to suggestions by all authors and the reviewers, and approved and submitted the final draft. KL, SS, NLD, KF, NP, JS, KC, FK and NGK are research coordinators and have a crucial role in the conduct of the study in all participating sites. They were involved in the design of the protocol, revised the manuscript and approved the final draft. JFD and LGF are the lead investigators in one of the participating sites. They made contributions to the study protocol, revised the manuscript and approved the final draft. DH advised on the laboratory measurements of vitamin D and related substances. He contributed to the study protocol and the first draft of the manuscript, revised subsequent drafts and approved the final version. GH is a consultant in clinical chemistry and advised on the laboratory measurements of vitamin D and related hormones. She contributed to the study protocol and the first draft of the manuscript, revised subsequent drafts and approved the final version. MO is the chief investigator of the study. She developed the study and wrote the main study protocol. She is overseeing the project and revised several drafts of the manuscript before approving the final version. All authors approved the final version and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Funding This work was supported by research funding from Fresenius Medical Care. The sponsor was not involved in the design of the protocol and has no involvement in the conduct of the study. The results will remain intellectual property of the research team. LKC’s involvement was supported by the Biomedical Research Centre at Guy’s and St Thomas’ NHS Foundation Trust.

Competing interests None declared.

Patient consent Obtained.

Ethics approval London — Camberwell St Giles Research Ethics Committee.

Provenance and peer review Not commissioned; externally peer reviewed.

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ETHICS AND DISSEMINATION

Full ethical approval was granted by the London — Camberwell St Giles National Research Ethics Committee on 30 March 2016.

Participants will be asked to give their consent to participate in the study. In the event that they are unable to give consent for themselves, a personal consultee (eg, next of kin) will be contacted and informed about the study and asked whether the patient would have any objections to participating in this observational study. They will be asked to sign a personal consultee declaration form. In case a personal consultee is not available, a nominated consultee (a clinician who is not connected to the research study) will be contacted. When the patient regains capacity, they will be informed about the study and asked to give consent to remain in the study. The Research Ethics Committee agreed that the study has the potential to benefit participants lacking capacity without imposing a disproportionate burden on them.

On completion, the study team will submit the study report for publication in a peer-reviewed scientific journal.

REFERENCES


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*BMJ Open* 2017 7:
doi: 10.1136/bmjopen-2017-016486

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